

PulseNet Standard Operating Procedure for PulseNet MLVA of *Salmonella enterica* Serotype Enteritidis – Applied Biosystems Genetic Analyzer 3500 Platform

1. PURPOSE:

To describe the standardized laboratory protocol for molecular subtyping of *Salmonella enterica* serotype Enteritidis.

2. SCOPE

To provide PulseNet participants with a single protocol for performing MLVA of *S. enterica* serotype Enteritidis, thus ensuring inter-laboratory comparability of the generated results using the Applied Biosystems Genetic Analyzer 3500 platform.

3. DEFINITIONS:

- 3.1. **MLVA:** Multiple-locus variable-number tandem repeat analysis
- 3.2. **VNTR:** Variable-number tandem repeat
- 3.3. **DNA:** Deoxyribonucleic acid
- 3.4. **DNase:** Deoxyribonuclease
- 3.5. **PCR:** Polymerase chain reaction
- 3.6. **HPLC:** High purity liquid chromatography
- 3.7. **dNTP:** Deoxyribonucleotide triphosphate
- 3.8. **CDC:** Centers for Disease Control and Prevention
- 3.9. **SOP:** Standard Operating Procedure

4. RESPONSIBILITIES/PROCEDURE

4.1. Biosafety Warning

4.1.1. *S. enterica* serotype Enteritidis is a human pathogen causing serious disease. Always use a minimum of Biosafety level 2 practices and extreme caution when transferring and handling strains of this serotype. Work in a biological safety cabinet when handling large amounts of cells. Disinfect or dispose of all plastic ware and glassware that come in contact with the cultures in a safe manner.

4.2. Reagents, supplies, and equipment needed for DNA template preparation

- 4.2.1. Trypticase soy agar with 5 % sheep blood (TSA-SB) or comparable media
- 4.2.2.1 µl and 10 µl inoculation loops
- 4.2.3. 0.5 ml microcentrifuge tubes
- 4.2.4. DNase-free, molecular biology -grade water
- 4.2.5. Vortex
- 4.2.6. Boiling water bath or thermal block / thermocycler accommodating 0.5 ml tubes
- 4.2.7. Tabletop centrifuge for high rpm (up to 13,000-14,000 rpm) spinning
- 4.2.8. Pipets (200 µl) for aliquoting 100 µl of DNase-free, molecular biology-grade water

4.2.9. Filtered sterile pipet tips

4.3. Reagents, supplies, and equipment needed for PCR

- 4.3.1. DNA lysates from isolates (store at -20°C or -80°C freezer for long term)
- 4.3.2. PCR primers (see appendix PNL30-1)
 - 4.3.2.1. Fluorescent-labeled forward primers
 - 4.3.2.1.1. HPLC-purified
 - 4.3.2.2. Unlabeled reverse primers
 - 4.3.2.2.1. Regular gel filtration purification
 - 4.3.2.3. Biosearch Technologies (Novato, CA; www.biosearchtech.com; 1-800-436-6631) sells primers labeled with the three dyes needed for the protocol
 - 4.3.2.4. Divide the concentrated stocks in portions and store at -80°C freezer
 - 4.3.2.4.1. One vial should contain enough to prepare 25-50 µl of working solution. Avoid repetitive freeze-thaw cycles of concentrated primer stocks.
 - 4.3.2.5. The 2.5 µM, and 12.5 µM working solutions can be stored at either -20°C or -80°C freezer
 - 4.3.2.6. Prepare new working solutions every month or if a significant drop in the fluorescence level is observed (for instructions refer to PNQ06_MLVA ABI certification, appendix PNQ06-5)
- 4.3.3. 96-well polypropylene PCR plates (Fisher, Cat. No. 07-200-613) or Microamp PCR tubes without caps (Life Technologies, Cat. No. N8010533)
- 4.3.4. 8-well strip caps for polypropylene plate (Fisher, Cat. No. 07-200-639) or MicroAmp strip caps for individual tubes (Life Technologies, Cat. No. N8010535)
- 4.3.5. DNase-free, molecular biology -grade water
- 4.3.6. 1.5 ml Eppendorf microcentrifuge tubes
- 4.3.7. PCR Nucleotide mix (ready-to-use dNTP mix containing all four nucleotides; Roche, Cat. No. 11 814 362 001)
- 4.3.8. Platinum Taq Polymerase with 50 mM MgCl₂ and 10X buffer (Life Technologies Cat. No. 10966-034)
- 4.3.9. PCR cooling block (VWR International, Cat. No. 62111-762)
- 4.3.10. DNA Engine (Bio-Rad), GeneAmp (Life Technologies) or similar thermocycler with a heated lid option and 96-well block format
- 4.3.11. Parafilm M, 4" width (VWR, Cat. No. 52858-032)
- 4.3.12. Complete set (1000 µl, 200 µl, 100 µl, 20 µl, 10 µl, and 2 µl) of single channel pipettors for mastermix set-up ("clean set")
- 4.3.13. A 1-10 µl single channel pipettor for adding DNA templates
- 4.3.14. Filtered tips for pipettors
- 4.3.15. Microfuge for low (up to 6,000 rpm) rpm spinning

4.4. Reagents, supplies and equipment needed for Genetic Analyzer 3500

- 4.4.1. DNase-free, molecular biology -grade water
- 4.4.2. PCR cooling block (VWR International, Cat. No. 62111-762)
- 4.4.3. 10 µl, 100 µl, and 1000 µl single channel pipettors
- 4.4.4. 1-10 µl and 20-200 µl multichannel pipettors
- 4.4.5. Filtered pipette tips
- 4.4.6. Sterile solution basins
- 4.4.7. 1.5 ml Eppendorf microcentrifuge tubes
- 4.4.8. 96-well polypropylene (non-PCR) V-bottom plate (for dilutions; Fisher Scientific, Cat. No. 07-200-698)
- 4.4.9. MicroAmp Optical 96-well reaction plates (Life Technologies, Cat. No. 4306737)

- 4.4.10. 96-well plate base (Life Technologies, Cat. No. 4317237)
- 4.4.11. Rubber septa for 96-well reaction plates (Life Technologies, Cat. No. 4315933)
- 4.4.12. 96-well plate retainer (Life Technologies, Cat. No. 4317241)
- 4.4.13. Hi-Di Formamide, 25 ml (Life Technologies, Cat. No. 4311320)
- 4.4.14. GeneFlo 625 DNA size standard – ROX, 800 µl (Chimerx, Cat. No. 3125-02)
- 4.4.15. Multi-Capillary DS-30 (Dye Set D) Matrix Standard Kit (Life Technologies, Cat. No. 4345827)
 - 4.4.15.1. Needed periodically to establish the system dye color spectra for the instrument. Required when analyzing fragments labeled with FAM, HEX, NED, and ROX.
 - 4.4.15.2. *NOTE: DS-33 Matrix Standard (Dye set G5, required for analyzing fragments labeled with FAM, VIC, NED, PET, and LIZ) is typically installed as default as part of the instrument installation process*
 - 4.4.15.3. In order to install the DS-30 Matrix, follow the instructions of the kit insert and the “Getting Started Guide”, chapter “Performing a Spectral Calibration”
- 4.4.16. 3500 24 Capillary Array 50 cm (Life Technologies, Cat. No. 4404689)
- 4.4.17. Genetic Analyzer Cathode Buffer Container (4 pack), 1X (Life Technologies, Cat. No. 4408256)
- 4.4.18. Genetic Analyzer Anode Buffer Container (4 pack), 1X (Life Technologies, Cat. No. 4393927)
- 4.4.19. 3500 Conditioning Reagent (Life Technologies, Cat. No. 4393718)
- 4.4.20. 3500 POP7 Performance Optimized Polymer, (384 samples) (Life Technologies, Cat. No. 4393708)
- 4.4.21. Parafilm M, 4” width (VWR, Cat. No. 52858-032)
- 4.4.22. Centrifuge with a microtiter plate rotor
- 4.4.23. Heating block or thermal cycler accommodating a 96-plate for denaturation
- 4.4.24. Microfuge for low (up to 6,000 rpm) rpm spinning

4.5. Isolate preparation

4.5.1. DAY 0

- 4.5.1.1. Streak an isolated colony from pure test cultures to TSA-SB plate (or comparable media). Incubate cultures at 37°C for 14-18 hrs.

4.5.2. DAY 1

- 4.5.2.1. For each isolate to be typed, aliquot 100 µl of sterile, molecular biology-grade water into 0.5 ml microfuge tubes. Use a sterile, disposable 1 µl loop to pick 2-3 colonies (about half of a loop-full); rotate the loop in the microfuge tube to release the bacteria into the water. Cap and vortex for 10-15 seconds to disperse any clumps.
- 4.5.2.2. Place the tubes in a 100°C water bath or heat block for 10 minutes. Cool briefly on ice or in fridge and centrifuge for 10 minutes at 10,000 rpm. Place on ice or in fridge while preparing PCR reactions. These DNA templates can be stored at -20°C or -80°C for several years.

4.6. PCR procedure

4.6.1. DAY 1

- 4.6.1.1. Fill out, save with the run name, and print a PCR template worksheet (see appendix PNL30-2) with appropriately labeled samples (a maximum of 46 isolates/plate; 44 unknowns + two positive controls and a negative control; two wells are reserved for the internal ladder).
 - 4.6.1.1.1. For each isolate, two wells must be labeled as follows: BNkeyR1 where “BNkey” represents the isolate-specific state laboratory identification number (be sure to use the exactly same isolate ID that is used in the PFGE gels uploaded to the

national database) and “R1” represents one of the two specific multiplex PCR reactions (R1, R2).

- 4.6.1.2. Fill out, and print a PCR mastermix calculation worksheet (see appendix PNL30-3) by typing the number of isolates to be tested (plus 2-3 extra) in the PCR mastermix calculators labeled R1 and R2. This number is highlighted in red and is next to “number of samples to be analyzed”. The mastermixes for reactions 1 and 2 (R1, R2) for one sample are as follows:

<i>R1</i>	<i>Volume (μl)</i>	<i>Final conc</i>
PCR water	4.91	
PCR buffer (10x)	1.00	1x
MgCl ₂ (50 mM)	0.40	2.0 mM
dNTPs (10 mM)	0.20	0.20 mM
SE1-F (2.5 μM)	0.20	0.05 μM
SE1-R (2.5 μM)	0.20	0.05 μM
SE2-F (12.5 μM)	0.32	0.40 μM
SE2-R (12.5 μM)	0.32	0.40 μM
SE8-F (2.5 μM)	0.28	0.07 μM
SE8-R (2.5 μM)	0.28	0.07 μM
SE6-F (12.5 μM)	0.34	0.43 μM
SE6-R (12.5 μM)	0.34	0.43 μM
Taq (5 U/μl)	0.20	1.00 U
Total volume	9.00	

<i>R2</i>	<i>Volume (μl)</i>	<i>Final conc</i>
PCR water	5.52	
PCR buffer (10x)	1.00	1x
MgCl ₂ (50 mM)	0.40	2.00 mM
dNTPs (10 mM)	0.20	0.20 mM
SE5-F (2.5 μM)	0.20	0.05 μM
SE5-R (2.5 μM)	0.20	0.05 μM
SE3-F (12.5 μM)	0.40	0.50 μM
SE3-R (12.5 μM)	0.40	0.50 μM
SE9-F (2.5 μM)	0.08	0.02 μM
SE9-R (2.5 μM)	0.08	0.02 μM
Taq (5 U/μl)	0.20	1.00 U
Total volume	9.00	

- 4.6.1.2.1. *NOTE: these primer concentrations serve as a starting point. Since laboratory-specific factors, such as the age of the primer stocks, calibration status of the thermocyclers and pipettes, etc. affect amplification efficiency, each laboratory will have to re-optimize the primer concentrations for optimal detection of all targets. However, any other parameters stated in the SOP should not be changed.*
- 4.6.1.3. Thaw all reagents and supplies needed for PCR reactions and place on ice; keep primers light protected as much as possible.
- 4.6.1.3.1. *NOTE: PCR mastermixes should be set up in a clean hood that is dedicated just for this purpose and where no cultures or DNA are handled.*
- 4.6.1.4. Prepare the two separate PCR mastermixes in 1.5 ml Eppendorf tubes following the instructions in the PCR mastermix calculation worksheet (see appendix PNL30-3) and place on ice. Add the mastermix components in the following order: water, 10x PCR buffer, Mg₂Cl, dNTPs, primers, and then finally Taq polymerase. Mix the reaction mixture by vortexing briefly.
- 4.6.1.4.1. Vortex all reagents except Taq polymerase before adding to the mastermix. Taq may be briefly centrifuged with low rpm, if necessary, to pull the enzyme down to the bottom of the tube.
- 4.6.1.5. Place a 96-well PCR plate or required number of PCR tubes in a PCR cooling block.
- 4.6.1.6. Dispense 9.0 μl of each mastermix into the appropriate columns of the 96-well polypropylene plate / PCR tubes as noted in the PCR template worksheet (see appendix PNL30-2)

- 4.6.1.7. Add 1 µl of PCR water to each of the two different wells representing the negative controls of the two reactions.
- 4.6.1.8. Add 1.0 µl of DNA lysate to each of the two different wells representing the two PCR reactions. Use *S. enterica* serotype Enteritidis strain K1891 (ATCC25928) as a positive control (it is recommended to run the positive control in duplicate). The internal ladder to be used will be comprised of pooled PCR products of the isolates H9560 and 2010K0017 (see appendix PNL30-5 for instructions for ladder preparation)
- 4.6.1.9. Cover all wells / tubes with 8-well strip caps and firmly clamp down to avoid any evaporation during PCR amplification.
- 4.6.1.10. Recommendation: briefly spin down the plate / tubes to remove any air bubbles.
- 4.6.1.11. Setup the following PCR parameters and save the program as “SEMLVA”:
 - Step 1 95°C for 5 minutes
 - Step 2 94°C for 20 seconds
 - Step 3 65°C for 20 seconds
 - Step 4 72°C for 20 seconds
 - Step 5 Go to Step 2, 34x
 - Step 6 72°C for 5 minutes
 - Step 7 Indefinite hold at 4°C
- 4.6.1.11.1. Make sure to use the heated lid option on the PCR block and “calculated” or tube control (instead of block or probe control) as a temperature control method.
- 4.6.1.12. When the PCR is complete, store the amplification products light-protected at 4°C until ready to run on the sequencer. If the fragment analysis is not performed the same day, the plate should be stored at -20°C or -80°C. The PCR products are stable for approximately one month, when stored frozen.

4.7. Initial setup of Genetic Analyzer 3500 instrument

NOTE: Steps 4.7 only needs to be performed before the very first run and every time a new working database is created.

- 4.7.1. Click on the “3500” icon to open the 3500 Data Collection Software. The software will open a window and the main screen will display the “Dashboard”. If the connections are functioning properly, the status light on the sequencer will be green, the “State” of the instrument will be “Idle”, and all of the consumables should have the number of days/samples remaining for each item.
- 4.7.2. Setting up the Size Standard
 - 4.7.2.1. Click on the “Library” icon at the top right of the screen.
 - 4.7.2.2. At the left-hand side of the screen select the “Size Standards” menu option.
 - 4.7.2.3. At the top of this screen select “Create” in the toolbar
 - 4.7.2.4. A new window will appear and in the box next to “Size Standard” enter in “GeneFlo625”
 - 4.7.2.5. From the “Dye Color” drop-down menu select “Red”
 - 4.7.2.6. In the “Enter new Size Standard definition” box enter the fragment sizes for each peak in the ladder (refer to the GeneFlo 625 product insert for the peak sizes). Separate each size using a comma.
 - 4.7.2.7. When finished, click the “Add Size(s)” button, then click “Save” and “Close”.
- 4.7.3. Setting up Run Parameters:
 - 4.7.3.1. Once the fragment sizes have been assigned the Assays, File Name Conventions, and Results Group will need to be set up.
 - 4.7.3.1.1. Assays

- 4.7.3.1.1.1. At the left-hand side of the screen select the “Assays” menu option and click “Create” in the toolbar at the top of the screen
- 4.7.3.1.1.2. For “Assay Name” enter “SE_Fragtest” and from the “Application Type” drop-down menu select “Fragment”. Under the “Protocols” heading make sure “No” is selected for assigning multiple instrument protocols.
- 4.7.3.1.1.3. Next to “Instrument Protocol” select the “Create New” button. A new window will appear.
 - 4.7.3.1.1.3.1. From the “Dye Set” drop-down menu select “DS-30”
 - 4.7.3.1.1.3.2. For “Run Module” select “FragmentAnalysis50_POP7xl”
 - 4.7.3.1.1.3.3. An alert window will pop up asking if you would wish to assign a protocol name. Click “Yes” and next to “Protocol Name” replace the ‘_1’ with ‘_2’ (i.e. - FragmentAnalysis50_POP7xl_2). Use the default values for all other parameters. This will create a new protocol with the specified criterion.
 - 4.7.3.1.1.3.3.1. *NOTE: These are the instrument default running conditions for Fragment Analysis with a 50 cm capillary array and POP7 polymer. You can check the running conditions by clicking the “Library” button at the top right of the dashboard and then selecting the “Assays” menu option on the left-hand side of the screen. The default conditions are:*
 - Oven_Temperature: 60°C
 - PreRun_Voltage: 15.0 kVolts
 - Pre-Run-Time: 180 sec.
 - Injection_Voltage: 1.6 kVolts
 - Injection_Time: 15 sec.
 - Data_Delay_Time: 1 sec.
 - Run_Voltage: 19.5 kVolts
 - Run time: 1330 sec.
 - 4.7.3.1.1.3.4. When finished, click on “Save to Library” and click “Yes” in the pop-up window that asks to apply the changes to the assay. Click “OK” to confirm, and then close this window.
- 4.7.3.1.1.4. Next to the “Sizcalling Protocol” option select the “Create New” button.
 - 4.7.3.1.1.4.1. In the new window enter “SEMLVA” as the “Protocol Name”.
 - 4.7.3.1.1.4.2. From the “Size Standard” drop-down menu select “GeneFlo625”
 - 4.7.3.1.1.4.3. Input the following analysis settings:

- Analysis: Full Range
- Sizing: Partial Sizing
- Start Size: 50
- Stop Size 625
- Size Calling Method: Local Southern Method
- Primer Peak: Present
- Minimum Peak Height:
 - Blue: 500
 - Green: 500
 - Yellow: 250
 - Red: 20
 - Uncheck Purple and Orange
- Smoothing: none
- Baseline window: 51 pts
- Min. Peak Half Width: 2 pts
- Peak Window Size: 21 pts
- Polynomial Degree: 2
- Slope Threshold Peak Start: 0.0
- Slope Threshold Peak End: 0.0

4.7.3.1.1.4.4. Click on “Save to Library” and “Yes” to apply to the assay. Select “OK” to confirm and close this window when finished.

4.7.3.1.1.5. Once all protocols have been established for the new assay, click “Save” and close the window to return to the “Library” main screen.

4.7.3.1.2. **File Name Conventions**

4.7.3.1.2.1. Access the “File Name Conventions” menu on the left-hand side of the screen. In the toolbar at the top of the screen select “Create”.

4.7.3.1.2.2. In the window that appears enter “MLVA” next to “Name”.

4.7.3.1.2.3. Use the default settings and click “Save” and close the window.

4.7.3.1.3. **Results Groups**

4.7.3.1.3.1. Access the “Results Groups” menu on the left-hand side of the screen. In the toolbar at the top of the screen select “Create”.

4.7.3.1.3.2. In the window that appears, enter “SResults” as the name for this results group.

4.7.3.1.3.3. Under the “Delimiters” section heading click the “Add” button to insert a dash in the “Selected Attributes” box on the right-hand side of the window.

4.7.3.1.3.4. From the “Available Attributes” list, scroll down to “Plate Name”. Select it and click the “Add” button in between the two boxes to add it to the “Selected Attributes” list.

4.7.3.1.3.5. Uncheck the “Include an Instrument Run Name folder” and “Include an Injection folder” options, keeping all other default parameters.

4.7.3.1.3.6. Save this results group to the library and close the window.

4.8. Genetic Analyzer 3500 instrument preparation before each run

4.8.1. DAY 1

4.8.1.1. Make sure service console is open and all components are green.

NOTE: Make sure a capillary array is installed in the instrument. For installation, follow the instructions of the “Install Capillary Wizard”. You can find the wizards by expanding the 3500 subfolder by highlighting

“Manual Control”. The “Wizards” drop-down menu will appear on top of the window.

- 4.8.1.1.1. *NOTE: The RFID tags on the packaging of the consumables automatically log in the hours spent on the instrument. The run will not start with expired reagents with the exception of an expired polymer*
- 4.8.1.2. Go back to the dashboard screen by clicking the “Dashboard” button at the top left of the screen
- 4.8.1.3. Select the “Create New Plate” icon at the top of the screen. Name the run following the standardized PulseNet naming system: use the unique identifier code that was assigned to your laboratory by PulseNet for the first two to four letters of the file name. The next two spaces will indicate the year and the next four spaces will indicate the month and the date the run was performed. For example GA070426 is a run made at the GA Public Health Laboratory on April 26, 2007. If several runs are performed the same day, separate the file names by using sequential numbers, for example GA070426-1, GA070426-2.
- 4.8.1.4. From the “Plate Type” drop-down menu, select “Fragment”
- 4.8.1.5. Type in the Owner name or initials
- 4.8.1.6. Use the default settings (50cm array, POP-7 polymer) for the rest of the options and click on the “Assign Plate Contents” button at the bottom of the screen.
- 4.8.1.7. The plate view screen will appear
- 4.8.1.8. Here you can type in the sample IDs or import a sample plate file from a template (see Appendix PNL30-5 for instructions on importing plate txt files).
- 4.8.1.9. To assign the Assay to the plate, proceed as follows:
 - 4.8.1.9.1. Click on “Add from Library” in the “Assays” box at the bottom of the screen.
 - 4.8.1.9.2. In the new window select “SE_Fragtest” from the list of assays.
 - 4.8.1.9.3. Add this assay by clicking on the “Add to Plate” button, and then close the window.
- 4.8.1.10. Adding the File Name Conventions (MLVA) and Results Groups (SEresults) can be done in the same manner, by selecting the “Add from Library” link in the respective boxes.
- 4.8.1.11. Once all of the files have been added to the plate, select the entire sample ID set by highlighting the individual cells or by clicking on column/row headings.
- 4.8.1.12. Check the box next to each of the run parameters: Assay, File Name Convention, and Results Group.
- 4.8.1.13. When each of the run parameters have been successfully assigned a dot will appear in the center of the cells in which the parameters were applied.
 - 4.8.1.13.1. *NOTE: If a plate layout has been imported from a .txt file the Assay, File Name Convention and Results Group should automatically be applied to the plate and check-marked for all samples.*
 - 4.8.1.13.2. *NOTE: What identifiers are shown in the sample ID cells can be changed by selecting or deselecting options in the “Show In Wells” drop-down menu above the sample plate layout. Assays, file name conventions, and results groups can be color coded by double-clicking the black dot to the left of the file name or during initial setup.*
- 4.8.1.14. Save the plate by clicking the “Save Plate” button in the toolbar at the top of the screen and selecting “Save” from the drop-down menu.
- 4.8.1.15. Go back to the dashboard screen by clicking the “Dashboard” button in the top left corner of the screen and continue with fragment analysis preparation.

- 4.8.1.16. If needed, install the POP7 polymer in the instrument.
- 4.8.1.16.1. Click on the “Maintenance” button in the toolbar at the top right of the window, and then select the “Maintenance Wizards” menu option on the left-hand side.
- 4.8.1.16.2. Follow the instructions of the “Replenish Polymer” wizard.
- 4.8.1.17. If needed, also install new anode and cathode buffer containers
- 4.8.1.17.1. *NOTE: if an old polymer pouch (been on the instrument > 7 days) is switched to a new one follow the instructions of the “Wash Pump and Channels” wizard instead. A conditioning pouch will be required for this process.*
- 4.8.1.18. *NOTE: To update the system after new reagents have been installed, click the “Refresh” button under “Consumables Information” on the Dashboard.*

4.9. Fragment analysis sample preparation

4.9.1. DAY 1

- 4.9.1.1. Thaw the Hi-Di Formamide, the GeneFlo 625 DNA size standard and the internal ladder (see appendix PNL30-4) and place on ice.
- 4.9.1.1.1. *NOTE: aliquot Hi-Di Formamide (500 µl / tube) and the size standard (50 µl / tube) in order to avoid frequent freeze-thaw cycles.*
- 4.9.1.2. Prepare a 96-well V-bottom plate for diluting the PCR reactions. Using a 200 µl multichannel pipettor and a solution basin, dispense 19 µl of molecular-grade water in the required number of wells.
- 4.9.1.3. Remove the plate / tubes with the PCR reactions from the thermocycler. Briefly spin down the plate / tubes, if necessary. Use a 10 µl multichannel pipettor to transfer 1 µl of each PCR reaction directly across to the corresponding set of wells in the dilution plate. In order to avoid cross-contamination, remove the strip cap from just one column at a time and recap the column before opening the next one.
- 4.9.1.4. For the internal ladder, combine R1 and R2 PCR products from the four PCR reactions of both internal ladder isolates to end up with a total of 40 µl. Mix well by pipetting up and down a few times and add 3 µl of internal ladder in two wells.
- 4.9.1.5. Using a 200 µl multichannel pipettor, mix the dilutions by pipetting up and down a few times. Cover the plate with parafilm and put in the fridge or on ice.
- 4.9.1.6. Prepare a fragment analysis master mix containing DNA size standard and Hi-Di Formamide for the samples following the calculations indicated in the table below. The fragment analysis mastermix calculations can also be performed using the autocalculate box at the bottom of the PCR template worksheet (see appendix PNL30-2. Vortex briefly and place on ice.

<i>Reagents</i>	<i>Fragment analysis mastermix</i>
Hi-Di Formamide	8 µl x (#samples +3)=
GeneFlo 625 bp size standard	1 µl x (# samples +3)=

- 4.9.1.7. Place a MicroAmp Optical 96-well sample plate in a cold block. Aliquot 9 µl of the prepared fragment analysis mastermix to the required number of wells. Cover the plate loosely with Parafilm.

- 4.9.1.8. Using the 10 µl multichannel pipettor, add 1 µl of 1:20 diluted PCR reactions to the appropriate columns in the sample plate. Keep sliding the Parafilm sheet from column to column to keep track of the sample order.
- 4.9.1.9. Briefly spin down the sample plate to remove any air bubbles
- 4.9.1.10. Denaturate templates by heating the reaction plate uncovered in a thermocycler at 95°C for 3 min.
- 4.9.1.11. While the templates denaturate, turn the oven of the 3500 on using the “Start Pre-Heat” button located on the middle, right-hand side of the dashboard. Make sure the set point is at 60°C.
- 4.9.1.12. Seal the plate with the rubber septa. Place the sample plate in a plate base. Snap the plate retainer onto the plate and the plate base.
- 4.9.1.13. Place the plate assembly and the buffer into the Genetic Analyzer 3500.
 - 4.9.1.13.1. Push on the tray button at the front of the Genetic Analyzer 3500 to bring the autosampler to the forward position. Open the instrument doors.
 - 4.9.1.13.2. Place the plate assembly on the autosampler in position A or B with the notched corner of the plate base facing you.
 - 4.9.1.13.3. Close the instrument doors and wait for the green light to illuminate
- 4.9.1.14. Open the previously created plate setup by clicking the blue “Edit Existing Plate” button at the top of the dashboard screen.
- 4.9.1.15. Select the plate to be run and click “Open”.
- 4.9.1.16. Link the plate by clicking “Link Plate for Run” at the bottom of the screen
- 4.9.1.17. A “Load Plate” pop-up window will establish that the plate has been successfully loaded. Click “OK”.
- 4.9.1.18. The “Start Run” button at the bottom of the screen will become functional indicating that the run can be started. Click on this button and then click “OK” on the alert window. The run will start.

4.10. Data export from the Genetic Analyzer 3500

4.10.1. DAY 2

- 4.10.1.1. *NOTE: Steps 4.10.1.3 and 4.10.1.4 only need to be performed before the very first analysis.*
- 4.10.1.2. Double-click on the shortcut icon for GeneMapper v.4.1 and enter the appropriate password to access the software. The main menu window will open.
- 4.10.1.3. **Set up the size standard:**
 - 4.10.1.3.1. From the “Tools” drop-down menu, select “GeneMapper Manager”. A “GeneMapper Manager” window will open.
 - 4.10.1.3.2. Select the “Size Standards” tab
 - 4.10.1.3.3. Click on “New”, leave the default option “Basic or Advanced” checked and click “OK”. “Size Standard Editor” window will appear.
 - 4.10.1.3.4. Name the new size standard “GeneFlo 625”
 - 4.10.1.3.5. Leave the default option “Red” as “Size Standard Dye”
 - 4.10.1.3.6. Enter sizes for each peak in the table (refer to the GeneFlo 625 product insert for the peak sizes)
 - 4.10.1.3.7. When finished, click “OK”.
 - 4.10.1.3.7.1. Click on “Apply”.
- 4.10.1.4. **Set up analysis method:**
 - 4.10.1.4.1. From the “Tools” drop-down menu, select “GeneMapper Manager”. A “GeneMapper Manager” window will open.
 - 4.10.1.4.2. Select the “Analysis Methods” tab

- 4.10.1.4.3. Click on “New”, leave the default option “Microsatellite” checked and click “OK”. “Analysis Methods Editor” window will appear.
- 4.10.1.4.4. Name the new method “SEMLVA” and click “OK”
- 4.10.1.4.5. Highlight the new method name “SEMLVA” and click “Open”
- 4.10.1.4.6. Select the “Peak Detector” tab and change “Peak Detection Algorithm” to “Advanced” from the drop-down menu
- 4.10.1.4.7. Input the following analysis settings and click “OK” when finished:
- Analysis: Full Range
 - Sizing: Partial Sizing
 - Start Size: 50
 - Stop Size 625
 - Smoothing: none
 - Baseline window: 51 pts
 - Size Calling Method: Local Southern Method
 - Peak Amplitude Thresholds
 - B: 500
 - G: 500
 - Y: 250
 - R: 20
 - O: 50
 - Min. Peak Half Width: 2 pts
 - Polynomial Degree: 2
 - Peak Window Size: 21 pts
 - Slope Threshold
 - Peak Start: 0.0
 - Peak End: 0.0
- 4.10.1.5. From the “File” drop-down menu select “Add Samples to Project”
- 4.10.1.6. Find the folder containing the data file to be analyzed: My Computer → AB SW & DATA (D:) → Applied Biosystems → 3500 → Data → SEresults-<PlateName>
- 4.10.1.7. Highlight the desired file(s) and click on “Add to List”. File(s) will appear in the window on the right. Click “Add” below the file list to return to the original screen.
- 4.10.1.8. Samples in the selected file(s) will be listed in a new window and the “Analyze” (play) button appears in green color in the toolbar indicating that the files are ready to be analyzed
- 4.10.1.9. Select the size standard GeneFlo 625 for the first sample, highlight the size standard column, and select “Fill Down” from the “Edit” drop-down menu
- 4.10.1.10. Select the analysis method SEMMLVA for the first sample, highlight the analysis method column, and select “Fill Down” from the “Edit” drop-down menu
- 4.10.1.11. Highlight all samples to be analyzed and click the “Play” icon
- 4.10.1.12. Name the project with the run name (for example, “GA070426”) and click “OK”
- 4.10.1.13. A successful analysis is indicated by green squares. Yellow triangles indicate problematic components (i.e. missing size standard peaks). Red circles indicate that results fell below acceptable quality values. Samples with yellow or red circles in the SQ column should be selected for re-analysis.
- 4.10.1.13.1. To resolve failed analyses due to sub-optimal molecular marker peak profile (i.e. miscalling of peaks), select a row with a yellow triangle or red circle in the SQ column and click on the “Size Match Editor” icon on the toolbar. “Size Match Editor” view will appear.

- 4.10.1.13.2. Place the cursor near the X-axis to activate the magnifying lens, and then pull up (mouse left-click and hold) to zoom in a specific area to facilitate editing
- 4.10.1.13.3. Left-click at the base of a peak to select. Right-click and select "Add", "Delete", or "Change".
- 4.10.1.13.4. Select the correct molecular weight for the selected peak from the drop-down menu. Repeat this process for all other miscalled peaks. Click "OK" when finished.
- 4.10.1.13.5. After the size standard has been adjusted, click the "Play" button to re-analyze the data. After a successful analysis, the samples will have green squares under the SQ column. If the size standard cannot be adjusted, the reaction is considered a fragment analysis failure and must be re-run
- 4.10.1.14. Check the fragment result data (the fluorescent peaks) for each well by highlighting the well ID and by clicking on the "Display Plots" icon on the toolbar.
 - 4.10.1.14.1. Make sure that all seven VNTRs amplified in the K1891 control and that the fragment sizes are within the range specified in the appendix PNL30-2 and record the fragment sizes on the PCR template worksheet.
 - 4.10.1.14.2. The size calling for the internal ladder should also be within the range specified in the appendix PNL30-2 (or PNL30-4).
 - 4.10.1.14.3. Write down any failed reactions in the PCR template worksheet. Make a note of non-specific bands and primer-dimers.
- 4.10.1.15. Export the peak file:
 - 4.10.1.15.1. *NOTE: the following columns should appear in the exported table in the following order from left to right: "Dye/Sample", "Sample file name", "Marker", "Size", "Height", "Area", "Data point". The format of the table can be modified by selecting "Table setting editor" from the "Tools" drop down menu. Select the "Genotype" tab and make sure that the boxes for the above mentioned columns are checked and no additional boxes are checked.*
 - 4.10.1.15.2. Highlight the samples for which you want to export peak data
 - 4.10.1.15.3. Click on "Display Plots" icon on the toolbar
 - 4.10.1.15.4. Click on "Sizing Table" icon on the toolbar and a table will appear below the electropherograms
 - 4.10.1.15.5. From the "File" drop-down menu, select "Export Table"
 - 4.10.1.15.6. Select the location (for example a flash drive) where you want to export the data
 - 4.10.1.15.7. Name the export file with the run name (for example GA070426) and make sure the file type is tab-delimited text (.txt) file
- 4.10.1.16. The remaining gel can stay in the instrument if it is going to be used within seven days

5. Bibliography

- 5.1. Seongbeom Cho, David J. Boxrud, Joanne M. Bartkus, Thomas S. Whittam, Mahdi Saeed. Multiple-locus variable-number tandem repeat analysis of *Salmonella* Enteritidis isolates from human and non-human sources using a single multiplex PCR. (2007) FEMS Microbiology Letters 275 (1), 16-23.
- 5.2. D. Boxrud, K. Pederson-Gulrud, J. Wotton, C. Medus, E. Lyszkowicz, J. Besser, and J. M. Bartkus. Comparison of Multiple-Locus Variable-Number Tandem Repeat Analysis, Pulsed-Field Gel Electrophoresis, and Phage Typing for Subtype Analysis of *Salmonella enterica* serotype Enteritidis. (2007) Journal of Clinical Microbiology, 45 (2), 536-543.

6. Contacts: PulseNet Next Generation Subtyping Unit, EDLB, DFWED, CDC

- | | | |
|-------------------------------|----------------|-----------------------|
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Appendix PNL30-1
***S. enterica* serotype Enteritidis MLVA PCR Primers**

<i>R1 Locus</i>	<i>Dye</i>	<i>Forward Primer (5' to 3')</i>	<i>Reverse Primer (5' to 3')</i>
SE-1	FAM	TGT GGG ACT GCT TCA ACC TTT GGG C	CCA GCC ATC CAT ACC AAG ACC AAC ACT CTA TGA
SE-2	CallRed590	GTG CTT CCT CAG GTT GCT TTT AGC CTT GTT CG	GGG GAA TGG ACG GAG GCG ATA GAC G
SE-8	HEX	GGT AGC TTG CCG CAT AGC AGC AGA AGT	GGC GGC AAG CGA GCG AAT CC
SE-6	FAM	CTG GTC GCA GGT GTG GC	GGT GAC GCC GTT GCT GAA GGT AAT AAC AGA GTC

<i>R2 Locus</i>	<i>Dye</i>	<i>Forward Primer (5' to 3')</i>	<i>Reverse Primer (5' to 3')</i>
SE-5	HEX	GGC TGG CGG GAA ACC ACC ATC	GCC GAA CAG CAG GAT CTG TCC ATT AGT CAC TG
SE-3	CallRed590	CGG GAT AAG TGC CAC ATA ACA CAG TCG CTA AGC	CGC CAG TGT TAA AGG AAT GAA TGA ACC TGC TGA TG
SE-9	FAM	CCA CCT CTT TAC GGA TAC TGT CCA CCA GC	GGC GTT ACT GGC GGC GTT CG

Primer sets are listed as multiplex PCR cocktails.

Appendix PNL30-2

S. enterica serotype Enteritidis MLVA PCR template Worksheet for ABI

PCR run date: _____
 PCR instrument: _____
 ABI run date: _____
 ABI instrument: _____

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												

	1	[put in # of samples only]
SLS	32	
600 bp	4.00	

Hi-Di lot no.
 625 bp lot no.
 Polymer lot no.
 Buffer lot no.

<i>Salmonella</i> Enteritidis - Positive Control (K1891)			
R1 fragment sizes		R2 fragment sizes	
SE1 (190 - 193)		SE9 (181 - 184)	
SE2 (335 - 339)		SE5 (201 - 203)	
SE8 (433 - 436)		SE3 (211 - 214)	
SE6 (479 - 482)			

<i>Salmonella</i> Enteritidis - Internal Ladder fragment sizes			
Expected Size Ranges		Fragment Sizes	
SE9	181 - 184		
SE1	190 - 193	211 - 212	
SE3	199 - 203	211 - 214	
SE5	201 - 203	218 - 221	
SE2	317.5 - 324	363 - 364	
SE8	346 - 350	433 - 436	
SE6	446 - 447	479 - 482	

- For each isolate, fill in an appropriate BioNumerics key number for PCR reactions R1 and R2 (for example: for an isolate K1720 the BN key numbers are K1720R1 and K1720R2).
- Fill in the exact number of fragment analysis reactions (2 per isolate + controls) in the red box (extra already included in calculation). Mix the Hi-Di Formamide and 625 bp ladder in an appropriate tube. Add 9 µl of mixture to each well.
- Using a multichannel pipettor, add 1 ul of diluted PCR reaction to wells containing 9 ul of Formamide/625 bp ladder mixture.
- Denature the plate at 95°C for 3 min
- Briefly spin down the plate.
- Place the plate in the plate base; cover the wells with a rubber septa; finish the plate assembly by snapping on the plate retainer.

Appendix PNL30-3

PCR Mastermix Calculations

NOTE: This is posted on SharePoint as an Excel file so users may save it locally, enter data, and use the auto calculate function.

Mastermix calculations

Enter number of reactions (+ 2 extra), currently a red zero, next to "Number to analyze" for autocalculations.

by:

cycler:

Appendix PNL30-3

Itids multiplex PCR **R1**

Number of samples to be analyzed

Total Reaction Volume

	Start	Final	Add	
PCR Buffer	10 X	1 X	1.00 ul	
MgCl2	50 mM	2 mM	0.40 ul	
dNTPs	10 mM	0.2 mM	0.20 ul	
SE1F	2.5 uM	0.05 uM	0.20 ul	
SE1R	2.5 uM	0.05 uM	0.20 ul	
SE2F	12.5 uM	0.4 uM	0.32 ul	
SE2R	12.5 uM	0.4 uM	0.32 ul	
SE3F	2.5 uM	0.05 uM	0.20 ul	
SE3R	2.5 uM	0.05 uM	0.20 ul	
SE6F	12.5 uM	0.43 uM	0.34 ul	
SE6R	12.5 uM	0.43 uM	0.34 ul	
Platinum taq	5 U/ul	1 U	0.20 ul	
PCR Water			4.91 ul	

Row 1	Row 2	Row 3	Row 4	Row 5	Row 6

Primer	Date prepared	PCR Reagents		
		Lot#	Exp.	
SE1F				
SE1R		10X Buffer		
SE2F		MgCl2		
SE2R		dNTPs		
SE3F		Platinum Taq		
SE3R				
SE6F				
SE6R				

Itids multiplex PCR **R2**

Number of samples to be analyzed

Total Reaction Volume

	Start	Final	Add	
PCR Buffer	10 X	1 X	1.00 ul	
MgCl2	50 mM	2 mM	0.40 ul	
dNTPs	10 mM	0.2 mM	0.20 ul	
SE5F	2.5 uM	0.05 uM	0.20 ul	
SE5R	2.5 uM	0.05 uM	0.20 ul	
SE3F	12.5 uM	0.5 uM	0.40 ul	
SE3R	12.5 uM	0.5 uM	0.40 ul	
SE9F	2.5 uM	0.02 uM	0.08 ul	
SE9R	2.5 uM	0.02 uM	0.08 ul	
Platinum taq	5 U/ul	1 U	0.20 ul	
PCR Water			5.84 ul	

Row 1	Row 2	Row 3	Row 4	Row 5	Row 6

Primers	Date prepared
SE5F	
SE5R	
SE3F	
SE3R	
SE9F	
SE9R	

Appendix PNL30-4

Instructions to prepare the internal ladder

1. Prepare DNA templates from isolates H9560 and 2010K0017 as described in the protocol step 4.5. Store the templates at -20°C or -80°C freezer.
2. Use the DNA templates to set up and run the PCR reactions R1 and R2 as described in the protocol step 4.6.
3. After the PCR amplification, pool the R1 and R2 reactions for isolates H9560 and 2010K0017 into one single PCR tube to gain a total volume of 40 µl.
4. Each new lot of internal ladder must be tested against the old ladder lot by running them in the same fragment analysis run.
5. Store the ladder in -20°C or -80°C freezer. It should remain stable at least 5-6 freeze-thaw cycles for a period of one month.

Expected fragment sizes (bp) of the thirteen fragments present in the internal ladder from smallest to the largest in the order they appear in the electropherogram:

SE9 (B)	181-184	
SE1 (B)	190-193	211-212
SE3 (B)	199-203	211-214
SE5 (G)	201-203	218-221
SE2 (G)	317.5-324	363-364
SE8 (Y)	346-350	433-436
SE6 (Y)	446-447	479-482

Expected fragment sizes (bp) of the thirteen fragments present in the internal ladder by the dye in the order they appear in the peak file:

SE3 (B)	199-203	211-214
SE2 (G)	317.5-324	363-364
SE5 (G)	201-203	218-221
SE8 (Y)	346-350	433-436
SE9 (B)	181-184	
SE1 (B)	190-193	211-212
SE6 (Y)	446-447	479-482

NOTE: fragment sizes for the internal ladder based on independent runs on multiple instruments at CDC and in PulseNet Participating Laboratories

Appendix PNL30-5

Steps for exporting and importing plate set ups on the ABI 3500

Note: the procedure described below will require the sequencer computer to have Microsoft Office. If the sequencer computer does not have Microsoft Office, the exported file needs to be saved on a flash drive so that steps 8 through 15 can be performed on a computer with Microsoft Office.

1. On the Dashboard screen click on the “Edit Existing Plate” button.
2. Select “Fragment” from the “Filter” drop-down menu.
3. Find the most recent full plate. Click on the ID to highlight the desired plate and then click the “Open” button at the bottom of the window.
NOTE: If you do not have a full plate, create a new one by filling in each sample ID space (A01 through H12) with “test” and saving it with the plate name “SE_Template”. Be sure to add the results group “SEresults”, file name convention “MLVA”, and Assay “SE_Fragtest” to each sample, making sure each one is check-marked. It is necessary to have all required wells filled in when exporting so that they will be available in your exported file.
4. In the plate layout window, click the “Export...” button in the toolbar at the top of the screen.
5. In the export window, create a new folder on the desktop by clicking the “Desktop” icon on the left side of the window. Find the ‘New Folder’ button in the toolbar at the top of the window and name the new folder “MLVA Plate Setup”.
6. Save the file to this folder by using your new plate name (e.g. - CDC110726) and clicking “Save”.
7. A window will pop up letting you know that the plate has been successfully exported. Click “OK”.
8. Click the “Close Plate” button in the toolbar at the top of the plate layout screen, and click “Yes” to close without saving changes.
9. Minimize the 3500 program window and open the “MLVA Plate Setup” folder on the desktop.
10. Right-click your plate name, select “Open With” and in the submenu select Microsoft Office Excel.
11. Sample IDs can be typed or copy and pasted from a separate Excel file into the spaces next to the correct wells under the “Sample Name” heading.
NOTE: Do not change/delete any of the column headings. The fields must be in the same format when importing as they were when exported. Additionally, the software will not import the file if the sample IDs contain special characters (e.g. - ! , / ,) , etc). You can use underscores and dashes.
12. The “Plate Name” should be changed to match the plate/file name, and initials should be placed under the ‘Owner Name’ heading. The defaults for “Application Type”, “Capillary Length (cm)”, “Polymer”, and “Number of Wells” should be used.
13. Once all of the IDs are inserted, make sure that for each sample ID and all controls, the assay is “SE_Fragtest”, the results group name is “SEresults”, the file name convention is “MLVA”, and “Sample” is listed under “Sample Type”.
14. Under the “File” drop-down menu select “Save”.
15. A warning window will appear asking if the workbook should be saved in the “Text (tab-delimited)” format. Click “Yes”.
16. Exit Microsoft Excel, clicking “No” when prompted to save changes.

NOTE: The removal of additional spaces from lines in the .txt file after modification in Excel may be necessary before importing into the "3500" program. To do this, open the file in "Notepad" making sure the window is opened in full screen. In lines 1, 2, and 4 remove any additional spaces after the last character in the line or from the left side of the page. Once all spaces have been removed from those lines, save the file by clicking "Save" in the "File" drop-down menu and close the window.

17. Go to the ABI 3500 plate layout window by clicking "Create New Plate" on the Dashboard.
18. Enter in the name of the plate and from the "Plate Type" drop-down menu select "Fragment".
19. Click on the "Assign Plate Contents" button at the bottom of the screen.
20. In the toolbar at the top of the "Plate View" screen, click the "Import..." button.
21. In the import window, navigate to the recently created plate under Desktop/MLVA Plate Setup. Select the appropriate .txt file and click "Open".
22. At this point an error message may appear asking if you would like to proceed. If you encounter this message, click the "Proceed" button at the bottom of the window.
23. When the file has been imported, an alert window will appear stating that the plate "has been successfully imported". Click "OK".
24. In the toolbar at the top of the screen click on "Save Plate" and from the drop-down menu select "Save As". Enter the plate name matching the name of the .txt file, and click "OK".

NOTE: When importing a plate layout from a .txt file the Assay, File Name Convention and Results Group should automatically be applied to the plate and check-marked for all samples. If the .txt file does not contain the correct assay file name (e.g. – the name is misspelled, etc.), the assay will not be imported and the file name convention and results group will be imported but not check-marked.

NOTE: For repeated use, plate templates can be created by exporting a plate to the "MLVA Plate Setup" folder. Save the file as a tab-delimited text file with "test" as the sample ID and with a new plate name (e.g. - "SE_Template"). After you have filled out the information for a new run on the template, save it in step 14 with the standardized run name (name (e.g. - CDC101020). Since the Assay and Results Group are different for each MLVA assay (STECO157, Salmonella Typhimurium, Salmonella Enteritidis, etc), separate templates must be exported and saved accordingly.