

Japanese Encephalitis Vaccine: Recommendations of the Advisory Committee on Immunization Practices



CONTENTS

Introduction	1
Methods.....	2
Background	2
JE-VC	8
Recommendations for the Prevention of JE	
Among U.S. Travelers	16
Recommendations for the Prevention of JE	
Among Laboratory Workers.....	16
Administration of JE Vaccine.....	17
Contraindications and Precaution for the Use of JE Vaccine	19
Special Populations	19
Reporting of Vaccine Adverse Events	19
Future Research on JE-VC.....	19
Additional Information.....	19
References.....	20

The *MMWR* series of publications is published by the Center for Surveillance, Epidemiology, and Laboratory Services, Centers for Disease Control and Prevention (CDC), U.S. Department of Health and Human Services, Atlanta, GA 30329-4027.

Suggested citation: [Author names; first three, then et al., if more than six.] [Title]. *MMWR Recomm Rep* 2019;68(No. RR-#):[inclusive page numbers].

Centers for Disease Control and Prevention

Robert R. Redfield, MD, *Director*
 Anne Schuchat, MD, *Principal Deputy Director*
 Chesley L. Richards, MD, MPH, *Deputy Director for Public Health Science and Surveillance*
 Rebecca Bunnell, PhD, MEd, *Director, Office of Science*
 Barbara Ellis, PhD, MS, *Acting Director, Office of Science Quality, Office of Science*
 Michael F. Iademarco, MD, MPH, *Director, Center for Surveillance, Epidemiology, and Laboratory Services*

MMWR Editorial and Production Staff (Serials)

Charlotte K. Kent, PhD, MPH, *Editor in Chief*
 Christine G. Casey, MD, *Editor*
 Mary Dott, MD, MPH, *Online Editor*
 Terisa F. Rutledge, *Managing Editor*
 David C. Johnson, *Lead Technical Writer-Editor*
 Catherine B. Lansdowne, MS, *Project Editor*

Martha F. Boyd, *Lead Visual Information Specialist*
 Maureen A. Leahy, Julia C. Martinroe,
 Stephen R. Spriggs, Tong Yang,
Visual Information Specialists
 Quang M. Doan, MBA, Phyllis H. King,
 Terraye M. Starr, Moua Yang,
Information Technology Specialists

MMWR Editorial Board

Matthew L. Boulton, MD, MPH	Timothy F. Jones, MD, <i>Chairman</i>	Stephen C. Redd, MD
Virginia A. Caine, MD	Robin Ikeda, MD, MPH	Patrick L. Remington, MD, MPH
Katherine Lyon Daniel, PhD	Phyllis Meadows, PhD, MSN, RN	Carlos Roig, MS, MA
Jonathan E. Fielding, MD, MPH, MBA	Jewel Mullen, MD, MPH, MPA	William Schaffner, MD
David W. Fleming, MD	Jeff Niederdeppe, PhD	Morgan Bobb Swanson, BS
William E. Halperin, MD, DrPH, MPH	Patricia Quinlisk, MD, MPH	

Japanese Encephalitis Vaccine: Recommendations of the Advisory Committee on Immunization Practices

Susan L. Hills, MBBS¹; Emmanuel B. Walter, MD²; Robert L. Atmar, MD³; Marc Fischer, MD¹

¹*Division of Vector-Borne Diseases, National Center for Emerging and Zoonotic Infectious Diseases, CDC, Fort Collins, Colorado;*

²*Duke University School of Medicine, Durham, North Carolina;* ³*Baylor College of Medicine, Houston, Texas*

Summary

This report updates the 2010 recommendations from the CDC Advisory Committee on Immunization Practices (ACIP) regarding prevention of Japanese encephalitis (JE) among U.S. travelers and laboratory workers (Fischer M, Lindsey N, Staples JE, Hills S. Japanese encephalitis vaccines: recommendations of the Advisory Committee on Immunization Practices (ACIP). *MMWR Recomm Rep* 2010;59[No. RR-1]). The report summarizes the epidemiology of JE, describes the JE vaccine that is licensed and available in the United States, and provides recommendations for its use among travelers and laboratory workers.

JE virus, a mosquito-borne flavivirus, is the most common vaccine-preventable cause of encephalitis in Asia. JE occurs throughout most of Asia and parts of the western Pacific. Approximately 20%–30% of patients die, and 30%–50% of survivors have neurologic, cognitive, or behavioral sequelae. No antiviral treatment is available.

Inactivated Vero cell culture–derived JE vaccine (Ixiaro [JE-VC]) is the only JE vaccine that is licensed and available in the United States. In 2009, the U.S. Food and Drug Administration (FDA) licensed JE-VC for use in persons aged ≥17 years; in 2013, licensure was extended to include children aged ≥2 months.

Most travelers to countries where the disease is endemic are at very low risk for JE. However, some travelers are at increased risk for infection on the basis of their travel plans. Factors that increase the risk for JE virus exposure include 1) traveling for a longer period; 2) travel during the JE virus transmission season; 3) spending time in rural areas; 4) participating in extensive outdoor activities; and 5) staying in accommodations without air conditioning, screens, or bed nets. All travelers to countries where JE is endemic should be advised to take precautions to avoid mosquito bites to reduce the risk for JE and other vectorborne diseases. For some persons who might be at increased risk for JE, the vaccine can further reduce the risk for infection. The decision about whether to vaccinate should be individualized and consider the 1) risks related to the specific travel itinerary, 2) likelihood of future travel to countries where JE is endemic, 3) high morbidity and mortality of JE, 4) availability of an effective vaccine, 5) possibility (but low probability) of serious adverse events after vaccination, and 6) the traveler's personal perception and tolerance of risk.

JE vaccine is recommended for persons moving to a JE-endemic country to take up residence, longer-term (e.g., ≥1 month) travelers to JE-endemic areas, and frequent travelers to JE-endemic areas. JE vaccine also should be considered for shorter-term (e.g., <1 month) travelers with an increased risk for JE on the basis of planned travel duration, season, location, activities, and accommodations and for travelers to JE-endemic areas who are uncertain about their specific travel duration, destinations, or activities. JE vaccine is not recommended for travelers with very low-risk itineraries, such as shorter-term travel limited to urban areas or outside of a well-defined JE virus transmission season.

Introduction

Japanese encephalitis (JE) virus, a mosquito-borne flavivirus, is the most common vaccine-preventable cause of encephalitis in Asia (1,2). JE occurs throughout most of Asia and parts of the western Pacific (3,4). Approximately 20%–30% of patients die, and 30%–50% of survivors have neurologic, cognitive, or behavioral sequelae (5–7). In countries where the disease is endemic, JE primarily affects children. Although

rare, travel-associated JE can occur among persons of any age (8–10). For most travelers to Asia, the risk for JE is very low but varies based on travel duration, season, location, activities, and accommodations (9,11).

JE virus is maintained in an enzootic cycle between mosquitoes and amplifying vertebrate hosts, primarily pigs and wading birds (12,13). JE virus is transmitted to humans by infected mosquitoes (1). JE virus transmission occurs primarily in rural agricultural areas. In most temperate areas of Asia, JE virus transmission is seasonal, and large outbreaks can occur. In the subtropics and tropics, transmission can occur year-round, often intensifying during the rainy season.

Corresponding author: Susan L. Hills, National Center for Emerging and Zoonotic Infectious Diseases, CDC. Telephone: 970-221-6400; E-mail: shills@cdc.gov.

Inactivated Vero cell culture–derived JE vaccine (Ixiaro [JE-VC]) is the only JE vaccine that is licensed and available in the United States. An inactivated mouse brain–derived vaccine (JE-VAX [JE-MB]) has been licensed in the United States since 1992 but is no longer produced, and all remaining doses expired in 2011. In 2009, the U.S. Food and Drug Administration (FDA) licensed JE-VC for use in persons aged ≥ 17 years (14). In 2013, licensure was extended to include children aged ≥ 2 months. This report updates the 2010 Advisory Committee on Immunization Practices (ACIP) recommendations for use of JE vaccine among U.S. travelers and laboratory workers (15).

Methods

The ACIP JE Vaccine Work Group was initially formed in 2006 to review and update information on JE vaccines available in the United States. FDA licensed JE-VC for adults aged ≥ 17 years in 2009. Updated ACIP recommendations for use of JE vaccine among U.S. travelers and laboratory workers were published in 2010 (15). ACIP subsequently approved recommendations for use of a booster dose of JE-VC in adults in 2011 and recommendations for use of JE-VC in children in 2013 after the FDA extension of JE-VC licensure to include children aged ≥ 2 months (16,17). The ACIP JE Vaccine Work Group was then disbanded and reformed in 2015 with revised membership. The objectives of the work group were to 1) review newly available safety and immunogenicity data for JE-VC, 2) review updated information on the epidemiology and risk for JE in travelers, and 3) review recommendations for use of JE vaccine in consideration of these data. Work group members included persons with expertise in JE, infectious diseases, pediatrics, travel medicine, public health, vaccination safety, and vaccine policy. The work group met approximately 34 times by teleconference during March 2015–January 2019. Presentations on vaccine immunogenicity and safety and on other topics related to the development of the JE vaccine recommendations were made to ACIP by the manufacturer or work group members.

Grading of Recommendations, Assessment, Development, and Evaluation (GRADE) methods were used to review and evaluate newly available data (18,19). Additional factors also were assessed in developing JE vaccine recommendations as outlined in the Evidence to Recommendations framework, including target population values, stakeholder acceptability, and feasibility of implementation (19,20). Details on the methods used for GRADE, including the search protocol, databases searched, and inclusion criteria, a summary of the evidence, the grading of the evidence, and information on the additional factors considered, are provided in

Japanese Encephalitis Vaccine Evidence to Recommendations (19). The work group presented preliminary recommendations to ACIP during its October 2018 meeting. Proposed recommendations were presented to ACIP and approved at the February 2019 meeting. ACIP will review additional data as they become available, and recommendations will be updated as needed.

Background

JE Virus Description

JE virus, an arthropodborne virus (arbovirus), is a single-stranded RNA virus that belongs to the genus *Flavivirus* and is closely related to West Nile, St. Louis encephalitis, yellow fever, and dengue viruses (21,22). Five genotypes of JE virus have been identified (23). Until the 1990s, the dominant JE virus genotype in Asia was genotype III but is now genotype I (23).

JE Virus Transmission

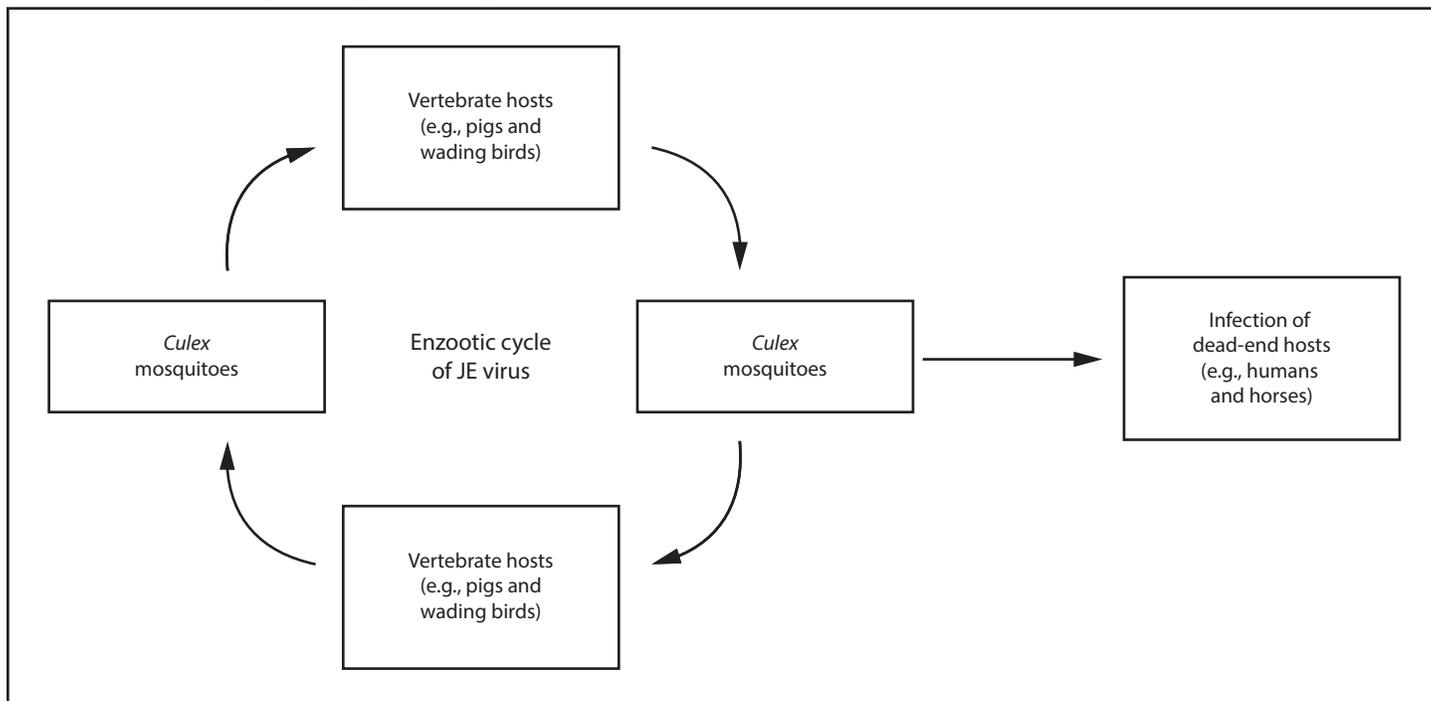
JE virus is transmitted in an enzootic cycle between mosquitoes and amplifying vertebrate hosts, primarily pigs and wading birds such as herons and egrets (Figure 1) (13,24–28). Because of rapid population turnover with numerous susceptible offspring and the development of high-titer viremia, domestic pigs are the main source of infection for mosquitoes that transmit JE virus to humans (12,28–32).

Culex mosquitoes, especially *Cx. tritaeniorhynchus*, are the principal vector for JE virus transmission in most of Asia (12,13,24,27,33–40). *Cx. tritaeniorhynchus* is an evening- and nighttime-biting mosquito that feeds preferentially on large domestic animals and birds but only infrequently on humans (41). *Cx. tritaeniorhynchus* feed most often in the outdoors, with peak feeding activity occurring after sunset (41). Larvae are found in flooded rice fields, marshes, and other stagnant collections of water (38,39). In temperate zones, the mosquito is present in the greatest density during June–November and is inactive during winter months (12,26,42). In certain parts of Asia and the Western Pacific, other mosquito species also are important JE virus vectors (13,37,39,43).

Infected mosquitoes transmit JE virus to humans. Humans are considered dead-end hosts in the JE virus transmission cycle because they do not develop a level or duration of viremia sufficient to infect mosquitoes (13,44). Therefore, travelers with JE virus infection who return to nonendemic areas pose minimal or no risk for subsequent transmission of the virus.

JE virus is not spread from person to person through direct contact. A small number of cases of transplacental transmission of JE virus has been reported. Four miscarriages were documented among nine infected pregnant women

FIGURE 1. Transmission cycle of Japanese encephalitis virus*



Abbreviation: JE = Japanese encephalitis.

* JE virus is transmitted in an enzoitic cycle between *Culex* mosquitoes and amplifying vertebrate hosts, primarily pigs and wading birds. Humans are a dead-end host in the JE virus transmission cycle, with brief and low levels of viremia. Humans play no role in the maintenance or amplification of JE virus, and the virus is not transmitted directly from person to person.

during outbreaks in India (3,45,46). All of the women who miscarried were in the first or second trimester of pregnancy, and JE virus was isolated from one of the four aborted fetuses. JE virus transmission through blood transfusion has been documented in a JE-endemic area, and on the basis of experience with similar flaviviruses, organ transplantation is considered a potential mode of transmission (47,48). In a laboratory setting, JE virus might be transmitted through accidental percutaneous exposure, or theoretically, mucosal or inhalational exposure. At least 22 laboratory-acquired JE virus infections have been reported (49).

Epidemiology of JE

Geographic Distribution and Spread

JE occurs throughout most of Asia and parts of the western Pacific (Figure 2). During the first half of the 20th century, the disease was recognized principally in temperate areas of Asia including Japan, Korea, Taiwan, and China (50–54). The virus then spread south and west, with increased transmission reported in Southeast Asia, India, Bangladesh, Sri Lanka, and Nepal (36,54–68). In the 1990s, JE virus spread east and was recognized for the first time in Saipan and then Australia, initially in the outer Torres Strait Islands and subsequently on

the northern mainland (43,69,70). More recently, transmission also has been detected in new areas, including in Tibet and mountain districts in Nepal (71,72). The reasons for this increased geographic distribution are uncertain but might include population shifts or changes in climate, ecology, agricultural practices, animal husbandry, or migratory bird patterns (39,56,70). These factors could contribute to further spread, including beyond Asia and the western Pacific.

Incidence

In the early 1970s, approximately 100,000 cases of JE were reported each year, with the vast majority from China (54). Because of vaccine use, increased urbanization, changes in agricultural practices, and mosquito control, annual JE case counts have decreased substantially. Up to 5,000 cases of JE are reported to the World Health Organization (WHO) each year (73). However, this number likely represents an underestimate of the actual number of cases because of limited diagnostic testing and surveillance capacity in many countries with endemic JE (5,7). In 2011, taking into account the status of vaccination programs at that time, a systematic review estimated that 67,900 JE cases typically occurred annually, with an overall incidence of 1.8 cases per 100,000 population. In children aged <15 years, the incidence was estimated to be 5.4 cases per 100,000 (5). However, incidence can vary

FIGURE 2. Approximate geographic range of Japanese encephalitis



Source: Hills SL, Lindsey NP, Fischer M. Japanese encephalitis. In: CDC Yellow Book 2020: health information for international travel. New York, NY: Oxford University Press; 2019:248–57.

substantially by year and area. Before the introduction of vaccination programs, the highest risk areas in Asia had incidence rates of laboratory-confirmed JE as high as 20 cases per 100,000 children per year (5,74–76). In countries with vaccination programs with high coverage, JE incidence is now less than one case per 100,000 children per year (5,77).

Ecologic and Seasonal Patterns

The risk for JE varies by local ecology and season. JE virus transmission primarily occurs in rural agricultural areas, often associated with rice production and flood irrigation, where large numbers of vector mosquitoes breed in proximity to animal reservoirs (24,27). In some areas of Asia, these ecologic conditions might occur near, or within (although rare), urban centers (78–80).

In temperate areas of Asia (e.g., China, Japan, Nepal, northern Vietnam, northern India, South Korea, and Taiwan), JE virus transmission is seasonal, and human disease usually

peaks in the summer and fall (50,52,53,57,66,68,81). The peak months of transmission and the length of the season vary by region, and large, explosive outbreaks can occur. In the subtropics and tropics, transmission can occur year-round, often with a peak during the rainy season (56,58,62,76,82).

Age-Specific Patterns

In areas with endemic JE, the disease primarily affects children, with the vast majority of cases occurring among children aged <15 years; most adults have protective immunity after natural exposure to the virus (52,53,57–59,66,68,76,83,84). However, in areas with childhood JE vaccination programs, the overall incidence of JE decreases, with a greater proportion of cases occurring among adults (50,51,85,86). Outbreaks that predominantly affected older adults have been reported in Japan, China, and India (87–89). Because unvaccinated travelers from nonendemic countries are

usually immunologically naïve, travel-associated JE can occur in persons of any age.

Clinical Manifestations and Diagnosis

Signs and Symptoms

The majority of JE virus infections in humans are asymptomatic, and <1% of persons infected with JE virus develop encephalitis (83,90–94). Acute encephalitis is the most commonly identified clinical syndrome among persons with JE virus infection, although milder forms of disease (e.g., aseptic meningitis or undifferentiated febrile illness) also can occur (6,13,95–97). Among patients who develop clinical symptoms, the incubation period is 5–15 days. Initial symptoms are usually nonspecific and might include fever, rigors, headache, vomiting, and diarrhea (6,61,98,99). Mental status changes, generalized weakness, focal neurologic deficits (e.g., hemiplegia, tetraplegia, or cranial nerve palsies), and movement disorders might occur over the next few days (61,98–103). Seizures are common, especially among children (61,98–100,103–105). A distinctive clinical presentation of JE is a parkinsonian syndrome resulting from extrapyramidal involvement, with mask-like facies, tremor, cogwheel rigidity, and choreoathetoid movements (6,99). Acute flaccid paralysis, with clinical and pathological features similar to poliomyelitis, also has been associated with JE virus infection (6,106,107). Status epilepticus, brain hypoxia, increased intracranial pressure, brainstem herniation, and aspiration pneumonia are the most common complications associated with poor outcome and death (6,98,104,108).

Clinical Laboratory Findings and Neuroimaging

Clinical laboratory findings with JE are nonspecific and might include moderately elevated white blood cell count, mild anemia, and hyponatremia (6,95,98,99,103). Thrombocytopenia and elevated hepatic enzymes have been reported (99). Cerebrospinal fluid (CSF) usually shows a lymphocytic pleocytosis with moderately elevated protein levels (6,59,61,95,98,100,103,109).

Magnetic resonance imaging (MRI) is the best means for detecting JE-associated abnormalities of the brain, including changes in the thalamus, basal ganglia, midbrain, pons, and medulla (110–112). Thalamic lesions are the most commonly described abnormality (110,112).

Laboratory Diagnosis

JE virus infections are usually confirmed by detection of virus-specific antibody in CSF or serum (13,113–117). Because humans have low or undetectable levels of viremia by the time the clinical illness occurs, virus isolation and nucleic acid

amplification tests (NAATs) are insensitive and should not be used for ruling out a JE diagnosis (118,119). In one study in Thailand, JE virus could not be isolated from 30 nonfatal JE cases with plasma and CSF samples (120). In contrast, JE virus was isolated from CSF from five (33%) of 15 patients who died and from brain tissue from eight (73%) of 11 who died. More recent studies have demonstrated the usefulness of NAAT to diagnose JE in some patients with encephalitis or aseptic meningitis, and JE virus RNA was detected in urine of a patient who died (97,121,122). However, testing by NAAT lacks the sensitivity needed for routine diagnosis.

Acute-phase specimens should be tested for JE virus immunoglobulin M (IgM) antibodies using an IgM antibody-capture enzyme-linked immunosorbent assay (MAC ELISA) (13,113–117). JE virus IgM antibodies can be measured in the CSF of most patients within 4 days of onset of symptoms and in serum by 7–8 days after onset (76,115,116,123,124). The presence of JE virus IgM antibodies in CSF provides evidence that JE virus infection is the cause of the neurologic illness (114,119). With clinical and epidemiologic correlation, a positive IgM test has good diagnostic predictive value, although cross-reaction with other flaviviruses can occur.

Plaque reduction neutralization tests (PRNTs) can be performed to confirm recent infection on the basis of a fourfold or higher rise in virus-specific neutralizing antibodies between acute- and convalescent-phase serum specimens or to discriminate between cross-reacting antibodies attributed to another primary flavivirus infection. In patients who have been infected previously by another flavivirus or vaccinated with a flaviviral vaccine (e.g., yellow fever), cross-reactive antibodies in both the ELISA and neutralization assays make identifying a specific etiologic agent difficult.

Vaccination history, date of onset of symptoms, and information regarding other arboviruses known to circulate in the geographic area that might cross-react in serologic assays should be considered when interpreting results. Diagnostic testing for JE is available in some state public health laboratories and at CDC.

Treatment and Management

JE treatment consists of supportive care and management of complications. No antiviral agent or specific medication is available to mitigate the effects of JE virus infection (125). In controlled clinical trials, clinical outcomes were not improved with corticosteroids, interferon alpha-2a, ribavirin, minocycline, or intravenous immunoglobulin (126–130). Infection with one JE virus genotype is thought to produce lifelong immunity against all genotypes.

Outcome and Sequelae

JE has a case-fatality ratio of 20%–30% (6,52,53,62,68,84,98–100,109,120,127,131,132). Some deaths occur after a short fulminant course, whereas others occur after a prolonged coma. Although some motor deficits and movement disorders improve after the acute illness, 30%–50% of JE survivors have neurologic or other sequelae even years later (6,100,106,127,131–138). These include seizures, upper and lower motor neuron weakness, cerebellar and extrapyramidal signs, flexion deformities of the arms, hyperextension of the legs, cognitive deficits, language impairment, psychiatric issues, learning difficulties, and behavioral problems (6).

JE Among Travelers

For most travelers to Asia, the risk for JE is very low but varies on the basis of travel destination, duration, season, activities, and accommodations (4,8,11,139). The overall incidence of JE among persons from nonendemic countries who travel to Asia is estimated to be less than one case per 1 million travelers. However, persons who stay for prolonged periods in rural areas with active JE virus transmission might have a risk level similar to that of the susceptible resident population. Travelers on brief trips might be at increased risk if they have extensive outdoor or nighttime exposure in rural areas during periods of active transmission (140–142). Shorter-term (e.g., <1 month) travelers whose visits are restricted to major urban areas are at minimal risk for JE.

Risk for infection for a traveler cannot be inferred from JE incidence among residents of JE-endemic countries. Very few cases might be reported among the local population because of vaccination or natural immunity from previous infection. However, because JE virus is maintained in an enzootic cycle between animals and mosquitoes, susceptible visitors might still be at risk for infection. JE should be suspected in any patient with evidence of a neurologic infection (e.g., encephalitis, meningitis, or acute flaccid paralysis) who recently has returned from a country in Asia or the western Pacific where JE is endemic.

JE Among All Travelers from Nonendemic Countries

During 1973–2017, a total of 85 JE cases among travelers or expatriates from nonendemic countries were published or reported to CDC (8–10,122,140–174). About twice as many cases were reported in the most recent 10-year period compared with the three previous 10-year periods: 2008–2017 (n = 34), 1998–2007 (n = 18), 1988–1997 (n = 17), and 1978–1987 (n = 13). This change might relate to increased numbers of travelers and increased testing and reporting of disease cases. Overall, 53 (62%) cases occurred in tourists, 16 (19%) in expatriates, six (7%) in soldiers, and one (1%)

in a researcher; the type of travel was unknown in nine (11%) cases. The tourist category included seven persons who were traveling to visit friends and relatives and two students on study-abroad programs. The patients were citizens of 20 different countries. The countries where the infection was most commonly acquired were Thailand (n = 26), Indonesia (n = 13), the Philippines (n = 11), China (n = 9), Vietnam (n = 4), and Japan (n = 4). The countries with the highest number of cases (i.e., Thailand and Indonesia) have high-risk areas but also are destinations with high numbers of tourists. In both countries, tourist beach resort areas can be close to rice fields or rural areas with high mosquito densities (e.g., Phuket, Thailand, and Bali, Indonesia). Among the 76 cases for which age was recorded, the median age was 36 years (range: 5 weeks to 91 years). Overall, 50 (59%) cases occurred among males, and 31 (36%) among females; sex was unknown in four cases (5%). Nineteen (22%) patients recovered fully, 39 (46%) survived but had sequelae, 14 (16%) died, and the outcome was unknown for 13 (15%). None of the patients were known to have received JE vaccine. No cases occurred among business or other shorter-term travelers who visited only urban areas.

JE Among U.S. Travelers

Before 1973, at least 300 cases of JE had been reported among U.S. military personnel or their family members (92,93,95,175–179). During 1973–1992, a total of 11 JE cases were reported among U.S. travelers and military personnel. During 1993–2017, after the first licensure of a JE vaccine in the United States in 1992, a total of 12 cases were reported among U.S. travelers, with a median of zero cases per year (range: 0–2) (10,143,144,165,173,174). On the basis of 12 reported cases during this 25-year period, and approximately 4–5 million U.S. citizen trips to Asia annually, the overall incidence of JE among U.S. travelers is estimated to be <1 case per million trips to Asia (180). Among the 12 cases, three (25%) were in children aged ≤11 years, and the remainder were in adults aged ≥17 years. Eight (67%) cases were in males. Six (50%) patients recovered, three (25%) survived but had sequelae, two (17%) died, and the outcome for one (8%) was unknown. Overall, four (33%) cases occurred in U.S. expatriates living in Asia and eight (67%) in tourists. Duration of travel ranged from 10 days to approximately 3 years, and for eight travelers (67%) was ≥1 month. On the basis of a 2007 study, approximately 20% of U.S. travelers to Asia travel for >30 days; therefore, approximately two thirds of U.S. traveler cases occurred among the smaller 20% of higher-risk, longer-term travelers (181). Among the four shorter-term travelers, three had traveled for 3 to <4 weeks, and one had traveled for 10 days. One shorter-term traveler spent most of the time in rural areas, two stayed in urban areas but took

at least one overnight trip to a rural area, and one had no exposure-related information.

The proportion of U.S. travelers who receive JE vaccine is unknown. However, studies suggest JE vaccination rates are low, even among higher-risk travelers. A 2007 survey of adult travelers on direct flights from the United States to Asia determined that 415 (25%) of 1,691 participants described itineraries for which JE vaccination should have been considered, including 330 (20%) who planned to spend ≥ 30 days in a JE-endemic country and another 85 (5%) shorter-term travelers who planned to spend at least 50% of their time in JE-endemic rural areas (181). Of these higher-risk travelers, 47 (11%) reported receiving at least 1 dose of JE vaccine. Among 164 unvaccinated higher-risk travelers who had visited a health care provider to prepare for their trip, 113 (69%) indicated that their health care provider had not offered or recommended JE vaccine. Results of another survey conducted among a group of U.S. clinical practices that provide pretravel health care indicated that 711 (9%) of 8,289 adults had an increased risk for JE on the basis of planned travel to one or more JE-endemic countries for ≥ 30 days during the JE virus transmission season with a visit to a rural area included in the itinerary (182). Among these 711 persons, 188 (26%) were vaccinated during the pretravel visit, and 11 (2%) had received JE vaccine within the previous 2 years; 512 (72%) were not administered JE vaccine. The main reasons noted for nonadministration included that JE vaccine was not indicated ($n = 282$; 55%), the patient declined ($n = 116$; 23%), or insufficient time to complete the vaccination series ($n = 85$; 17%) (182). In these two U.S. studies, 2%–4% of lower-risk travelers had been vaccinated (181,182).

Subclinical JE Virus Infection Among Travelers

Two studies have investigated the frequency of subclinical JE virus infection. Among 1,000 unvaccinated U.S. infantry soldiers deployed to Korea for at least 330 days during 2008–2011, predeployment and postdeployment serologic testing suggested one possible subclinical infection (183). In a study of 387 Australian adult travelers not vaccinated at their pretravel visit who traveled to Asia for a median of 21 days (range: 7–326 days) and had pretravel and posttravel serologic testing, no JE virus infections occurred; therefore, the risk for subclinical infection was zero per 10,000 traveler-days (95% confidence interval [CI] = 0–3.9) (184).

JE Vaccines

Four types of JE vaccines are manufactured and available in different countries, including a live attenuated vaccine, a live recombinant (chimeric) vaccine, inactivated mouse brain-derived vaccines, and inactivated Vero cell culture-derived

vaccines (3,7,185,186). JE-VC, manufactured as Ixiaro, is the only JE vaccine licensed and available in the United States. JE-MB, manufactured as JE-VAX, was previously available in the United States; production has ceased, and all doses expired in May 2011 (16).

Correlates of Protection

Because several effective JE vaccines are available in Asia, randomized, controlled efficacy trials to evaluate new JE vaccines would be logistically difficult and potentially unethical. JE-VC was licensed based on its ability to induce JE virus neutralizing antibodies, which is thought to be a reliable surrogate of efficacy (187,188). Observations from the 1930s indicated that laboratory workers who had been accidentally exposed to JE virus were protected from disease when they had measurable neutralizing antibodies (187). These observations were further supported by passive antibody transfer and active vaccination studies in animals using both licensed and experimental JE vaccines. Studies in mice indicated that passive transfer of neutralizing antibodies protected animals against JE virus challenge and established a dose-response relationship between antibody titer and level of protection (189–192). These studies also indicated that animals that were actively primed but had no detectable neutralizing antibodies against JE virus were protected from lethal challenge, demonstrating an effective anamnestic immune response (192). A more recent study indicated that hyperimmune ascitic fluid raised against two JE vaccines derived from genotype III JE virus strains (i.e., JE-MB derived from the Nakayama strain and a chimeric vaccine derived from the SA14-14-2 strain) protected mice against intracerebral challenge with JE virus strains of four genotypes. These data demonstrate that neutralizing antibodies provide protection against heterologous JE virus genotypes (193). In another study, mice were passively vaccinated with pooled sera with varying titers of neutralizing antibodies against JE virus from humans vaccinated with JE-VC. Mice were challenged 18 hours later with a lethal dose of either a genotype I (KE-093) or genotype III (SA14) JE virus strain (194). Mice with ex vivo neutralizing antibody titers of ≥ 10 had survival rates of 100% (10 of 10) and 90% (nine of 10) after challenge with the genotype I and III JE virus strains, respectively. In mice receiving lower titer sera, survival correlated with the neutralizing antibody titer of the immunizing sera. Mice actively vaccinated with varying doses of JE-VC and JE-MB also had dose-dependent protection against intraperitoneal challenge with the JE virus SA14 strain (194). Finally, in a study designed to develop a JE animal model in nonhuman primates, 16 rhesus macaques were given an intranasal challenge with a 90% effective dose

of JE virus (i.e., a dose that when administered via intranasal challenge would be expected to cause encephalitis in 90% of the animals), including four monkeys that were given four doses of an inactivated mouse brain–derived JE vaccine, eight monkeys immunized with one of two developmental poxvirus JE vaccines, and four JE virus–naïve control monkeys (195,196). The minimum neutralizing antibody titer required to protect the monkeys from lethal challenge was between 30 and 46. The higher titers required for protection in this study might have been caused by the high challenge dose used to develop the model.

PRNT is used to measure functional antibody that inactivates or neutralizes virus. A PRNT₅₀ titer is the reciprocal of the endpoint serum dilution that reduces the challenge virus plaque count by 50%. A WHO expert panel accepted a PRNT₅₀ titer of ≥ 10 as an immunologic correlate of protection against JE in humans (188). Although a correlate of protection has been defined, a vaccinated person whose neutralizing antibody titer has waned to a level of < 10 might still be protected because of persisting immunologic memory. Several studies have shown that vaccinated persons without measurable neutralizing antibodies can mount a rapid anamnestic response to infection (197–199). T cells likely also play a key role in clearing JE virus.

PRNT can be performed using various protocols, and the validity and comparability of PRNT results depend on detailed components of the selected assay (e.g., endpoint neutralization, incubation conditions, cell substrate, and target virus) (188,200). Although JE virus PRNTs are performed only at selected reference laboratories, careful attention must be paid to the characteristics and validation of a PRNT assay that is used to measure JE virus neutralizing antibody titers as a surrogate for efficacy.

JE-VC

Manufacture and Licensure

In March 2009, FDA approved JE-VC for use in persons aged ≥ 17 years; in May 2013, licensure was extended to include children aged ≥ 2 months (17). The booster dose was approved for persons aged ≥ 17 years in October 2010 and for children in April 2018.

JE-VC is an inactivated vaccine derived from the attenuated SA14-14-2 JE virus strain propagated in Vero cells (Box 1) (14,201,202). Each 0.5-mL dose contains 6 antigen units of purified, inactivated JE virus and approximately 250 μg aluminum hydroxide as an adjuvant.

Immunogenicity of JE-VC in Adults

Primary Series at 0 and 28 Days

No efficacy data exist for JE-VC. The vaccine was licensed on the basis of its ability to induce JE virus neutralizing antibodies as a surrogate for protection. The pivotal noninferiority immunogenicity study compared 2 doses of JE-VC given on days 0 and 28 to 3 doses of JE-MB given on days 0, 7, and 28 to adults aged ≥ 18 years in the United States, Austria, and Germany (203). Prevacination PRNT₅₀ titers were < 10 for all participants. In the per-protocol analysis, 352 (96%) of 365 JE-VC recipients developed a PRNT₅₀ titer ≥ 10 , compared with 347 (94%) of 370 JE-MB recipients at 28 days after the last dose (14). PRNT₅₀ titers were > 80 among 91% of the 361 JE-VC recipients for whom data were available. The difference in seroconversion rates was 2.6% (95% CI = -0.5%–6%), and noninferiority of JE-VC compared with JE-MB was established (14,203). The PRNT₅₀ geometric mean titer (GMT) for JE-VC recipients was 244 (95% CI = 216–274), compared with 102 (95% CI = 90–115) for JE-MB recipients. However, the target JE virus strain in the neutralizing antibody assay was SA14-14-2 (i.e., the JE virus strain used in JE-VC), whereas JE-MB is produced from the Nakayama JE virus strain. The GMT ratio was 2.3 (95% CI = 2.0–2.8), and noninferiority was again established.

The licensed vaccine schedule was derived in part from a study that compared 2 6- μg doses of vaccine administered 28 days apart to a single dose of either 6 μg or 12 μg (204). Twenty-eight days after receiving 1 dose of the standard 6- μg regimen, only 95 (41%) of 230 JE-VC recipients had seroconverted with a PRNT₅₀ titer ≥ 10 . Fifty-six days after receiving their first dose of vaccine, 110 (97%) of 113 participants who had received 2 doses had a PRNT₅₀ titer ≥ 10 , compared with 30 (26%) of 117 and 47 (41%) of 114 of those who received a single 6- μg or 12- μg dose, respectively; GMTs in the three groups were 218, 8, and 11, respectively. All of the 2-dose recipients who seroconverted had protective antibodies by 7 days after receiving the second dose of vaccine. In all other prelicensure and postlicensure randomized controlled trials and observational studies, $\geq 95\%$ of adults were seroprotected after receiving 2 doses of JE-VC administered 28 days apart, with the exception of one study that showed lower seroprotection rates among adults aged ≥ 64 years (Table 1) (201,203–209).

Adults Aged ≥ 65 Years

Prelicensure clinical trials did not include sufficient numbers of persons aged ≥ 65 years to allow an adequate assessment of immunogenicity in this age group. One immunogenicity study of JE-VC included 24 persons aged ≥ 65 years who received

BOX 1. Composition, storage, dose, and administration of inactivated Vero cell culture–derived Japanese encephalitis vaccine

Trade name: Ixiaro
JE virus strain: SA14-14-2
JE virus seed: Attenuated
Substrate: Vero cells
Adjuvant: Aluminum hydroxide
Stabilizer: None
Preservative: None
Final preparation:* Liquid
Storage: 35°F–46°F (2°C–8°C)
Presentation: Prefilled syringe
Route: Intramuscular

Dose and schedule, by age group:

- 2–35 months: 2 doses (0.25 mL each) administered on days 0 and 28[†]
- 3–17 years: 2 doses (0.5 mL each) administered on days 0 and 28
- 18–65 years: 2 doses (0.5 mL each) administered on days 0 and 7–28[§]
- >65 years: 2 doses (0.5 mL each) administered on days 0 and 28

Booster dose, by age group (if ongoing exposure or reexposure is expected):

- <3 years: 1 dose (0.25 mL) at ≥1 year after the second dose^{†,¶}
- ≥3 years: 1 dose (0.5 mL) at ≥1 year after the second dose[¶]

* The final preparation contains residues of protamine sulfate.

[†] To administer a 0.25-mL dose, expel and discard half of the volume from the 0.5-mL prefilled syringe.

[§] Studies showing the second dose can be administered as early as 7 days after the first dose only included adults aged 18–65 years.

[¶] No data are available on the response to a booster dose administered >2 years after the second dose.

the 2-dose primary series per protocol. At 28 days after the second dose, 23 (96%) persons had a seroprotective titer, and the GMT was 255 (14,203).

One postlicensure phase IV observational study was conducted to investigate immunogenicity of JE-VC in older adults (206). The median age was 69 years (range: 64–83 years). Forty-two days after the second dose of a 2-dose primary series, 128 (65%) of 197 persons were seroprotected and the GMT was 37. Both the seroprotection rate and GMT were substantially lower compared with results in the pivotal immunogenicity study of JE-VC in which study participants had a median age of 41 years (203). Seroprotection after the second dose was measured at 42 days in the study among older adults compared with 28 days in the pivotal immunogenicity study; however, that difference is unlikely to explain the results. In a subanalysis of 173 persons aged 65–74 years compared with 23 persons aged 75–83 years, the seroprotection rates and GMTs were similar in both of these groups. No data were gathered on seroprotection rates at >42 days after the second dose, or immunologic response to an additional dose or early booster dose of JE-VC.

Delayed Administration of the Second Dose of the Primary Series

In one study, persons who had previously received a single 6- μ g dose of JE-VC and had a PRNT₅₀ titer <10 at month 6 received a second 6- μ g dose at month 11 (14,210). At 28 days after the second dose, 99 (99%) of 100 persons were seroprotected, and the GMT was 504 (95% CI = 367–692). Compared with 2 doses administered at a 28-day interval, 2 doses administered at an interval of 11 months resulted in a similar rate of seroprotection and a higher GMT. At 13 months after dose 2 of the 0- and 11-month schedule, 85 (89%) of 96 participants were still seroprotected, and the GMT was 121 (95% CI = 87–168). Other than anecdotal reports of a small number of study participants who seroconverted when 2 doses were administered 23 months apart, no data are available on the immunogenicity of the primary series administered at an interval of >11 months.

Accelerated Primary Series in Adults Aged 18–65 years

A randomized, controlled trial in adults aged 18–65 years in Austria, Germany, and Switzerland investigated immunogenicity

after JE-VC administered in an accelerated primary schedule on days 0 and 7, given concomitantly with purified chick embryo cell rabies vaccine (14,205). In two comparison groups, JE-VC was administered according to a conventional 2-dose primary series on days 0 and 28, with or without rabies vaccine. Rabies vaccine was administered in the accelerated schedule group on days 0, 3, and 7 (an unlicensed regimen in the United States) and in the conventional group on days 0, 7, and 28. Twenty-eight days after the second JE-VC dose, 203 (99%) of 206 persons in the accelerated schedule group, 157 (100%) of 157 persons in the JE-VC conventional schedule with rabies vaccine group, and 49 (100%) of 49 persons in the JE-VC conventional schedule alone group were seroprotected (Table 2). The PRNT₅₀ GMT in the accelerated schedule group was 690 compared with 299 for the conventional JE-VC schedule with rabies vaccine and 337 for the conventional JE-VC schedule alone. At 10–12 months after the second dose, seroprotection rates were 94% for the accelerated schedule group and 86% and 88% for the other two groups (14,211). The GMT in the accelerated schedule group was 117, threefold higher than the GMTs of 39 in the other two groups. The reason for the higher GMT in the accelerated schedule group is unknown; no study participants reported vaccination with other flavivirus vaccines during the study period (211).

Data on a shorter schedule also are available from a phase II study of JE-VC that investigated alternate dosing schedules among adults aged 18–49 years (201). One study arm included persons who received JE-VC on a 0-, 14- and 28 -day schedule, and a blood sample was collected before vaccination on day 28. At 14 days after administration of the 0- and 14-day doses, 22 (96%) of 23 persons were seroprotected, and the GMT was 328 (95% CI = 189–570).

Adults with Preexisting Flavivirus Antibodies

A study that evaluated the effect of preexisting antibodies against tickborne encephalitis (TBE) virus, another flavivirus, determined that TBE virus antibodies enhanced the response to JE-VC after the first dose but had no effect after the 2-dose primary series (212). After 1 dose of JE-VC, 62 (77%) of 81 persons with preexisting TBE virus IgG antibodies developed protective antibodies against JE virus compared with 166 (49%) of 339 JE-VC recipients with no preexisting TBE virus antibodies. However, after the second dose of JE-VC, persons with and without TBE virus antibodies had similarly high rates of seroprotection against JE virus, with 78 (96%) of 81 and 310 (91%) of 339, respectively, with protective antibodies; this difference was not statistically significant ($p = 0.17$). JE virus PRNT₅₀ GMTs also were similar between the groups after 2 doses of JE-VC (207 and 187, respectively; $p = 0.56$).

Duration of Neutralizing Antibodies After JE-VC Primary Series

Three clinical trials provided data on persistence of protective neutralizing antibodies after a primary JE-VC series of 2 doses administered 28 days apart. In a study performed in central Europe (Austria, Germany, and Romania), seroprotection rates ranged from 95% at 6 months to 82% at 60 months after receiving the first dose of the 2-dose series (Table 3) (14,213,214). A study that used similar methods but was performed in western and northern Europe (Germany and Northern Ireland) found that among adults receiving 2 doses of JE-VC, seroprotection rates were 83% (96 of 116) at 6 months, 58% (67 of 116) at 12 months, and 48% (56 of 116) at 24 months after their first vaccination (165). In a third clinical trial, conducted in Austria and Germany, at 15 months after the first dose of the 2-dose JE-VC vaccination series, 69% (137 of 198) of participants had a protective neutralizing antibody titer (215).

To investigate possible reasons for the substantially different seroprotection rates at similar time points in the three studies, a subsequent analysis was conducted using participant data from the study conducted in central Europe (Austria, Germany, and Romania) and stratifying participants by TBE vaccination status (Table 4) (214,216). In the stratified analysis, seroprotection rates were lower at all time points from 6 to 60 months after the first dose of a 2-dose primary series in the group that had not received TBE vaccine compared with the group with persons who had received TBE vaccine before or during the study. In the United States, TBE vaccine is not available, and other flavivirus vaccines are not routinely administered with JE-VC; therefore, the immunologic response after JE-VC is likely to be most similar to the participants who did not receive the TBE vaccine.

Immunologic Response After a Booster Dose

Two clinical trials provided data on the response to a booster dose of JE-VC. In a study conducted in Austria and Germany, 198 adults aged ≥ 18 years who had received a 2-dose primary series of JE-VC were administered a booster dose 15 months after the first dose (215). The percentage of participants with a protective neutralizing antibody titer increased from 69% (137 of 198) before the booster dose to 100% (198 of 198) at 28 days after the booster dose, and a protective titer was found in 98% of persons at 6 months and 12 months after the booster dose (Table 5). The GMT before the booster was 23 and increased fortyfold to 900 at 28 days after the booster dose. At approximately 76 months after the booster dose, 64 (96%) of 67 participants still had a PRNT₅₀ titer ≥ 10 and the GMT was 148, indicating good seroprotection

rates for at least 6 years after a booster dose (217). GMTs at month 76 were not significantly different in persons with and without a history of TBE or yellow fever vaccination. Data from this study were used in a mathematical model to estimate that protection after the booster dose would last an average of 14 years (range: 2–25 years).

In a second study, a booster dose was administered to 40 persons who had received a 2-dose primary series but no longer had protective neutralizing antibody titers (210). The booster was administered at 11 months ($n = 16$) or 23 months ($n = 24$) after the first dose and resulted in protective titers in all persons. GMTs at 1 month after the booster increased to 676 (95% CI = 365–1,253) in the group administered the dose at 11 months and 2,496 (95% CI = 1,408–4,427) in those vaccinated at 23 months after the primary series. Among the 16 persons who received the booster dose at 11 months, all still had seroprotective titers 13 months later.

Use of JE-VC After Primary Vaccination with JE-MB

In a study among U.S. military personnel who had received at least 3 doses of JE-MB or were JE vaccine naïve, persons were vaccinated with 2 doses of JE-VC on days 0 and 28 and immunogenicity was assessed at 28 days after 1 dose in the previously JE-MB-vaccinated persons and 2 doses in the vaccine-naïve persons (207). The previously JE-vaccinated persons had received their last JE-MB dose a median of 2.9 years earlier (range: 1.8–10.2 years). In the per-protocol analysis, the seroprotection rate among previously vaccinated participants on day 28 after 1 dose of JE-VC was 100% (44 of 44) and in previously unvaccinated participants at 28 days after 2 doses was 93% (53 of 57). The GMT was significantly higher in previously vaccinated participants after 1 dose (GMT 315; 95% CI = 191–520) compared with the previously unvaccinated participants after 2 doses (GMT 79; 95% CI = 54–114). Among previously JE-vaccinated persons, the time since receiving their last dose did not significantly affect the neutralizing antibody titers achieved after 1 dose of JE-VC; however, only 12 (27%) participants had received their last dose of JE-MB ≥ 5 years before enrollment.

In another U.S. study using archived sera from military personnel, immunogenicity at 12–23 months after a single dose of JE-VC was assessed in adults previously vaccinated with at least 3 doses of JE-MB compared with JE vaccine-naïve adults vaccinated with a 2-dose JE-VC series (218). Persons with a history of JE-MB vaccination had received their last JE-MB dose a median of 2.9 years (range: 1 day–19 years) before the JE-VC dose and had received a median of three JE-MB doses. At 12–23 months, seroprotection rates were 94% (235 of 250) in previously JE-MB-vaccinated personnel and 54% (135 of 250) in previously unvaccinated personnel.

The GMT of 75 (95% CI = 63–90) in the previously JE-MB-vaccinated personnel was significantly higher than the GMT of 12 (95% CI = 11–14) in the previously unvaccinated personnel.

An observational study was conducted at travel clinics in Scandinavia among adults planning travel to a JE-endemic area (208). One study cohort included participants who had received 2 or 3 doses of JE-MB and were vaccinated with 1 dose of JE-VC; the comparison group included JE vaccine-naïve persons who were vaccinated with 2 doses of JE-VC on days 0 and 28. Among the previously vaccinated persons who received 1 dose of JE-VC, their last JE-MB dose was a median of 5.2 years earlier (range: 1–21 years). At 4–8 weeks after the JE-VC dose, 98% (41 of 42) were seroprotected with a GMT of 504. Among unvaccinated persons, 4–8 weeks after two JE-VC doses, 97% (30 of 31) were seroprotected with a GMT of 499. In a follow-up study investigating duration of protection, only 47% of persons from the original study cohorts were available (219). A mean of 2.1 years after the final JE-VC dose, the seroprotection rate among previously JE-MB-vaccinated persons who had received 1 dose of JE-VC was 100% (18 of 18) and among previously unvaccinated persons who had received 2 doses of JE-VC was 93% (14 of 15).

The results of these studies indicate that among persons who previously received JE-MB vaccine, a single dose of JE-VC results in seroprotection rates and GMTs at approximately 4 weeks that are noninferior to those in unvaccinated persons who receive a standard 2-dose JE-VC series. At 12–23 months, the immunological response after 1 JE-VC dose in adults previously vaccinated with at least 3 doses of JE-MB remains noninferior to the response in JE vaccine-naïve adults vaccinated with the 2-dose primary series of JE-VC.

Concomitant Administration of JE-VC with Other Vaccines

JE-VC with Hepatitis A Vaccine

A clinical trial in which the first dose of JE-VC was administered concomitantly with hepatitis A vaccine indicated no interference with the immune response to JE-VC or hepatitis A vaccine (209). Among the 58 persons who received both JE-VC and hepatitis A vaccine in the per-protocol analysis, all had protective neutralizing antibodies compared with 98% (57 of 58) of persons who received JE-VC alone. GMTs also were similar at 203 (95% CI = 154–261) and 192 (95% CI = 148–250), respectively. In addition, persons receiving JE-VC and hepatitis A vaccine had similar seroconversion rates for antihepatitis A virus (anti-HAV) antibody (100%, 58 of 58) compared with persons receiving hepatitis A vaccine alone (96%, 50 of 52) and HAV antibody

GMTs were similar (150 and 124, respectively). However, some differences were noted between men and women in the levels of anti-HAV antibody achieved, and both seroconversion rates and antibody titers varied depending on which anti-HAV assay was used; whether these observations have any clinical significance is not known.

JE-VC with Rabies Vaccine

A randomized trial evaluated immunologic responses to JE-VC and a purified chick embryo cell culture rabies vaccine when vaccines were administered alone or concomitantly to adults aged 18–65 years (14,205,220). JE-VC was administered in a 2-dose schedule on days 0 and 28 and rabies vaccine in a 3-dose schedule on days 0, 7, and 28. Twenty-eight days after the second JE-VC dose, all persons in the concomitant administration group (157/157) and in the group administered JE-VC alone (49 of 49) had seroprotective neutralizing antibody titers against JE virus and GMTs were similar at 299 and 337, respectively (Table 2). At 12 months after the first JE-VC dose, JE seroprotection rates were similar at 86% (132 of 154) and 88% (42 of 48), and the GMT for both groups was 39 (211). The percentage of persons with protective neutralizing antibody concentrations against rabies virus (i.e., ≥ 0.5 IU/mL) at 28 days after the third rabies vaccine dose was 100% (157 of 157) in the concomitant administration group and 99% (203 of 204) in the group administered rabies vaccine alone (220). Noninferiority of the immunologic responses to JE-VC and rabies vaccine was established for concomitant administration compared with separate administration of either vaccine.

JE-VC and Rabies Vaccine with Meningococcal Vaccine

Another study was conducted in which JE-VC and purified chick embryo cell rabies vaccine were administered concomitantly, with or without a quadrivalent meningococcal conjugate vaccine, to adults aged 18–60 years (221). Two additional groups received rabies vaccine alone or meningococcal vaccine alone. JE-VC was administered according to a 0- and 28-day schedule, rabies vaccine on a 0-, 7-, and 28-day schedule, and meningococcal vaccine as a single dose on day 0. In the per-protocol analysis, at 28 days after the final doses of JE-VC and rabies vaccine, 95 (98%) of 97 adults who received all three vaccines had seroprotective titers against JE virus, compared with 95 (99%) of 96 persons who received JE-VC and rabies vaccine. GMTs also were similar at 165 (95% CI = 136–199) and 183 (95% CI = 151–221), respectively. All persons in the three groups that received rabies vaccine developed protective rabies virus antibody concentrations of ≥ 0.5 IU/mL. Measurement of the response to meningococcal vaccine was at 56 days in persons who received

the three vaccines concomitantly and at 28 days in the group that received meningococcal vaccine alone. No significant differences were found in the percentage of participants who achieved human serum bactericidal assay antibody titers $\geq 1:8$ against meningococcal serogroups A, C, W, or Y; however, the GMT against each serogroup was higher when meningococcal vaccine was administered alone. Interpretation of these results is complicated by the different time points of the blood draws for the two groups.

Immunogenicity of JE-VC Against Different JE Virus Genotypes

JE-VC is derived from the genotype III SA14-14-2 JE virus. To assess cross-protection provided against different JE virus genotypes, a study was conducted among European travelers who were administered a 2-dose primary JE-VC series and evaluated for the presence of neutralizing antibodies against JE virus strains representing genotypes I, II, III, and IV (222). At 4–8 weeks after the primary series, among 29 persons vaccinated with JE-VC, $\geq 93\%$ had PRNT titers ≥ 10 against each of the virus strains. GMTs ranged from 55 for the genotype I strain to 811 for the genotype II strain.

Immunogenicity of JE-VC in Children

Primary Series

The pivotal pediatric clinical trial of JE-VC was conducted among children aged 2 months–17 years in the Philippines (14,223). Among children randomly assigned to receive 2 age-appropriate doses of JE-VC, 384 (99.7%) of 385 were seroprotected at 28 days after the second dose (95% CI = 96%–100%) (Table 6). Seroprotection rates were similar in children who received the 0.25-mL and 0.5-mL doses.

In a randomized, controlled trial conducted in India among children aged 1–2 years, 22 (96%) of 23 children (95% CI = 87%–100%) were seroprotected at 28 days after receiving 2 0.25-mL doses of JE-VC compared with 10 (91%) of 11 (95% CI = 74%–100%) children who received 3 doses of an inactivated mouse brain-derived JE vaccine (14,224). GMTs were 201 (95% CI = 106–380) and 230 (95% CI = 68–784), respectively. In an observational study of 62 children from countries without endemic JE, all had protective neutralizing antibodies 28 days after the second dose of JE-VC (14,225).

Duration of Neutralizing Antibodies After JE-VC Primary Series

In the pediatric clinical trial of JE-VC in the Philippines, 6 months after completing the primary series, 134 (88%) of 152

children aged 2 months–2 years (95% CI = 82%–92%) and 224 (95%) of 237 children aged 3–17 years (95% CI = 91%–97%) had protective neutralizing antibodies (14,223). A subset of 300 children were enrolled in a follow-up study and were randomly assigned to groups that received or did not receive a booster dose (226). Among the 150 children who did not receive a booster dose, 90% (134 of 149) were seroprotected 11 months after the second dose of the 2-dose primary series, 89% (130 of 146) at 23 months, and 90% (128 of 142) at 35 months (14,226).

In the observational study of children from countries without endemic JE, 91% (31 of 34) children had protective neutralizing antibodies 6 months after the second dose of the 2-dose primary series (14,225). Among a subset enrolled in a follow-up study, seroprotection rates were 89% (17 of 19) at 11 months, 91% (21 of 23) at 23 months, and 89% (17 of 19) at 35 months (14).

Immunologic Response After a Booster Dose

Among 300 children enrolled in a follow-up to the pediatric immunogenicity study conducted in the Philippines, 150 were randomly assigned to receive an age-appropriate booster dose 11 months after the second dose of the primary series. Among these children, 94% (139 of 148) had a protective neutralizing antibody titer before the booster dose, and 100% were seroprotected 1 month, 12 months, and 24 months after the booster dose (Table 7) (14,226). The GMT was 53 before the booster dose and increased approximately fortyfold to 2,067 at 1 month after the booster dose. GMTs remained high at 428 at 12 months and 350 at 24 months after the booster dose. Data also were used in a mathematical model to estimate the duration of protection after the booster dose and suggested a median duration of 9 years (range: <5 years to ≥30 years) (227).

Local and Systemic Adverse Events with JE-VC

JE-VC was studied in approximately 5,000 adults in prelicensure clinical trials (15). Local and systemic adverse events caused by JE-VC were similar to those reported for JE-MB or placebo adjuvant alone (228,229). Subsequent postlicensure studies included children and older adults, and no safety concerns were identified (206,230). Since the vaccine's licensure, approximately 1 million doses have been distributed and no patterns of serious hypersensitivity, neurologic, or other serious adverse events considered to be vaccine related have been identified (231,232).

Among adults, the most common local reactions after a dose of JE-VC are pain and tenderness, and the most common systemic reactions are headache and myalgia (14,228).

Among children, the most frequently reported local reactions are redness in children aged 2 months to <3 years and pain and tenderness in children aged 3–17 years, and the most commonly reported systemic reaction is fever (14,230).

Adverse Events with JE-VC Compared with Placebo Adjuvant

The pivotal safety study comparing 1,993 adults aged ≥18 years randomly assigned to receive 2 doses of JE-VC and 657 persons assigned to receive 2 doses of placebo adjuvant (phosphate buffered saline with 0.1% aluminum hydroxide) indicated similar reactogenicity and adverse events, including medically attended and serious adverse events (228). The most common local reactions after administration of dose 1 or 2 of JE-VC were pain (28% and 18% after doses 1 and 2, respectively) and tenderness (29% and 23%), and the most common systemic reactions were headache (22% and 13%) and myalgia (13% and 6%) (14,228). Two patients had urticaria (228). The first patient had a rash localized on the thighs, which occurred 6 days after the second placebo vaccination. The second patient had generalized urticaria of the face, chest, arms, and abdomen, which occurred 8 days after the second dose of JE-VC and was described as moderate; angioedema was not observed. The patient was treated with cetirizine hydrochloride, and the rash resolved after 3 days (228). A total of 17 persons, 12 (0.6%) in the JE-VC group and five (0.8%) in the placebo group, terminated the study prematurely because of adverse events (228). In the JE-VC group, two of these events (gastroenteritis and rash) were considered severe, and eight of them (headache [two events], influenza-like illness, allergic dermatitis, injection site pain, nausea, fatigue, and rash) were considered to be at least possibly related to the study treatment. No serious neurologic events were identified.

Adverse Events with JE-VC Compared with JE-MB

In the noninferiority immunogenicity trial among adults aged ≥18 years, the overall frequency of adverse events reported after JE-VC vaccination (n = 428 persons) was similar to that reported by those receiving JE-MB (n = 435 persons) (203). Severe redness, swelling, tenderness, or pain at the injection site were each reported by ≤1% of JE-VC recipients (Table 8). Reported systemic adverse events after JE-VC vaccination generally were mild; the most commonly reported adverse events in the 7 days after each dose were headache (26%), myalgia (21%), influenza-like illness (13%), and fatigue (13%). One serious adverse event was reported in the JE-VC group; a man aged 50 years had a nonfatal myocardial infarction 3 weeks after the second vaccination. The event was considered by the investigator as unlikely to be related to the study vaccine.

Pooled Safety Data

A pooled analysis of 6-month safety data from seven prelicensure studies among adults aged ≥ 18 years included 3,558 persons administered at least 1 dose JE-VC, 435 persons administered at least 1 dose of JE-MB, and 657 placebo adjuvant recipients (229). Local injection site reactions within 7 days of dose 1 were reported by 48% of the JE-VC persons, 46% of the JE-MB recipients, and 48% of the placebo adjuvant recipients. However, severe local reactions after dose 1 were more frequent after JE-MB (6%) compared with JE-VC (3%) and placebo adjuvant recipients (2%) ($p < 0.01$). Systemic adverse events were reported with similar frequency among persons who received JE-VC (64%), JE-MB (64%), or placebo (61%). Serious adverse events were reported by 1% of the persons in the JE-VC group. Serious allergic reactions did not occur in any of the study groups. Systemic adverse events were reported by a lower percentage of participants after the second dose compared with the first dose.

Adverse Events with JE-VC Administered in an Accelerated Primary Series

In the study of adults administered JE-VC in an accelerated schedule concomitantly with a purified chick embryo cell rabies vaccine ($n = 217$), JE-VC in the standard schedule concomitantly with rabies vaccine ($n = 167$), or JE-VC in the standard schedule alone ($n = 56$), local adverse events were reported in 74%, 75%, and 63% of persons, respectively (205). Systemic adverse events were reported by 66%, 60%, and 54% of persons in the three groups, respectively. Overall, rates of local and systemic adverse events were similar when JE-VC was administered in an accelerated or standard schedule.

Adverse Events with JE-VC in Adults Aged ≥ 65 Years

In adults aged ≥ 65 years vaccinated with a 2-dose primary series of JE-VC, in the 7 days after each dose the most common local reaction was tenderness (26%) and the most common systemic reaction was headache (18%) (206). Serious or medically attended adverse events were reported in 38 (19%) of 200 persons by day 42 after the second dose, but none were considered by study investigators to be causally related to vaccination.

Adverse Events in Children

In an open-label trial in the Philippines, 195 infants aged 2–11 months were randomly assigned to receive JE-VC ($n = 131$) or 7-valent pneumococcal conjugate vaccine ($n = 64$). An additional 1,674 children aged 1–17 years were randomly assigned to receive JE-VC ($n = 1,280$) or hepatitis A vaccine ($n = 394$) (14,230). The incidences of local, systemic,

medically attended, and serious adverse events were similar between children who received JE-VC or the comparison vaccines. Adverse events were most frequent in children aged 2–11 months. The most frequently reported local reactions were redness in children aged 2 months to < 3 years and pain and tenderness in children aged 3–17 years. Overall, 9% (122 of 1,411) of JE-VC recipients had fever ($\geq 100.4^\circ\text{F}$ [38.0°C]) within 7 days after the first dose, and 6% (84 of 1,405) had fever within 7 days after the second dose (17). Rates of fever were higher in children aged < 3 years compared with older children. Within 1 month after either dose, four ($< 1\%$) recipients had urticaria or hypersensitivity reactions, and five ($< 1\%$) had neurologic adverse events, including febrile seizures ($n = 3$), drooling ($n = 1$), and dizziness ($n = 1$); all rates were similar to rates for recipients of the comparison vaccines. Solicited local and systemic adverse events were more frequent after dose 1 than dose 2. In children aged 2–11 months, 46% reported events after dose 1 and 28% after dose 2, and in those aged 1–17 years, 32% and 18% reported events after dose 1 and 2, respectively. Among the 1,411 children who received JE-VC, 23 (2%) reported a serious adverse event within 7 months of the first dose. The most common serious adverse events were pneumonia ($n = 6$) and febrile seizures ($n = 5$). Only three serious adverse events were reported within 2 weeks after a dose of JE-VC, including one report each of a febrile convulsion, cellulitis, and gastroenteritis. One death from disseminated intravascular coagulation after suspected bacterial meningitis was reported in a boy aged 12 years at 4 months after receipt of the second dose of JE-VC and was considered unrelated to vaccination by the investigator and the trial's Data Safety Monitoring Board. No other neurologic or hypersensitivity events were reported as serious adverse events.

Among 48 children aged 1–2 years who were randomly assigned to receive JE-VC in a trial in India, five (10%) cases of injection site tenderness and one (2%) case of fever within 7 days after either dose were reported (224). The only unsolicited adverse events were one report each of a skin lesion and a skin rash. No serious adverse events or deaths were reported.

In an observational study of children aged 2 months–17 years from countries without endemic JE, adverse events were evaluated among 12 children who received a 0.25-mL dose and 88 children who received a 0.5-mL dose (14,225). Among children who received the 0.25-mL dose, the most common solicited adverse reactions within 7 days after either JE-VC dose were injection-site redness ($n = 3$, 25%) and diarrhea ($n = 2$, 17%). Among children who received the 0.5-mL dose, the most common solicited adverse reactions were injection-site tenderness ($n = 44$, 50%) and muscle pain ($n = 27$, 31%). Three serious adverse events were reported. None occurred

within 28 days of either dose of JE-VC. One child each had diabetes mellitus (3 months after dose 2), dizziness (4 months after dose 2), and intentional self-injury.

Adverse Events After a JE-VC Booster Dose in Adults and Children

Among adults aged ≥ 18 years who received a JE-VC booster dose at 15 months after the first dose of a 2-dose primary series, during the 7 days after the booster dose the most frequent local reactions were tenderness in 19% (37 of 193) and pain in 13% (25 of 195), and the most commonly reported systemic reactions were headache in 11% (21 of 194) and fatigue in 10% (18 of 188) (16,215,233). No serious adverse events were reported during the 28 days after the booster dose.

In a similar trial conducted among children, within the 7 days after the booster dose, 12 (8%) of 148 children had a local adverse event, and 21 (14%) had a systemic adverse event (226). Two children experienced serious adverse events within 1 month after the booster dose. One child had an abscess in the lumbar area 1 day after vaccination, and one had dengue fever approximately 4 weeks after the booster dose.

Adverse Events with Concomitant Administration of JE-VC and Hepatitis A or Rabies Vaccines

Persons in a clinical trial who received the first dose of JE-VC administered concomitantly with hepatitis A vaccine were more likely to report pain, redness, and swelling than persons who received either vaccine alone (209). No other differences were reported in safety or reactogenicity with concomitant administration of JE-VC and hepatitis A vaccine compared with administration of each vaccine alone.

Adults receiving JE-VC and rabies vaccine reported more local and systemic adverse events when the vaccines were coadministered compared with administered alone (205,220). Among 166 persons who received the vaccines concomitantly, local reactions were reported in 125 (75%) and systemic reactions in 100 (60%). Among 56 persons who received JE-VC alone, 35 (63%) reported local reactions and 30 (54%) reported systemic reactions.

Postlicensure JE-VC Surveillance

Two reviews of vaccine safety data from the U.S. Vaccine Adverse Event Reporting System (VAERS) have occurred since vaccine licensure, covering a total of 7 years during May 2009–April 2016, when >1 million doses of JE-VC were distributed (231,232). VAERS is the national passive surveillance system for monitoring adverse events after vaccination with reports submitted by health care providers, vaccine recipients, and vaccine manufacturers. The overall rates of adverse events in the two analyses were similar, with a rate of 15.2 adverse events per

100,000 doses distributed in the first analysis and 14.8 adverse events per 100,000 doses during the second period. The rates were similar to or lower than the 15.0 and 23.7 adverse events per 100,000 doses distributed previously reported to VAERS for JE-MB (234,235). In the two analyses of JE-VC data reported to VAERS, the rates of serious adverse events defined according to the FDA definition were 1.8 and 1.1 per 100,000 doses distributed; among the 14 serious event reports, 11 (79%) were reports in which JE-VC was administered with one or more other vaccines (236). Hypersensitivity events were reported at rates of 3.0 and 4.4 per 100,000 doses distributed, and 56% (20 of 36) occurred after concomitant administration of JE-VC with other vaccines. Neurologic events were reported at rates of 2.2 and 1.2 events per 100,000 doses distributed. The neurologic adverse event reports included four reports of seizures after vaccination, and all occurred after administration of JE-VC with other vaccines. VAERS data cannot generally be used to determine causality, especially among persons who receive multiple vaccines. However, the majority of reports in both analyses were not serious, and no unexpectedly high reporting rates for specific events were identified.

In a postmarketing adverse event surveillance study conducted among U.S. military personnel, rates of hypersensitivity and neurologic reactions were much higher, reflecting the different study methods (237). An active surveillance approach was used, events were identified using *International Classification of Diseases, Ninth Revision*, codes, and a retrospective review of medical records was conducted. However, complete descriptions of events often were lacking, preventing clarification of the nature of some events. In addition, the assessment was conducted among military personnel who sometimes received multiple other vaccines with JE-VC, including reactogenic vaccines.

Vaccination of Pregnant or Breastfeeding Women

No controlled studies have assessed the safety, immunogenicity, or efficacy of JE-VC in pregnant women. Preclinical studies of JE-VC in pregnant rats did not show evidence of harm to the fetus (14). No studies have investigated the safety or immunogenicity of JE-VC in breastfeeding women, and no data are available on whether JE-VC is excreted in human milk. ACIP general guidelines for best vaccination practices indicate inactivated vaccines administered to breastfeeding women do not affect the safety of breastfeeding for women or their infants (238).

Cost-Effectiveness of JE Vaccines

Several studies have demonstrated that JE vaccination among children in JE-endemic countries is cost-effective or cost-saving compared with no vaccination (239–241). Because of the substantially lower risk for disease among U.S. travelers and use of a much higher cost vaccine than those used for routine vaccination programs in Asia, JE vaccination for travelers would not be expected to be cost-effective. However, cost-effectiveness is less relevant for travel vaccines that usually are paid for by the travelers themselves and are not covered by most insurance plans or the Vaccines for Children program.

One comparative analysis compared strategies for JE vaccination for U.S. travelers to Asia among three groups (242). Group 1 included higher-risk travelers who planned to spend ≥ 1 month in JE-endemic areas, group 2 included travelers who would spend < 1 month in JE-endemic areas with at least 20% of their time participating in outdoor activities in rural areas, and group 3 included the remainder of shorter-term and lower-risk U.S. travelers to Asia. An analytic horizon of 6 years was used, although productivity losses were evaluated over average life expectancy. To prevent one JE case, the number of travelers who would need to be vaccinated was 0.7 million, 1.6 million, and 9.8 million in groups 1, 2, and 3, respectively. The cost to prevent one JE case from a societal perspective was approximately \$0.6 billion, \$1.3 billion, and \$7.9 billion for each group. The variable with the greatest influence on the cost-effectiveness of vaccination was disease incidence among travelers, and a sensitivity analysis was conducted increasing baseline incidence 100 times. Using this higher incidence, in groups 1, 2, and 3 the numbers of travelers needed to be vaccinated to prevent a case were 7,000, 16,000, and 98,000, and the cost per case averted was \$5 million, \$12 million, and \$78 million, respectively. Although the cost per case averted was high for all groups of travelers, this comparative analysis supported focusing on vaccination of travelers at increased risk for disease compared with those at lower risk.

Recommendations for the Prevention of JE Among U.S. Travelers

JE is a very low-risk disease for most U.S. travelers to JE-endemic countries. However, some travelers are at increased risk for infection on the basis of their planned itinerary. Factors that increase the risk for JE virus exposure include 1) longer duration of travel; 2) travel during the JE virus transmission season; 3) spending time in rural areas; 4) participating in extensive outdoor activities; and 5) staying in accommodations without air conditioning, screens, or bed nets (Box 2).

Health care providers should assess each traveler's risk for mosquito exposure and JE virus infection on the basis of their planned itinerary and discuss ways to reduce their risk (Figure 3). All travelers to JE-endemic countries should be advised to take precautions to avoid mosquito bites to reduce the risk for JE and other vectorborne diseases. These precautions include using insect repellent, permethrin-impregnated clothing, and bed nets and staying in accommodations with screened or air-conditioned rooms.

For some persons who might be at increased risk for JE based on travel duration, season, location, activities, and accommodations, JE vaccine can further reduce the risk for infection. The decision whether to vaccinate should be individualized and consider the 1) risks related to the specific travel itinerary, 2) likelihood of future travel to JE-endemic countries, 3) high morbidity and mortality of JE, 4) availability of an effective vaccine, 5) possibility but low probability of serious adverse events after vaccination, and 6) traveler's personal perception and tolerance of risk.

JE vaccine is recommended for persons moving to a JE-endemic country to take up residence, longer-term (e.g., ≥ 1 month) travelers to JE-endemic areas, and frequent travelers to JE-endemic areas. JE vaccine also should be considered for shorter-term (e.g., < 1 month) travelers with an increased risk for JE based on planned travel duration, season, location, activities, and accommodations (Box 2). Vaccination also should be considered for travelers to JE-endemic areas who are uncertain of specific duration of travel, destinations, or activities. JE vaccine is not recommended for travelers with very low-risk itineraries, such as shorter-term travel limited to urban areas or travel that occurs outside of a well-defined JE virus transmission season.

Recommendations for the Prevention of JE Among Laboratory Workers

Work with JE virus is primarily restricted to biosafety level 3 (BSL-3) facilities and practices; however, the attenuated SA14-14-2 JE vaccine virus can be handled at BSL-2 (243). In a laboratory setting, JE virus might be transmitted through accidental percutaneous, or theoretically, mucosal or inhalational exposures. Vaccine-induced immunity presumably protects against exposure through a percutaneous route. Exposure to aerosolized JE virus, particularly high concentrations that might occur during viral purification, might lead to infection through mucous membranes or through the olfactory epithelium directly into the central nervous system. Whether vaccination provides protection after such exposures is unknown.

BOX 2. Factors that increase risk for Japanese encephalitis among travelers**Duration**

- Highest incidence of disease has been reported among longer-term travelers.
- Although no specific duration of travel puts a traveler at risk for JE, longer-term travel increases the likelihood that a traveler might be exposed to an infected mosquito.
- Longer-term travel includes cumulative periods in JE-endemic areas; this includes frequent travelers and persons living in urban areas who are likely to visit higher-risk rural areas.

Season

- JE virus transmission occurs seasonally in some areas and year-round in others.
- Information on expected JE virus transmission by country is available from the *Yellow Book* on the CDC website (<https://wwwnc.cdc.gov/travel/yellowbook/2018/infectious-diseases-related-to-travel/japanese-encephalitis>). These data should be interpreted cautiously because JE virus transmission varies within countries and from year to year.

Location

- The highest risk occurs from mosquito exposure in rural or agricultural areas.
- Mosquitoes that transmit JE virus typically breed in flooded rice fields, marshes, and other stagnant collections of water.
- Some cases have been reported among travelers to coastal areas or resorts located in or adjacent to rural or rice growing areas.
- JE can occur in large, focal outbreaks indicating extensive active JE virus transmission in those areas.

Activities

- The mosquitoes that transmit JE virus feed most often in the outdoors, particularly from sunset through dawn; therefore, examples of activities that increase risk include the following:
 - Outdoor recreational activities such as camping, hiking, trekking, biking, rafting, fishing, hunting, or farming.
 - Spending substantial time outdoors, especially during the evening or night.

Accommodations

- Accommodations without air conditioning, screens, or bed nets increase risk for mosquito exposure.

Vaccination is recommended for all laboratory workers with a potential for exposure to JE viruses other than SA14-14-2 JE vaccine virus. Vaccination generally is not required for those who work only with SA14-14-2 JE virus; however, for those working with SA14-14-2 virus at high concentrations or volumes, or passaging virus, individual risk assessments with consideration of biosafety level and vaccination should be undertaken by a local biosafety committee. Vaccination is not required for workers handling routine clinical samples.

Administration of JE Vaccine

Vaccine Composition, Presentation, and Storage

Each 0.5-mL dose of JE-VC contains 250 μg aluminum hydroxide as an adjuvant. The finished product does not include gelatin stabilizers, antibiotics, or thimerosal. JE-VC is supplied in a 0.5-mL prefilled glass syringe with a plunger stopper (chlorobutyl elastomer, with no natural latex rubber). The vaccine should be stored at 35°F–46°F (2°C–8°C) and should not be frozen. The vaccine should be protected from light.

Dosage, Schedule, and Administration

Primary Vaccination Series

The vaccination dose and primary schedule for JE-VC vary by age (Box 1).

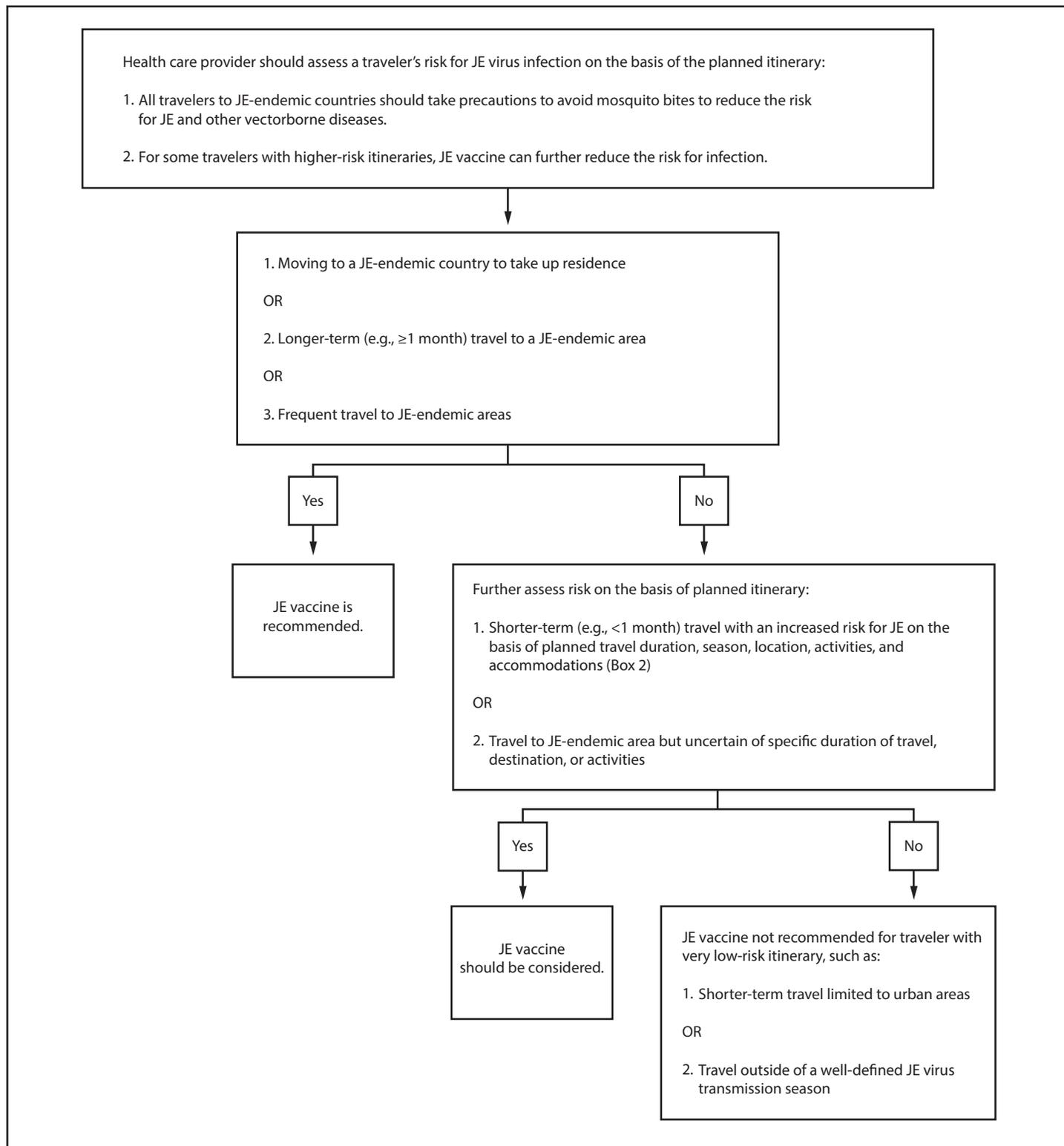
- 2–35 months: 2 doses (0.25 mL each) administered intramuscularly (IM) on days 0 and 28.
- 3–17 years: 2 doses (0.5 mL each) administered IM on days 0 and 28.
- 18–65 years: 2 doses (0.5 mL each) administered IM on days 0 and 7–28; this is the only age group for which an accelerated schedule is approved.
- >65 years: 2 doses (0.5 mL each) administered IM on days 0 and 28.

For all age groups, the 2-dose series should be completed at least 1 week before potential exposure to JE virus.

Booster Dose

For adults and children, a booster dose (i.e., third dose) should be given ≥ 1 year after completion of the primary JE-VC series if ongoing exposure or reexposure to JE virus is expected. The booster dose for children aged <3 years

FIGURE 3. Vaccine recommendations for U.S. travelers to areas with endemic Japanese encephalitis



Abbreviation: JE = Japanese encephalitis.

is 0.25 mL and for adults and children aged ≥ 3 years is 0.5 mL. No data are available on the response to a booster dose administered >2 years after the primary series. Clinical trial data show high rates of seroprotection for at least 6 years after a booster dose (217); no longer-term study data are available. No U.S. recommendations exist on the need for subsequent booster doses.

Vaccine Preparation

During storage, the vaccine might appear as a clear liquid with a white precipitate. Before administration, shake the syringe well to obtain a white, opaque, homogeneous suspension. To administer a 0.25-mL dose, expel and discard half of the volume from the 0.5-mL prefilled syringe by pushing the plunger stopper up to the edge of the red line on the syringe barrel before injection. See the prescribing information for additional information on preparing the 0.25-mL dose (14).

Simultaneous Administration of Other Vaccines or Drugs

A clinical trial in which the first dose of JE-VC was administered concomitantly with hepatitis A vaccine indicated no interference with the immune response to JE-VC or hepatitis A vaccine (209). Similarly, noninferiority of the immunological responses to JE-VC and purified chick embryo cell culture rabies vaccine was established for concomitant administration of the two vaccines compared with separate administration of either vaccine (205,220). If JE-VC and other vaccines are administered concomitantly, they should be administered with separate syringes and at different anatomical sites (i.e., >1 inch apart if possible).

Contraindications and Precaution for the Use of JE Vaccine

Allergy to Vaccine Components

A severe allergic reaction (e.g., anaphylaxis) after a previous dose of JE-VC, any other JE vaccine, or any component of JE-VC is a contraindication to administration of a subsequent dose. JE-VC contains protamine sulfate, a compound known to cause hypersensitivity reactions in some persons (14).

Pregnancy

Pregnancy is a precaution for the use of JE-VC. Vaccination with JE vaccine usually should be deferred because of a theoretical risk for the developing fetus. However, pregnant

women who must travel to an area in which risk for JE is high should be vaccinated if the benefits outweigh the risks of vaccination to the mother and developing fetus.

Special Populations

Infants aged <2 months: Safety and effectiveness of JE-VC have not been established for infants aged <2 months.

Adults aged ≥ 65 years: In a postlicensure observational study conducted among older adults, both the seroprotection rate and GMT were substantially lower after the primary JE-VC series compared with rates in younger persons. However, no data are available on the safety or immunogenicity of an additional dose or early booster dose of JE-VC for adults aged ≥ 65 years.

Breastfeeding women: Breastfeeding is not a contraindication or precaution to vaccination with JE-VC.

Persons with altered immune states: No data exist on the use of JE-VC in immunocompromised persons or patients receiving immunosuppressive therapies; however, these persons might have a diminished response to JE-VC.

Reporting of Vaccine Adverse Events

Surveillance for adverse events associated with administration of JE vaccine is important. Even if a causal relation to vaccination is not certain, all clinically significant adverse events should be reported to the VAERS (<https://vaers.hhs.gov> or 800-822-7967).

Future Research on JE-VC

Additional studies of JE-VC would be useful to evaluate persistence of protective immunity beyond 6 years after a booster dose, response to a booster dose administered >2 years after the primary JE-VC series, and response to a booster dose in adults aged >65 years.

Additional Information

Additional information about JE is available from CDC at <https://www.cdc.gov/japaneseencephalitis> and in the CDC *Yellow Book* (4). Additional licensure information for JE-VC is available from the U.S. Food and Drug Administration (<https://www.fda.gov/vaccines-blood-biologics/vaccines/ixiaro>).

Acknowledgments

Lorry Rubin and Cynthia Pellegrini previously were ACIP members on the ACIP JE Vaccine Work Group. Amanda Cohn and Jessica MacNeil provided advice in development of the JE vaccine recommendations and this document. Ann Powers provided advice in development of the JE vaccine recommendations for laboratory workers.

Conflicts of Interest

No conflicts of interest were disclosed.

References

- Fischer M, Hills S, Staples E, Johnson B, Yaich M, Solomon T. Japanese encephalitis prevention and control: advances, challenges, and new initiatives. In: Scheld WM, Hammer SM, Hughes JM, eds. *Emerging infections 8*. Washington, DC: ASM Press; 2008:93–124.
- Heffelfinger JD, Li X, Batmunkh N, et al. Japanese encephalitis surveillance and immunization—Asia and Western Pacific regions, 2016. *MMWR Morb Mortal Wkly Rep* 2017;66:579–83. <https://doi.org/10.15585/mmwr.mm6622a3>
- Halstead SB, Hills SL, Dubischar K. Japanese encephalitis vaccines. In: Plotkin SA, Orenstein WA, Offit PA, Edwards KM, eds. *Plotkin's vaccines*, 7th ed. Philadelphia, PA: Elsevier; 2017:511–48.
- Hills SL, Lindsey NP, Fischer M. Japanese encephalitis. In: CDC. *CDC Yellow Book 2020: health information for international travel*. New York, NY: Oxford University Press; 2019:248–57.
- Campbell GL, Hills SL, Fischer M, et al. Estimated global incidence of Japanese encephalitis: a systematic review. *Bull World Health Organ* 2011;89:766–74. <https://doi.org/10.2471/BLT.10.085233>
- Solomon T, Dung NM, Kneen R, Gainsborough M, Vaughn DW, Khanh VT. Japanese encephalitis. *J Neurol Neurosurg Psychiatry* 2000;68:405–15. <https://doi.org/10.1136/jnnp.68.4.405>
- World Health Organization. Japanese encephalitis vaccines: WHO position paper—February 2015. *Wkly Epidemiol Rec* 2015;90:69–87.
- CDC. Inactivated Japanese encephalitis virus vaccine. Recommendations of the Advisory Committee on Immunization Practices (ACIP). *MMWR Recomm Rep* 1993;42(No. RR-1).
- Hills SL, Griggs AC, Fischer M. Japanese encephalitis in travelers from non-endemic countries, 1973–2008. *Am J Trop Med Hyg* 2010;82:930–6. <https://doi.org/10.4269/ajtmh.2010.09-0676>
- Hills SL, Stoltey J, Martínez D, et al. A case series of three U.S. adults with Japanese encephalitis, 2010–2012. *J Travel Med* 2014;21:310–3. <https://doi.org/10.1111/jtm.12127>
- Shlim DR, Solomon T. Japanese encephalitis vaccine for travelers: exploring the limits of risk. *Clin Infect Dis* 2002;35:183–8. <https://doi.org/10.1086/341247>
- Endy TP, Nisalak A. Japanese encephalitis virus: ecology and epidemiology. *Curr Top Microbiol Immunol* 2002;267:11–48. https://doi.org/10.1007/978-3-642-59403-8_2
- Vaughn DW, Hoke CH Jr. The epidemiology of Japanese encephalitis: prospects for prevention. *Epidemiol Rev* 1992;14:197–221. <https://doi.org/10.1093/oxfordjournals.epirev.a036087>
- Food and Drug Administration. *Ixiaro: Japanese encephalitis vaccine, inactivated, adsorbed [package insert]*. Vienna, Austria: Valneva Austria GmbH; 2018. <https://www.fda.gov/media/75777/download>
- Fischer M, Lindsey N, Staples JE, Hills S. Japanese encephalitis vaccines: recommendations of the Advisory Committee on Immunization Practices (ACIP). *MMWR Recomm Rep* 2010;59(No. RR-1).
- CDC. Recommendations for use of a booster dose of inactivated Vero cell culture-derived Japanese encephalitis vaccine: Advisory Committee on Immunization Practices, 2011. *MMWR Morb Mortal Wkly Rep* 2011;60:661–3.
- CDC. Use of Japanese encephalitis vaccine in children: recommendations of the Advisory Committee on Immunization Practices, 2013. *MMWR Morb Mortal Wkly Rep* 2013;62:898–900.
- Ahmed F. Advisory Committee on Immunization Practices handbook for developing evidence-based recommendations. Version 1.2. Atlanta, GA: US Department of Health and Human Services, CDC; 2013. <https://www.cdc.gov/vaccines/acip/recs/grade/downloads/handbook.pdf>
- CDC. Japanese encephalitis vaccine evidence to recommendations. Atlanta, GA: US Department of Health and Human Services, CDC. <https://www.cdc.gov/vaccines/acip/recs/grade/table-refs.html>
- Lee G, Carr W; ACIP Evidence-Based Recommendations Work Group. Updated framework for development of evidence-based recommendations by the Advisory Committee on Immunization Practices. *MMWR Morb Mortal Wkly Rep* 2018;67:1271–2. <https://doi.org/10.15585/mmwr.mm6745a4>
- Kuno G, Chang GJ, Tsuchiya KR, Karabatsos N, Cropp CB. Phylogeny of the genus *Flavivirus*. *J Virol* 1998;72:73–83.
- Mackenzie JS, Barrett AD, Deubel V. The Japanese encephalitis serological group of flaviviruses: a brief introduction to the group. *Curr Top Microbiol Immunol* 2002;267:1–10. https://doi.org/10.1007/978-3-642-59403-8_1
- Schuh AJ, Ward MJ, Leigh Brown AJ, Barrett AD. Dynamics of the emergence and establishment of a newly dominant genotype of Japanese encephalitis virus throughout Asia. *J Virol* 2014;88:4522–32. <https://doi.org/10.1128/JVI.02686-13>
- Buescher EL, Scherer WF. Ecologic studies of Japanese encephalitis virus in Japan. IX. Epidemiologic correlations and conclusions. *Am J Trop Med Hyg* 1959;8:719–22. <https://doi.org/10.4269/ajtmh.1959.8.719>
- Buescher EL, Scherer WF, McClure HE, et al. Ecologic studies of Japanese encephalitis virus in Japan. IV. Avian infection. *Am J Trop Med Hyg* 1959;8:678–88. <https://doi.org/10.4269/ajtmh.1959.8.678>
- Buescher EL, Scherer WF, Rosenberg MZ, Gresser I, Hardy JL, Bullock HR. Ecologic studies of Japanese encephalitis virus in Japan. II. Mosquito infection. *Am J Trop Med Hyg* 1959;8:651–64. <https://doi.org/10.4269/ajtmh.1959.8.651>
- Rosen L. The natural history of Japanese encephalitis virus. *Annu Rev Microbiol* 1986;40:395–414. <https://doi.org/10.1146/annurev.mi.40.100186.002143>
- Scherer WF, Moyer JT, Izumi T, Gresser I, McCown J. Ecologic studies of Japanese encephalitis virus in Japan. VI. Swine infection. *Am J Trop Med Hyg* 1959;8:698–706. <https://doi.org/10.4269/ajtmh.1959.8.698>
- Konno J, Endo K, Agatsuma H, Ishida N. Cyclic outbreaks of Japanese encephalitis among pigs and humans. *Am J Epidemiol* 1966;84:292–300. <https://doi.org/10.1093/oxfordjournals.aje.a120643>
- Kodama K, Sasaki N, Inoue YK. Studies of live attenuated Japanese encephalitis vaccine in swine. *J Immunol* 1968;100:194–200.
- Ueba N, Maeda A, Otsu K, Mitsuda B, Kimoto T. Natural infection of swine by Japanese encephalitis virus and its modification by vaccination. *Biken J* 1972;15:67–79.
- Simpson DIH, Smith CEG, Marshall TF, et al. Arbovirus infections in Sarawak: the role of the domestic pig. *Trans R Soc Trop Med Hyg* 1976;70:66–72. [https://doi.org/10.1016/0035-9203\(76\)90010-9](https://doi.org/10.1016/0035-9203(76)90010-9)

33. Gresser I, Hardy JL, Hu SM, Scherer WF. Factors influencing transmission of Japanese B encephalitis virus by a colonized strain of *Culex tritaeniorhynchus* Giles, from infected pigs and chicks to susceptible pigs and birds. *Am J Trop Med Hyg* 1958;7:365–73. <https://doi.org/10.4269/ajtmh.1958.7.365>
34. Hill MN. Japanese encephalitis in Sarawak: studies on adult mosquito populations. *Trans R Soc Trop Med Hyg* 1970;64:489–96. [https://doi.org/10.1016/0035-9203\(70\)90068-4](https://doi.org/10.1016/0035-9203(70)90068-4)
35. Simpson DI, Bowen ET, Way HJ, et al. Arbovirus infections in Sarawak, October 1968–February 1970: Japanese encephalitis virus isolations from mosquitoes. *Ann Trop Med Parasitol* 1974;68:393–404. <https://doi.org/10.1080/00034983.1974.11686966>
36. Peiris JS, Amerasinghe FP, Amerasinghe PH, Ratnayake CB, Karunaratne SH, Tsai TF. Japanese encephalitis in Sri Lanka—the study of an epidemic: vector incrimination, porcine infection and human disease. *Trans R Soc Trop Med Hyg* 1992;86:307–13. [https://doi.org/10.1016/0035-9203\(92\)90325-7](https://doi.org/10.1016/0035-9203(92)90325-7)
37. Gajanana A, Rajendran R, Samuel PP, et al. Japanese encephalitis in south Arcot district, Tamil Nadu, India: a three-year longitudinal study of vector abundance and infection frequency. *J Med Entomol* 1997;34:651–9. <https://doi.org/10.1093/jmedent/34.6.651>
38. Kanojia PC, Shetty PS, Geevarghese G. A long-term study on vector abundance & seasonal prevalence in relation to the occurrence of Japanese encephalitis in Gorakhpur district, Uttar Pradesh. *Indian J Med Res* 2003;117:104–10.
39. Keiser J, Maltese ME, Erlanger TE, et al. Effect of irrigated rice agriculture on Japanese encephalitis, including challenges and opportunities for integrated vector management. *Acta Trop* 2005;95:40–57. <https://doi.org/10.1016/j.actatropica.2005.04.012>
40. Olson JG, Ksiazek TG, Tan R, Atmosoedjono S, Lee VH, Converse JD. Correlation of population indices of female *Culex tritaeniorhynchus* with Japanese encephalitis viral activity in Kapuk, Indonesia. *Southeast Asian J Trop Med Public Health* 1985;16:337–42.
41. Gould DJ, Edelman R, Grossman RA, Nisalak A, Sullivan MF. Study of Japanese encephalitis virus in Chiangmai Valley, Thailand. IV. Vector studies. *Am J Epidemiol* 1974;100:49–56. <https://doi.org/10.1093/oxfordjournals.aje.a112008>
42. Reisen WK, Aslamkhan M, Basio RG. The effects of climatic patterns and agricultural practices on the population dynamics of *Culex tritaeniorhynchus* in Asia. *Southeast Asian J Trop Med Public Health* 1976;61–71.
43. Hanna JN, Ritchie SA, Phillips DA, et al. An outbreak of Japanese encephalitis in the Torres Strait, Australia, 1995. *Med J Aust* 1996;165:256–60. <https://doi.org/10.5694/j.1326-5377.1996.tb124960.x>
44. Scherer WF, Kitaoka M, Okuno T, Ogata T. Ecologic studies of Japanese encephalitis virus in Japan. VII. Human infection. *Am J Trop Med Hyg* 1959;8:707–15. <https://doi.org/10.4269/ajtmh.1959.8.707>
45. Chaturvedi UC, Mathur A, Chandra A, Das SK, Tandon HO, Singh UK. Transplacental infection with Japanese encephalitis virus. *J Infect Dis* 1980;141:712–5. <https://doi.org/10.1093/infdis/141.6.712>
46. Mathur A, Tandon HO, Mathur KR, Sarkari NB, Singh UK, Chaturvedi UC. Japanese encephalitis virus infection during pregnancy. *Indian J Med Res* 1985;81:9–12.
47. Cheng VCC, Sridhar S, Wong SC, et al. Japanese encephalitis virus transmitted via blood transfusion, Hong Kong, China. *Emerg Infect Dis* 2018;24:49–57. <https://doi.org/10.3201/eid2401.171297>
48. Iwamoto M, Jernigan DB, Guasch A, et al; West Nile Virus in Transplant Recipients Investigation Team. Transmission of West Nile virus from an organ donor to four transplant recipients. *N Engl J Med* 2003;348:2196–203. <https://doi.org/10.1056/NEJMoa022987>
49. The Subcommittee on Arbovirus Laboratory Safety of the American Committee on Arthropod-Borne Viruses. Laboratory safety for arboviruses and certain other viruses of vertebrates. *Am J Trop Med Hyg* 1980;29:1359–81. <https://doi.org/10.4269/ajtmh.1980.29.1359>
50. Wu YC, Huang YS, Chien LJ, et al. The epidemiology of Japanese encephalitis on Taiwan during 1966–1997. *Am J Trop Med Hyg* 1999;61:78–84. <https://doi.org/10.4269/ajtmh.1999.61.78>
51. Sohn YM. Japanese encephalitis immunization in South Korea: past, present, and future. *Emerg Infect Dis* 2000;6:17–24.
52. Grayston JT, Wang SP, Yen CH. Encephalitis on Taiwan. I. Introduction and epidemiology. *Am J Trop Med Hyg* 1962;11:126–30. <https://doi.org/10.4269/ajtmh.1962.11.126>
53. Kono R, Kim KH. Comparative epidemiological features of Japanese encephalitis in the Republic of Korea, China (Taiwan) and Japan. *Bull World Health Organ* 1969;40:263–77.
54. Igarashi A. Epidemiology and control of Japanese encephalitis. *World Health Stat Q* 1992;45:299–305.
55. Okuno T. An epidemiological review of Japanese encephalitis. *World Health Stat Q* 1978;31:120–33.
56. Umenai T, Krzysko R, Bektimirov TA, Assaad FA. Japanese encephalitis: current worldwide status. *Bull World Health Organ* 1985;63:625–31.
57. Yongxin Y. Japanese encephalitis in China. *Southeast Asian J Trop Med Public Health* 1995;26(suppl 3):17–21.
58. Carey DE, Myers RM, Webb JK, Reuben R. Japanese encephalitis in South India. A summary of recent knowledge. *J Indian Med Assoc* 1969;52:10–5.
59. Yamada T, Rojanasuphot S, Takagi M, Wungkobkiat S, Hirota T. Studies on an epidemic of Japanese encephalitis in the northern region of Thailand in 1969 and 1970. *Biken J* 1971;14:267–96.
60. Khan AM, Khan AQ, Dobrzynski L, Joshi GP, Myat A. A Japanese encephalitis focus in Bangladesh. *J Trop Med Hyg* 1981;84:41–4.
61. Poneprasert B. Japanese encephalitis in children in northern Thailand. *Southeast Asian J Trop Med Public Health* 1989;20:599–603.
62. Ha D, Huong V, Loan H, Thong D, Deubel V. Current situation of Japanese encephalitis in the south of Vietnam, 1976–1992. *Trop Med* 1994;36:202–14.
63. Cardoso MJ, Hooi TP, Kaur P. Japanese encephalitis virus is an important cause of encephalitis among children in Penang. *Southeast Asian J Trop Med Public Health* 1995;26:272–5.
64. Chunsuttiwat S, Warachit P. Japanese encephalitis in Thailand. *Southeast Asian J Trop Med Public Health* 1995;26(suppl 3):43–6.
65. Joshi D. Current status of Japanese encephalitis in Nepal. *Southeast Asian J Trop Med Public Health* 1995;26(suppl 3):34–40.
66. Tam N, Yen N. Japanese encephalitis in Vietnam 1985–93. *Southeast Asian J Trop Med Public Health* 1995;26(suppl3):47–50.
67. Vitarana T. Japanese encephalitis in Sri Lanka. *Southeast Asian J Trop Med Public Health* 1995;26(suppl 3):41–2.
68. Bista MB, Shrestha JM. Epidemiological situation of Japanese encephalitis in Nepal. *JNMA J Nepal Med Assoc* 2005;44:51–6. <https://doi.org/10.31729/jnma.397>
69. Paul WS, Moore PS, Karabatsos N, et al. Outbreak of Japanese encephalitis on the island of Saipan, 1990. *J Infect Dis* 1993;167:1053–8. <https://doi.org/10.1093/infdis/167.5.1053>

70. Hanna JN, Ritchie SA, Phillips DA, et al. Japanese encephalitis in north Queensland, Australia, 1998. *Med J Aust* 1999;170:533–6. <https://doi.org/10.5694/j.1326-5377.1999.tb127878.x>
71. Li YX, Li MH, Fu SH, et al. Japanese encephalitis, Tibet, China. *Emerg Infect Dis* 2011;17:934–6. <https://doi.org/10.3201/eid1705.101417>
72. Bhattachan A, Amatya S, Sedai TR, Upreti SR, Partridge J. Japanese encephalitis in hill and mountain districts, Nepal. *Emerg Infect Dis* 2009;15:1691–2. <https://doi.org/10.3201/eid1510.081641>
73. World Health Organization. Japanese encephalitis reported cases. Geneva, Switzerland: World Health Organization. http://apps.who.int/immunization_monitoring/globalsummary/timeseries/tsincidencejapenc.html
74. Wierzbica TE, Ghimire P, Malla S, et al. Laboratory-based Japanese encephalitis surveillance in Nepal and the implications for a national immunization strategy. *Am J Trop Med Hyg* 2008;78:1002–6. <https://doi.org/10.4269/ajtmh.2008.78.1002>
75. Touch S, Hills S, Sokhal B, et al. Epidemiology and burden of disease from Japanese encephalitis in Cambodia: results from two years of sentinel surveillance. *Trop Med Int Health* 2009;14:1365–73. <https://doi.org/10.1111/j.1365-3156.2009.02380.x>
76. Kari K, Liu W, Gautama K, et al. A hospital-based surveillance for Japanese encephalitis in Bali, Indonesia. *BMC Med* 2006;4:8. <https://doi.org/10.1186/1741-7015-4-8>
77. Arai S, Matsunaga Y, Takasaki T, et al. Vaccine Preventable Diseases Surveillance Program of Japan. Japanese encephalitis: surveillance and elimination effort in Japan from 1982 to 2004. *Jpn J Infect Dis* 2008;61:333–8.
78. Gingrich JB, Nisalak A, Latendresse JR, et al. Japanese encephalitis virus in Bangkok: factors influencing vector infections in three suburban communities. *J Med Entomol* 1992;29:436–44. <https://doi.org/10.1093/jmedent/29.3.436>
79. Lindahl J, Chirico J, Boqvist S, Thu HTV, Magnusson U. Occurrence of Japanese encephalitis virus mosquito vectors in relation to urban pig holdings. *Am J Trop Med Hyg* 2012;87:1076–82. <https://doi.org/10.4269/ajtmh.2012.12-0315>
80. Di Francesco J, Choeung R, Peng B, et al. Comparison of the dynamics of Japanese encephalitis virus circulation in sentinel pigs between a rural and a peri-urban setting in Cambodia. *PLoS Negl Trop Dis* 2018;12:e0006644. <https://doi.org/10.1371/journal.pntd.0006644>
81. Kanagasabai K, Joshua V, Ravi M, et al. Epidemiology of Japanese encephalitis in India: analysis of laboratory surveillance data, 2014–2017. *J Infect* 2018;76:317–20. <https://doi.org/10.1016/j.jinf.2017.09.018>
82. Touch S, Hills S, Sokhal B, et al. Epidemiology and burden of disease from Japanese encephalitis in Cambodia: results from two years of sentinel surveillance. *Trop Med Int Health* 2009;14:1365–73. <https://doi.org/10.1111/j.1365-3156.2009.02380.x>
83. Grossman RA, Edelman R, Willhight M, Pantuwatana S, Udomsakdi S. Study of Japanese encephalitis virus in Chiangmai Valley, Thailand. 3. Human seroepidemiology and inapparent infections. *Am J Epidemiol* 1973;98:133–49. <https://doi.org/10.1093/oxfordjournals.aje.a121538>
84. Potula R, Badrinath S, Srinivasan S. Japanese encephalitis in and around Pondicherry, South India: a clinical appraisal and prognostic indicators for the outcome. *J Trop Pediatr* 2003;49:48–53. <https://doi.org/10.1093/tropej/49.1.48>
85. Yin Z, Wang H, Yang J, et al. Acute Meningitis and Encephalitis Syndrome (AMES) Study Group. Japanese encephalitis disease burden and clinical features of Japanese encephalitis in four cities in the People's Republic of China. *Am J Trop Med Hyg* 2010;83:766–73. <https://doi.org/10.4269/ajtmh.2010.09-0748>
86. Sunwoo JS, Jung KH, Lee ST, Lee SK, Chu K. Reemergence of Japanese Encephalitis in South Korea, 2010–2015. *Emerg Infect Dis* 2016;22:1841–3. <https://doi.org/10.3201/eid2210.160288>
87. Ayukawa R, Fujimoto H, Ayabe M, et al. An unexpected outbreak of Japanese encephalitis in the Chugoku district of Japan, 2002. *Jpn J Infect Dis* 2004;57:63–6.
88. Wang LH, Fu SH, Wang HY, et al. Japanese encephalitis outbreak, Yuncheng, China, 2006. *Emerg Infect Dis* 2007;13:1123–5. <https://doi.org/10.3201/eid1307.070010>
89. Gurav YK, Bondre VP, Tandale BV, et al. A large outbreak of Japanese encephalitis predominantly among adults in northern region of West Bengal, India. *J Med Virol* 2016;88:2004–11. <https://doi.org/10.1002/jmv.24556>
90. Gajanana A, Thenmozhi V, Samuel PP, Reuben R. A community-based study of subclinical flavivirus infections in children in an area of Tamil Nadu, India, where Japanese encephalitis is endemic. *Bull World Health Organ* 1995;73:237–44.
91. Halstead SB, Grosz CR. Subclinical Japanese encephalitis. I. Infection of Americans with limited residence in Korea. *Am J Hyg* 1962;75:190–201.
92. Benenson MW, Top FH Jr, Gresso W, Ames CW, Altstatt LB. The virulence to man of Japanese encephalitis virus in Thailand. *Am J Trop Med Hyg* 1975;24:974–80. <https://doi.org/10.4269/ajtmh.1975.24.974>
93. Hsieh WC, Wallace CK, Wang SP, Rasmussen AF Jr. Inapparent infection with Japanese encephalitis of American servicemen on Okinawa in 1960. *Am J Trop Med Hyg* 1963;12:413–6. <https://doi.org/10.4269/ajtmh.1963.12.413>
94. Southam CM. Serological studies of encephalitis in Japan. II. Inapparent infections by Japanese B encephalitis virus. *J Infect Dis* 1956;99:163–9. <https://doi.org/10.1093/infdis/99.2.163>
95. Lincoln AF, Sivertson SE. Acute phase of Japanese B encephalitis; two hundred and one cases in American soldiers, Korea, 1950. *J Am Med Assoc* 1952;150:268–73. <https://doi.org/10.1001/jama.1952.03680040010003>
96. Watt G, Jongsakul K. Acute undifferentiated fever caused by infection with Japanese encephalitis virus. *Am J Trop Med Hyg* 2003;68:704–6. <https://doi.org/10.4269/ajtmh.2003.68.704>
97. Kuwayama M, Ito M, Takao S, et al. Japanese encephalitis virus in meningitis patients, Japan. *Emerg Infect Dis* 2005;11:471–3.
98. Kumar R, Mathur A, Kumar A, Sharma S, Chakraborty S, Chaturvedi UC. Clinical features & prognostic indicators of Japanese encephalitis in children in Lucknow (India). *Indian J Med Res* 1990;91:321–7.
99. Kumar R, Tripathi P, Singh S, Bannerji G. Clinical features in children hospitalized during the 2005 epidemic of Japanese encephalitis in Uttar Pradesh, India. *Clin Infect Dis* 2006;43:123–31. <https://doi.org/10.1086/505121>
100. Schneider RJ, Firestone MH, Edelman R, Chieowanich P, Pornpibul R. Clinical sequelae after Japanese encephalitis: a one year follow-up study in Thailand. *Southeast Asian J Trop Med Public Health* 1974;5:560–8.
101. Misra UK, Kalita J. Prognosis of Japanese encephalitis patients with dystonia compared to those with parkinsonian features only. *Postgrad Med J* 2002;78:238–41. <https://doi.org/10.1136/pmj.78.918.238>

102. Kalita J, Misra UK. Markedly severe dystonia in Japanese encephalitis. *Mov Disord* 2000;15:1168–72. [https://doi.org/10.1002/1531-8257\(200011\)15:6<1168::AID-MDS1016>3.0.CO;2-V](https://doi.org/10.1002/1531-8257(200011)15:6<1168::AID-MDS1016>3.0.CO;2-V)
103. Rayamajhi A, Singh R, Prasad R, Khanal B, Singhi S. Clinico-laboratory profile and outcome of Japanese encephalitis in Nepali children. *Ann Trop Paediatr* 2006;26:293–301. <https://doi.org/10.1179/146532806X152818>
104. Solomon T, Dung NM, Kneen R, et al. Seizures and raised intracranial pressure in Vietnamese patients with Japanese encephalitis. *Brain* 2002;125:1084–93. <https://doi.org/10.1093/brain/awf116>
105. Misra UK, Kalita J. Seizures in Japanese encephalitis. *J Neurol Sci* 2001;190:57–60. [https://doi.org/10.1016/S0022-510X\(01\)00589-5](https://doi.org/10.1016/S0022-510X(01)00589-5)
106. Misra UK, Kalita J. Anterior horn cells are also involved in Japanese encephalitis. *Acta Neurol Scand* 1997;96:114–7. <https://doi.org/10.1111/j.1600-0404.1997.tb00250.x>
107. Solomon T, Kneen R, Dung NM, et al. Poliomyelitis-like illness due to Japanese encephalitis virus. *Lancet* 1998;351:1094–7. [https://doi.org/10.1016/S0140-6736\(97\)07509-0](https://doi.org/10.1016/S0140-6736(97)07509-0)
108. Halstead SB, Jacobson J. Japanese encephalitis. *Adv Virus Res* 2003;61:103–38. [https://doi.org/10.1016/S0065-3527\(03\)61003-1](https://doi.org/10.1016/S0065-3527(03)61003-1)
109. Libraty DH, Nisalak A, Endy TP, Suntayakorn S, Vaughn DW, Innis BL. Clinical and immunological risk factors for severe disease in Japanese encephalitis. *Trans R Soc Trop Med Hyg* 2002;96:173–8. [https://doi.org/10.1016/S0035-9203\(02\)90294-4](https://doi.org/10.1016/S0035-9203(02)90294-4)
110. Maschke M, Kastrup O, Forsting M, Diener HC. Update on neuroimaging in infectious central nervous system disease. *Curr Opin Neurol* 2004;17:475–80. <https://doi.org/10.1097/01.wco.0000137540.29857.bf>
111. Wang HS. Comparison of magnetic resonance imaging abnormalities in Japanese encephalitis and acute necrotizing encephalopathy of childhood. *Arch Neurol* 2004;61:1149–50, author reply 1150. <https://doi.org/10.1001/archneur.61.7.1149>
112. Dung NM, Turtle L, Chong WK, et al. An evaluation of the usefulness of neuroimaging for the diagnosis of Japanese encephalitis. *J Neurol* 2009;256:2052–60. <https://doi.org/10.1007/s00415-009-5249-5>
113. Burke DS, Nisalak A, Hoke CH Jr. Field trial of a Japanese encephalitis diagnostic kit. *J Med Virol* 1986;18:41–9. <https://doi.org/10.1002/jmv.1890180106>
114. Burke DS, Nisalak A, Ussery MA, Laorakpongse T, Chantavibul S. Kinetics of IgM and IgG responses to Japanese encephalitis virus in human serum and cerebrospinal fluid. *J Infect Dis* 1985;151:1093–9. <https://doi.org/10.1093/infdis/151.6.1093>
115. Chanama S, Sukprasert W, Sa-ngasang A, et al. Detection of Japanese encephalitis (JE) virus-specific IgM in cerebrospinal fluid and serum samples from JE patients. *Jpn J Infect Dis* 2005;58:294–6.
116. Martin DA, Biggerstaff BJ, Allen B, Johnson AJ, Lanciotti RS, Roehrig JT. Use of immunoglobulin m cross-reactions in differential diagnosis of human flaviviral encephalitis infections in the United States. *Clin Diagn Lab Immunol* 2002;9:544–9.
117. Martin DA, Muth DA, Brown T, Johnson AJ, Karabatsos N, Roehrig JT. Standardization of immunoglobulin M capture enzyme-linked immunosorbent assays for routine diagnosis of arboviral infections. *J Clin Microbiol* 2000;38:1823–6.
118. Srey VH, Sadones H, Ong S, et al. Etiology of encephalitis syndrome among hospitalized children and adults in Takeo, Cambodia, 1999–2000. *Am J Trop Med Hyg* 2002;66:200–7. <https://doi.org/10.4269/ajtmh.2002.66.200>
119. Hills S, Dabbagh A, Jacobson J, et al; Japanese Encephalitis Core Working Group. Evidence and rationale for the World Health Organization recommended standards for Japanese encephalitis surveillance. *BMC Infect Dis* 2009;9:214. <https://doi.org/10.1186/1471-2334-9-214>
120. Burke DS, Lorsomrudee W, Leake CJ, et al. Fatal outcome in Japanese encephalitis. *Am J Trop Med Hyg* 1985;34:1203–10. <https://doi.org/10.4269/ajtmh.1985.34.1203>
121. Swami R, Ratho RK, Mishra B, Singh MP. Usefulness of RT-PCR for the diagnosis of Japanese encephalitis in clinical samples. *Scand J Infect Dis* 2008;40:815–20. <https://doi.org/10.1080/00365540802227102>
122. Huang GKL, Tio SY, Caly L, et al. Prolonged detection of Japanese encephalitis virus in urine and whole blood in a returned short-term traveler. *Open Forum Infect Dis* 2017;4:ofx203. <https://doi.org/10.1093/ofid/ofx203>
123. Han XY, Ren QW, Xu ZY, Tsai TF. Serum and cerebrospinal fluid immunoglobulins M, A, and G in Japanese encephalitis. *J Clin Microbiol* 1988;26:976–8.
124. Solomon T, Thao LT, Dung NM, et al. Rapid diagnosis of Japanese encephalitis by using an immunoglobulin M dot enzyme immunoassay. *J Clin Microbiol* 1998;36:2030–4.
125. Turtle L, Solomon T. Japanese encephalitis—the prospects for new treatments. *Nat Rev Neurol* 2018;14:298–313. <https://doi.org/10.1038/nrneurol.2018.30>
126. Kumar R, Tripathi P, Baranwal M, Singh S, Tripathi S, Banerjee G. Randomized, controlled trial of oral ribavirin for Japanese encephalitis in children in Uttar Pradesh, India. *Clin Infect Dis* 2009;48:400–6. <https://doi.org/10.1086/596309>
127. Hoke CH Jr, Vaughn DW, Nisalak A, et al. Effect of high-dose dexamethasone on the outcome of acute encephalitis due to Japanese encephalitis virus. *J Infect Dis* 1992;165:631–7. <https://doi.org/10.1093/infdis/165.4.631>
128. Solomon T, Dung NM, Wills B, et al. Interferon alfa-2a in Japanese encephalitis: a randomised double-blind placebo-controlled trial. *Lancet* 2003;361:821–6. [https://doi.org/10.1016/S0140-6736\(03\)12709-2](https://doi.org/10.1016/S0140-6736(03)12709-2)
129. Rayamajhi A, Nightingale S, Bhatta NK, et al. A preliminary randomized double blind placebo-controlled trial of intravenous immunoglobulin for Japanese encephalitis in Nepal. *PLoS One* 2015;10:e0122608. <https://doi.org/10.1371/journal.pone.0122608>
130. Kumar R, Basu A, Sinha S, et al. Role of oral Minocycline in acute encephalitis syndrome in India—a randomized controlled trial. *BMC Infect Dis* 2016;16:67. <https://doi.org/10.1186/s12879-016-1385-6>
131. Kumar R, Mathur A, Singh KB, et al. Clinical sequelae of Japanese encephalitis in children. *Indian J Med Res* 1993;97:9–13.
132. Maha MS, Moniaga VA, Hills SL, et al. Outcome and extent of disability following Japanese encephalitis in Indonesian children. *Int J Infect Dis* 2009;13:e389–93. <https://doi.org/10.1016/j.ijid.2009.01.009>
133. Pieper SJ Jr, Kurland LT. Sequelae of Japanese B and mumps encephalitis: recent follow-up of patients affected in 1947–1948 epidemic on Guam. *Am J Trop Med Hyg* 1958;7:481–90. <https://doi.org/10.4269/ajtmh.1958.7.481>
134. Richter RW, Shimoyjo S. Neurologic sequelae of Japanese B encephalitis. *Neurology* 1961;11:553–9. <https://doi.org/10.1212/WNL.11.7.553>
135. Huy BV, Tu HC, Luan TV, Lindqvist R. Early mental and neurological sequelae after Japanese B encephalitis. *Southeast Asian J Trop Med Public Health* 1994;25:549–53.
136. Murgod UA, Muthane UB, Ravi V, Radhesh S, Desai A. Persistent movement disorders following Japanese encephalitis. *Neurology* 2001;57:2313–5. <https://doi.org/10.1212/WNL.57.12.2313>

137. Ding D, Hong Z, Zhao SJ, et al. Long-term disability from acute childhood Japanese encephalitis in Shanghai, China. *Am J Trop Med Hyg* 2007;77:528–33. <https://doi.org/10.4269/ajtmh.2007.77.528>
138. Hills SL, Van Cuong N, Touch S, et al. Disability from Japanese encephalitis in Cambodia and Viet Nam. *J Trop Pediatr* 2011;57:241–4. <https://doi.org/10.1093/tropej/fmp133>
139. Hatz C, Werlein J, Mutsch M, Hufnagel M, Behrens RH. Japanese encephalitis: defining risk incidence for travelers to endemic countries and vaccine prescribing from the UK and Switzerland. *J Travel Med* 2009;16:200–3. <https://doi.org/10.1111/j.1708-8305.2009.00334.x>
140. Macdonald WB, Tink AR, Ouvrier RA, et al. Japanese encephalitis after a two-week holiday in Bali. *Med J Aust* 1989;150:334–6,9.
141. Wittesjö B, Eitrem R, Niklasson B, Vene S, Mangiafico JA. Japanese encephalitis after a 10-day holiday in Bali. *Lancet* 1995;345:856–7. [https://doi.org/10.1016/S0140-6736\(95\)92990-8](https://doi.org/10.1016/S0140-6736(95)92990-8)
142. Caramello P, Canta F, Balbiano R, et al. A case of imported JE acquired during short travel in Vietnam. Are current recommendations about vaccination broader? *J Travel Med* 2007;14:346–8. <https://doi.org/10.1111/j.1708-8305.2007.00140.x>
143. CDC. Japanese encephalitis among three U.S. travelers returning from Asia, 2003–2008. *MMWR Morb Mortal Wkly Rep* 2009;58:737–40.
144. CDC. Japanese encephalitis in a U.S. traveler returning from Thailand, 2004. *MMWR Morb Mortal Wkly Rep* 2005;54:123–5.
145. Rose MR, Hughes SM, Gatus BJ. A case of Japanese B encephalitis imported into the United Kingdom. *J Infect* 1983;6:261–5. [https://doi.org/10.1016/S0163-4453\(83\)93693-9](https://doi.org/10.1016/S0163-4453(83)93693-9)
146. Trillen C. American chronicles. Zei-da-man. *New Yorker* 1985;61–93.
147. Burdon JT, Stanley PJ, Lloyd G, Jones NC. A case of Japanese encephalitis. *J Infect* 1994;28:175–9. [https://doi.org/10.1016/S0163-4453\(94\)95640-5](https://doi.org/10.1016/S0163-4453(94)95640-5)
148. Buhl MR, Lindquist L. Japanese encephalitis in travelers: review of cases and seasonal risk. *J Travel Med* 2009;16:217–9. <https://doi.org/10.1111/j.1708-8305.2009.00333.x>
149. Buhl MR, Black FT, Andersen PL, Laursen A. Fatal Japanese encephalitis in a Danish tourist visiting Bali for 12 days. *Scand J Infect Dis* 1996;28:189. <https://doi.org/10.3109/00365549609049074>
150. Pogodina VV, Bochkova NG, Leshchinskaia EV, Levina LS. [Japanese encephalitis in citizens of Russia who travel abroad]. *Vopr Virusol* 1996;41:8–11.
151. Bernard P, Jambaud E, Berbineau A, Brunot J, Flechère A. [Japanese encephalitis: an exceptional imported arbovirus]. *Presse Med* 1998;27:1327.
152. Saito M, Sunagawa T, Makino Y, et al. Three Japanese encephalitis cases in Okinawa, Japan, 1991. *Southeast Asian J Trop Med Public Health* 1999;30:277–9.
153. Monnet FP. Behavioural disturbances following Japanese B encephalitis. *Eur Psychiatry* 2003;18:269–73. <https://doi.org/10.1016/j.eurpsy.2003.09.001>
154. Geraghty CM, McCarthy JS. Japanese encephalitis vaccine: is it being sufficiently used in travellers? *Med J Aust* 2004;181:269–70.
155. Hanson JR, Taylor CT, Richards AR, Smith IL, Boutlis CS. Japanese encephalitis acquired near Port Moresby: implications for residents and travellers to Papua New Guinea. *Med J Aust* 2004;181:282–3.
156. Ostlund MR, Kan B, Karlsson M, Vene S. Japanese encephalitis in a Swedish tourist after travelling to Java and Bali. *Scand J Infect Dis* 2004;36:512–3. <https://doi.org/10.1080/00365540410020640>
157. Cutfield NJ, Anderson NE, Brickell K, Hueston L, Pikhholz C, Roxburgh RH. Japanese encephalitis acquired during travel in China. *Intern Med J* 2005;35:497–8. <https://doi.org/10.1111/j.1445-5994.2005.00852.x>
158. Delsing CE, Ardesch J, Nihom J, Mulder L, Kootstra GJ, Hylkema BS. [An unusual cause of meningo-encephalitis: Japanese encephalitis]. *Ned Tijdschr Geneesk* 2005;149:2423–7.
159. Lehtinen VA, Huhtamo E, Siikamäki H, Vapalahti O. Japanese encephalitis in a Finnish traveler on a two-week holiday in Thailand. *J Clin Virol* 2008;43:93–5. <https://doi.org/10.1016/j.jcv.2008.05.001>
160. Poland JD, Cropp CB, Craven RB, Monath TP. Evaluation of the potency and safety of inactivated Japanese encephalitis vaccine in US inhabitants. *J Infect Dis* 1990;161:878–82. <https://doi.org/10.1093/infdis/161.5.878>
161. Artsob H, Spence L. Imported arbovirus infections in Canada 1974–89. *Can J Infect Dis* 1991;2:95–100. <https://doi.org/10.1155/1991/678906>
162. Jeurissen A, Strauven T. A case of aseptic meningitis due to Japanese encephalitis virus in a traveller returning from the Philippines. *Acta Neurol Belg* 2011;111:143–5.
163. Tappe D, Nemecek A, Zipp F, et al. Two laboratory-confirmed cases of Japanese encephalitis imported to Germany by travelers returning from Southeast Asia. *J Clin Virol* 2012;54:282–5. <https://doi.org/10.1016/j.jcv.2012.03.004>
164. Werlinrud AM, Christiansen CB, Koch A. Japanese encephalitis in a Danish short-term traveler to Cambodia. *J Travel Med* 2011;18:411–3. <https://doi.org/10.1111/j.1708-8305.2011.00565.x>
165. CDC. Japanese encephalitis in two children—United States, 2010. *MMWR Morb Mortal Wkly Rep* 2011;60:276–8.
166. Langevin S, Libman M, Drebot MA, Laverdière M. A case of Japanese encephalitis virus infection acquired during a trip in Thailand. *J Travel Med* 2012;19:127–9. <https://doi.org/10.1111/j.1708-8305.2011.00582.x>
167. Lee DW, Choe YJ, Kim JH, et al. Epidemiology of Japanese encephalitis in South Korea, 2007–2010. *Int J Infect Dis* 2012;16:e448–52. <https://doi.org/10.1016/j.ijid.2012.02.006>
168. Australian Government Department of Health. National Notifiable Diseases Surveillance System. Canberra, Australia: Australian Government Department of Health. <http://www9.health.gov.au/cda/source/cda-index.cfm>
169. Doti P, Castro P, Martínez MJ, et al. A case of Japanese encephalitis in a 20 year-old Spanish sportsman, February 2013. *Euro Surveill* 2013;18:20573. <https://doi.org/10.2807/1560-7917.ES2013.18.35.20573>
170. Lagarde S, Lagier JC, Charrel R, et al. Japanese encephalitis in a French traveler to Nepal. *J Neurovirol* 2014;20:99–102. <https://doi.org/10.1007/s13365-013-0226-2>
171. Knope KE, Doggett SL, Kurucz N, et al; National Arbovirus and Malaria Advisory Committee. Arboviral diseases and malaria in Australia, 2011–12: annual report of the National Arbovirus and Malaria Advisory Committee. *Commun Dis Intell Q Rep* 2014;38:E122–42.
172. Schwermer B, Eschle D, Bloch-Infanger C. Fever and headache after a vacation in Thailand. *Dtsch Med Wochenschr* 2017;142:1063–6.
173. Shin ES, Park O, Kong IS. Review of the incidence of Japanese encephalitis in foreign-born and Korean nationals living in the Republic of Korea, 2007–2016. *Osong Public Health Res Perspect* 2018;9:126–9. <https://doi.org/10.24171/j.phrp.2018.9.3.08>
174. Huang MZ, Novara SC, Hough-Telford C, Oliver SE, Wu CL, Pinninti S. Japanese encephalitis in an American adolescent after travel to Asia. *J Investig Med* 2015;63:395.
175. Ketel WB, Ognibene AJ. Japanese B encephalitis in Vietnam. *Am J Med Sci* 1971;261:271–9. <https://doi.org/10.1097/00000441-197105000-00006>

176. Aidem HP, Garagusi VF. Japanese B encephalitis: a case report from New York and a brief review of the literature. *Ann Intern Med* 1961;55:324–7. <https://doi.org/10.7326/0003-4819-55-2-324>
177. Long AP. Current status of immunization procedures; tetanus, and exotic diseases of military importance. *Am J Public Health Nations Health* 1948;38:485–9. <https://doi.org/10.2105/AJPH.38.4.485>
178. Perez-Pina F, Merikangas UR. Japanese B encephalitis in an American soldier returning from Korea. *N Engl J Med* 1953;249:531–2. <https://doi.org/10.1056/NEJM195309242491305>
179. Tigertt WD, Hammon WM, Berge TO, et al. Japanese B encephalitis; a complete review of experience on Okinawa 1945-1949. *Am J Trop Med Hyg* 1950;30:689–722. <https://doi.org/10.4269/ajtmh.1950.s1-30.689>
180. National Travel and Tourism Office. U.S. citizen travel to international regions. Washington, DC: US Department of Commerce, National Travel and Tourism Office. <https://travel.trade.gov>
181. Duffy MR, Reed C, Edelson PJ, et al. A survey of U.S. travelers to Asia to assess compliance with recommendations for the use of Japanese encephalitis vaccine. *J Travel Med* 2013;20:165–70. <https://doi.org/10.1111/jtm.12020>
182. Deshpande BR, Rao SR, Jentes ES, et al; The Global TravEpiNet Consortium. Use of Japanese encephalitis vaccine in U.S. travel medicine practices in Global TravEpiNet. *Am J Trop Med Hyg* 2014;91:694–8. <https://doi.org/10.4269/ajtmh.14-0062>
183. Eick-Cost AA, Hu Z, Klein TA, Putnak RJ, Jarman RG. Seroconversion to Japanese encephalitis virus among U.S. infantry forces in Korea. *Am J Trop Med Hyg* 2015;93:1052–4. <https://doi.org/10.4269/ajtmh.15-0307>
184. Ratnam I, Leder K, Black J, et al. Low risk of Japanese encephalitis in short-term Australian travelers to Asia. *J Travel Med* 2013;20:206–8. <https://doi.org/10.1111/jtm.12019>
185. Hills S, Martin R, Marfin A, Fischer M. Control of Japanese encephalitis in Asia: the time is now. *Expert Rev Anti Infect Ther* 2014;12:901–4. <https://doi.org/10.1586/14787210.2014.929498>
186. Batchelor P, Petersen K. Japanese encephalitis: a review of clinical guidelines and vaccine availability in Asia. *Trop Dis Travel Med Vaccines* 2015;1:11. <https://doi.org/10.1186/s40794-015-0013-6>
187. Markoff L. Points to consider in the development of a surrogate for efficacy of novel Japanese encephalitis virus vaccines. *Vaccine* 2000;18(Suppl 2):26–32. [https://doi.org/10.1016/S0264-410X\(00\)00038-4](https://doi.org/10.1016/S0264-410X(00)00038-4)
188. Hombach J, Solomon T, Kurane I, Jacobson J, Wood D. Report on a WHO consultation on immunological endpoints for evaluation of new Japanese encephalitis vaccines, WHO, Geneva, 2–3 September, 2004. *Vaccine* 2005;23:5205–11. <https://doi.org/10.1016/j.vaccine.2005.07.002>
189. Oya A. Japanese encephalitis vaccine. *Acta Paediatr Jpn* 1988;30:175–84. <https://doi.org/10.1111/j.1442-200X.1988.tb02516.x>
190. Hammon WM, Sather GE. Passive immunity for arbovirus infection. I. Artificially induced prophylaxis in man and mouse for Japanese (B) encephalitis. *Am J Trop Med Hyg* 1973;22:524–34. <https://doi.org/10.4269/ajtmh.1973.22.524>
191. Lubiniecki AS, Cypess RH, Hammon WM. Passive immunity for arbovirus infection. II. Quantitative aspects of naturally and artificially acquired protection in mice for Japanese (B) encephalitis virus. *Am J Trop Med Hyg* 1973;22:535–42. <https://doi.org/10.4269/ajtmh.1973.22.535>
192. Konishi E, Yamaoka M, Khin-Sane-Win, Kurane I, Takada K, Mason PW. The anamnestic neutralizing antibody response is critical for protection of mice from challenge following vaccination with a plasmid encoding the Japanese encephalitis virus premembrane and envelope genes. *J Virol* 1999;73:5527–34.
193. Beasley DW, Li L, Suderman MT, et al. Protection against Japanese encephalitis virus strains representing four genotypes by passive transfer of sera raised against ChimeriVax-JE experimental vaccine. *Vaccine* 2004;22:3722–6. <https://doi.org/10.1016/j.vaccine.2004.03.027>
194. Van Gessel Y, Klade CS, Putnak R, et al. Correlation of protection against Japanese encephalitis virus and JE vaccine (IXIARO) induced neutralizing antibody titers. *Vaccine* 2011;29:5925–31. <https://doi.org/10.1016/j.vaccine.2011.06.062>
195. Raengsakulrach B, Nisalak A, Gettayacamin M, et al. An intranasal challenge model for testing Japanese encephalitis vaccines in rhesus monkeys. *Am J Trop Med Hyg* 1999;60:329–37. <https://doi.org/10.4269/ajtmh.1999.60.329>
196. Raengsakulrach B, Nisalak A, Gettayacamin M, et al. Safety, immunogenicity, and protective efficacy of NYVAC-JEV and ALVAC-JEV recombinant Japanese encephalitis vaccines in rhesus monkeys. *Am J Trop Med Hyg* 1999;60:343–9. <https://doi.org/10.4269/ajtmh.1999.60.343>
197. Feroldi E, Capeding MR, Boaz M, Gailhardou S, Meric C, Bouckennooghe A. Memory immune response and safety of a booster dose of Japanese encephalitis chimeric virus vaccine (JE-CV) in JE-CV-primed children. *Hum Vaccin Immunother* 2013;9:889–97. <https://doi.org/10.4161/hv.23087>
198. Sohn YM, Tandan JB, Yoksan S, Ji M, Ohrr H. A 5-year follow-up of antibody response in children vaccinated with single dose of live attenuated SA14-14-2 Japanese encephalitis vaccine: immunogenicity and anamnestic responses. *Vaccine* 2008;26:1638–43. <https://doi.org/10.1016/j.vaccine.2008.01.021>
199. Monath TP, Guirakhoo F, Nichols R, et al. Chimeric live, attenuated vaccine against Japanese encephalitis (ChimeriVax-JE): phase 2 clinical trials for safety and immunogenicity, effect of vaccine dose and schedule, and memory response to challenge with inactivated Japanese encephalitis antigen. *J Infect Dis* 2003;188:1213–30. <https://doi.org/10.1086/378356>
200. Ferguson M, Johnes S, Li L, Heath A, Barrett A. Effect of genomic variation in the challenge virus on the neutralization titres of recipients of inactivated JE vaccines—report of a collaborative study on PRNT50 assays for Japanese encephalitis virus (JE) antibodies. *Biologicals* 2008;36:111–6. <https://doi.org/10.1016/j.biologicals.2007.07.002>
201. Lyons A, Kanesa-thasan N, Kuschner RA, et al. A Phase 2 study of a purified, inactivated virus vaccine to prevent Japanese encephalitis. *Vaccine* 2007;25:3445–53. <https://doi.org/10.1016/j.vaccine.2006.12.046>
202. Srivastava AK, Putnak JR, Lee SH, et al. A purified inactivated Japanese encephalitis virus vaccine made in Vero cells. *Vaccine* 2001;19:4557–65. [https://doi.org/10.1016/S0264-410X\(01\)00208-0](https://doi.org/10.1016/S0264-410X(01)00208-0)
203. Tauber E, Kollaritsch H, Korinek M, et al. Safety and immunogenicity of a Vero-cell-derived, inactivated Japanese encephalitis vaccine: a non-inferiority, phase III, randomised controlled trial. *Lancet* 2007;370:1847–53. [https://doi.org/10.1016/S0140-6736\(07\)61780-2](https://doi.org/10.1016/S0140-6736(07)61780-2)
204. Schuller E, Klade CS, Wölfel G, Kaltenböck A, Dewasthaly S, Tauber E. Comparison of a single, high-dose vaccination regimen to the standard regimen for the investigational Japanese encephalitis vaccine, IC51: a randomized, observer-blind, controlled Phase 3 study. *Vaccine* 2009;27:2188–93. <https://doi.org/10.1016/j.vaccine.2008.12.062>

205. Jelinek T, Burchard GD, Dieckmann S, et al. Short-term immunogenicity and safety of an accelerated pre-exposure prophylaxis regimen with Japanese encephalitis vaccine in combination with a rabies vaccine: A phase III, multicenter, observer-blind study. *J Travel Med* 2015;22:225–31. <https://doi.org/10.1111/jtm.12210>
206. Cramer JP, Dubischar K, Eder S, et al. Immunogenicity and safety of the inactivated Japanese encephalitis vaccine IXIARO® in elderly subjects: Open-label, uncontrolled, multi-center, phase 4 study. *Vaccine* 2016;34:4579–85. <https://doi.org/10.1016/j.vaccine.2016.07.029>
207. Woolpert T, Staples JE, Faix DJ, et al. Immunogenicity of one dose of Vero cell culture-derived Japanese encephalitis (JE) vaccine in adults previously vaccinated with mouse brain-derived JE vaccine. *Vaccine* 2012;30:3090–6. <https://doi.org/10.1016/j.vaccine.2012.02.063>
208. Erra EO, Askling HH, Rombo L, et al. A single dose of Vero cell-derived Japanese encephalitis (JE) vaccine (Ixio) effectively boosts immunity in travelers primed with mouse brain-derived JE vaccines. *Clin Infect Dis* 2012;55:825–34. <https://doi.org/10.1093/cid/cis542>
209. Kaltenböck A, Dubischar-Kastner K, Eder G, et al. Safety and immunogenicity of concomitant vaccination with the cell-culture based Japanese Encephalitis vaccine IC51 and the hepatitis A vaccine HAVRIX1440 in healthy subjects: A single-blind, randomized, controlled Phase 3 study. *Vaccine* 2009;27:4483–9. <https://doi.org/10.1016/j.vaccine.2009.05.034>
210. Dubischar-Kastner K, Eder S, Buerger V, et al. Long-term immunity and immune response to a booster dose following vaccination with the inactivated Japanese encephalitis vaccine IXIARO, IC51. *Vaccine* 2010;28:5197–202. <https://doi.org/10.1016/j.vaccine.2010.05.069>
211. Cramer JP, Jelinek T, Paulke-Korinek M, et al. One-year immunogenicity kinetics and safety of a purified chick embryo cell rabies vaccine and an inactivated Vero cell-derived Japanese encephalitis vaccine administered concomitantly according to a new, 1-week, accelerated primary series. *J Travel Med* 2016;23:1–8.
212. Schuller E, Klade CS, Heinz FX, et al. Effect of pre-existing anti-tick-borne encephalitis virus immunity on neutralising antibody response to the Vero cell-derived, inactivated Japanese encephalitis virus vaccine candidate IC51. *Vaccine* 2008;26:6151–6. <https://doi.org/10.1016/j.vaccine.2008.08.056>
213. Schuller E, Jilma B, Voicu V, et al. Long-term immunogenicity of the new Vero cell-derived, inactivated Japanese encephalitis virus vaccine IC51 Six and 12 month results of a multicenter follow-up phase 3 study. *Vaccine* 2008;26:4382–6. <https://doi.org/10.1016/j.vaccine.2008.05.081>
214. Dubischar-Kastner K. New clinical data for IXIARO Japanese encephalitis vaccine, inactivated, adsorbed. Presentation to Advisory Committee on Immunization Practices (ACIP), February 24, 2016. Atlanta, GA: US Department of Health and Human Services, CDC; 2016. <https://stacks.cdc.gov/view/cdc/60592>
215. Eder S, Dubischar-Kastner K, Firbas C, et al. Long term immunity following a booster dose of the inactivated Japanese Encephalitis vaccine IXIARO®, IC51. *Vaccine* 2011;29:2607–12. <https://doi.org/10.1016/j.vaccine.2011.01.058>
216. Taucher C, Kollaritsch H, Dubischar KL. Persistence of the immune response after vaccination with the Japanese encephalitis vaccine, IXIARO® in healthy adults: A five year follow-up study. *Vaccine* 2019;37:2529–31. <https://doi.org/10.1016/j.vaccine.2019.03.030>
217. Paulke-Korinek M, Kollaritsch H, Kundi M, Zwazl I, Seidl-Friedrich C, Jelinek T. Persistence of antibodies six years after booster vaccination with inactivated vaccine against Japanese encephalitis. *Vaccine* 2015;33:3600–4. <https://doi.org/10.1016/j.vaccine.2015.05.037>
218. Krow-Lucal E, Fischer M, Laven J, et al. Seroprotection at 12–23 months following a single dose of Vero cell culture-derived Japanese encephalitis (JE) vaccine in adults previously vaccinated with mouse brain-derived JE vaccine. The 15th Conference of the International Society of Travel Medicine; May 14–18, 2017; Barcelona, Spain.
219. Erra EO, Askling HH, Yoksan S, et al. Cross-protection elicited by primary and booster vaccinations against Japanese encephalitis: a two-year follow-up study. *Vaccine* 2013;32:119–23. <https://doi.org/10.1016/j.vaccine.2013.10.055>
220. Jelinek T, Cramer JP, Dieckmann S, et al. Evaluation of rabies immunogenicity and tolerability following a purified chick embryo cell rabies vaccine administered concomitantly with a Japanese encephalitis vaccine. *Travel Med Infect Dis* 2015;13:241–50. <https://doi.org/10.1016/j.tmaid.2015.05.008>
221. Alberer M, Burchard G, Jelinek T, et al. Co-administration of a meningococcal glycoconjugate ACWY vaccine with travel vaccines: a randomized, open-label, multi-center study. *Travel Med Infect Dis* 2014;12:485–93. <https://doi.org/10.1016/j.tmaid.2014.04.011>
222. Erra EO, Askling HH, Yoksan S, et al. Cross-protective capacity of Japanese encephalitis (JE) vaccines against circulating heterologous JE virus genotypes. *Clin Infect Dis* 2013;56:267–70. <https://doi.org/10.1093/cid/cis883>
223. Dubischar KL, Kadlecsek V, Sablan JB, et al. Immunogenicity of the inactivated Japanese encephalitis virus vaccine IXIARO in children from a Japanese encephalitis virus-endemic region. *Pediatr Infect Dis J* 2017;36:898–904. <https://doi.org/10.1097/INF.0000000000001615>
224. Kaltenböck A, Dubischar-Kastner K, Schuller E, Datla M, Klade CS, Kishore TS. Immunogenicity and safety of IXIARO (IC51) in a Phase II study in healthy Indian children between 1 and 3 years of age. *Vaccine* 2010;28:834–9. <https://doi.org/10.1016/j.vaccine.2009.10.024>
225. Jelinek T, Cromer MA, Cramer JP, et al. Safety and immunogenicity of an inactivated Vero cell-derived Japanese encephalitis vaccine (IXIARO®, JESPECT®) in a pediatric population in JE non-endemic countries: An uncontrolled, open-label phase 3 study. *Travel Med Infect Dis* 2018;22:18–24. <https://doi.org/10.1016/j.tmaid.2018.03.003>
226. Kadlecsek V, Borja-Tabora CF, Eder-Lingelbach S, et al. Antibody persistence up to 3 years after primary immunization with inactivated Japanese encephalitis vaccine IXIARO in Philippine children and effect of a booster dose. *Pediatr Infect Dis J* 2018;37:e233–40. <https://doi.org/10.1097/INF.0000000000002124>
227. Taucher C, Kundi M. Estimating the duration of protection following a booster dose of the inactivated Japanese encephalitis vaccine IXIARO®, IC51 in children. American Society of Tropical Medicine and Hygiene. 67th Annual Meeting; New Orleans, Louisiana; October 28–November 1, 2018.
228. Tauber E, Kollaritsch H, von Sonnenburg F, et al. Randomized, double-blind, placebo-controlled phase 3 trial of the safety and tolerability of IC51, an inactivated Japanese encephalitis vaccine. *J Infect Dis* 2008;198:493–9. <https://doi.org/10.1086/590116>
229. Dubischar-Kastner K, Kaltenboeck A, Klingler A, Jilma B, Schuller E. Safety analysis of a Vero-cell culture derived Japanese encephalitis vaccine, IXIARO (IC51), in 6 months of follow-up. *Vaccine* 2010;28:6463–9. <https://doi.org/10.1016/j.vaccine.2010.07.040>
230. Dubischar KL, Kadlecsek V, Sablan B Jr, et al. Safety of the inactivated Japanese encephalitis virus vaccine IXIARO in children. *Pediatr Infect Dis J* 2017;36:889–97. <https://doi.org/10.1097/INF.0000000000001623>

231. Rabe IB, Miller ER, Fischer M, Hills SL. Adverse events following vaccination with an inactivated, Vero cell culture-derived Japanese encephalitis vaccine in the United States, 2009–2012. *Vaccine* 2015;33:708–12. <https://doi.org/10.1016/j.vaccine.2014.11.046>
232. Walker WL, Hills SL, Miller ER, Fischer M, Rabe IB. Adverse events following vaccination with an inactivated, Vero cell culture-derived Japanese encephalitis vaccine in the United States, 2012–2016. *Vaccine* 2018;36:4369–74. <https://doi.org/10.1016/j.vaccine.2018.04.038>
233. Dubischar-Kastner K. Data supporting the use of a booster dose of Ixiaro. Presentation to Advisory Committee on Immunization Practices (ACIP); February 23, 2011; Atlanta, GA. <https://www.cdc.gov/vaccines/acip/meetings/index.html>
234. Lindsey NP, Staples JE, Jones JF, et al. Adverse event reports following Japanese encephalitis vaccination in the United States, 1999–2009. *Vaccine* 2010;29:58–64. <https://doi.org/10.1016/j.vaccine.2010.10.016>
235. Takahashi H, Pool V, Tsai TF, Chen RT; The VAERS Working Group. Adverse events after Japanese encephalitis vaccination: review of post-marketing surveillance data from Japan and the United States. *Vaccine* 2000;18:2963–9. [https://doi.org/10.1016/S0264-410X\(00\)00111-0](https://doi.org/10.1016/S0264-410X(00)00111-0)
236. Food and Drug Administration. Postmarketing reporting of adverse experiences. 21 C.F.R. Sect. 600.80 (2018). <https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/cfrsearch.cfm?fr=600.80>
237. Taucher C. VLA-401: JE-VC post-marketing adverse event surveillance among U.S. military personnel. October 26, 2017. Presentation to Advisory Committee on Immunization Practices (ACIP), October 26, 2017; Atlanta, GA.
238. CDC. Special situations. General best practice guidelines for immunization: Best Practices Guidance of the Advisory Committee on Immunization Practices (ACIP). <https://www.cdc.gov/vaccines/hcp/acip-recs/general-recs/special-situations.html>
239. Siraprasiri T, Sawaddiwudhipong W, Rojanasuphot S. Cost benefit analysis of Japanese encephalitis vaccination program in Thailand. *Southeast Asian J Trop Med Public Health* 1997;28:143–8.
240. Yin Z, Beeler Asay GR, Zhang L, et al; Guizhou JE Study Group. An economic evaluation of the use of Japanese encephalitis vaccine in the expanded program of immunization of Guizhou province, China. *Vaccine* 2012;30:5569–77. <https://doi.org/10.1016/j.vaccine.2012.05.068>
241. Touch S, Suraratdecha C, Samnang C, et al. A cost-effectiveness analysis of Japanese encephalitis vaccine in Cambodia. *Vaccine* 2010;28:4593–9. <https://doi.org/10.1016/j.vaccine.2010.04.086>
242. Meltzer M. Comparative analysis of strategies for JE vaccination for U.S. travelers to Asia. Presentation to the Advisory Committee on Immunization Practices (ACIP), June 21, 2018; Atlanta, GA. <https://www.cdc.gov/vaccines/acip/meetings/downloads/slides-2018-06/JE-03-Hills-Fischer-508.pdf>
243. US Department of Health and Human Services. Biosafety in microbiological and biomedical laboratories. 5th ed. HHS Publication No. (CDC) 21-1112. Washington, DC: US Department of Health and Human Services; 2018. <https://www.cdc.gov/biosafety/publications/bmb15>

Advisory Committee on Immunization Practices

Membership as of February 27, 2019

Chair: José R. Romero, MD, University of Arkansas for Medical Sciences and Arkansas Children's Hospital, Little Rock, Arkansas.

Executive Secretary: Amanda Cohn, MD, National Center for Immunization and Respiratory Diseases, CDC, Atlanta, Georgia.

Members: Robert L. Atmar, MD, Baylor College of Medicine, Houston, Texas; Kevin A. Ault, MD, University of Kansas Medical Center, Kansas City, Kansas; Henry Bernstein, DO, Zucker School of Medicine at Hofstra/Northwell Cohen Children's Medical Center, New Hyde Park, New York; Echezona Ezeanolue, MD, University of Nevada, Las Vegas, Nevada; Sharon E. Frey, MD, Saint Louis University Medical School, Saint Louis, Missouri; Stefan Gravenstein, MD, Providence Veterans Administration Hospital, Providence, Rhode Island; Paul Hunter, MD, City of Milwaukee Health Department, Milwaukee, Wisconsin; Grace M. Lee, MD, Lucile Packard Children's Hospital, Stanford University School of Medicine, Stanford, California; Veronica V. McNally, JD, Fanny Strong Foundation, West Bloomfield, Michigan; Kelly Moore, MD, Vanderbilt University School of Medicine, Nashville, Tennessee; José R. Romero, MD, University of Arkansas for Medical Sciences and Arkansas Children's Hospital, Little Rock, Arkansas; David Stephens, MD, Emory University School of Medicine, Atlanta, Georgia; Peter Szilagyi, MD, University of California, Los Angeles (UCLA), Los Angeles, California; Helen Keipp Talbot, MD, Vanderbilt University, Nashville, Tennessee; Emmanuel Walter, Jr., MD, Duke University School of Medicine, Durham, North Carolina.

Ex Officio Members: Mary Beth Hance, Centers for Medicare and Medicaid Services; Eric Deussing, MD, Department of Defense; Jane A. Kim, MD, Department of Veterans Affairs; Doran Fink, MD, Food and Drug Administration; Narayan Nair, MD, Health Resources and Services Administration; Thomas Weiser, MD, Indian Health Service; Tammy Beckham, National Vaccine Program Office; John Beigel, MD, National Institutes of Health.

Liaison Representatives: American Academy of Family Physicians, Pamela G. Rockwell, DO, Ann Arbor, Michigan; American Academy of Pediatrics, Yvonne Maldonado, MD, Stanford, California; American Academy of Pediatrics, David Kimberlin, MD, Birmingham, Alabama; American Academy of Physician Assistants, Marie-Michèle Léger, MPH, Alexandria, Virginia; American College Health Association, Susan Even, MD, Columbia, Missouri; American College of Nurse-Midwives, Carol E. Hayes, MN, Atlanta, Georgia; American College of Nurse-Midwives, Pamela M. Meharry, PhD; American College of Obstetricians and Gynecologists, Linda O'Neal Eckert, MD, Seattle, Washington; American College of Physicians, Jason M. Goldman, MD, Boca Raton, Florida; American Geriatrics Society, Kenneth Schmader, MD, Durham, North Carolina; America's Health Insurance Plans, Mark J. Netoskie, MD, Houston, Texas; American Immunization Registry Association, Rebecca Coyle, MEd, Washington DC; American Medical Association, Sandra Adamson Fryhofer, MD, Atlanta, Georgia; American Nurses Association, Charles Rittle, MPH, Pittsburgh, Pennsylvania; American Osteopathic Association, Stanley Grogg, DO, Tulsa, Oklahoma; American Pharmacists Association, Stephan L. Foster, PharmD, Memphis, Tennessee; Association of Immunization Managers, Christine Finley, MPH, Burlington, Vermont; Association for Prevention Teaching and Research, Paul W. McKinney, MD, Louisville, Kentucky; Association of State and Territorial Health Officials, Nathaniel Smith, MD, Little Rock, Arkansas; Biotechnology Industry Organization, Phyllis A. Arthur, MBA, Washington, DC; Council of State and Territorial Epidemiologists, Christine Hahn, MD, Boise, Idaho; Canadian National Advisory Committee on Immunization, Caroline Quach, MD, Montreal, Québec, Canada; Infectious Diseases Society of America, Carol J. Baker, MD, Houston, Texas; National Association of County and City Health Officials, Matthew Zahn, MD, Santa Ana, California; National Association of County and City Health Officials, Jeffrey Duchin, MD, Seattle, Washington; National Association of Pediatric Nurse Practitioners, Patricia A. Stinchfield, MS, St. Paul, Minnesota; National Foundation for Infectious Diseases, William Schaffner, MD, Nashville, Tennessee; Mexico National Immunization Council and Child Health Program, Luis Duran, MD, Mexico; National Medical Association, Patricia Whitley-Williams, MD, New Brunswick, New Jersey; Pediatric Infectious Diseases Society, Sean O'Leary, MD, Colorado; Pediatric Infectious Diseases Society, Mark H. Sawyer, MD, San Diego, California; Pharmaceutical Research and Manufacturers of America, David R. Johnson, MD, Swiftwater, Pennsylvania; Society for Adolescent Health and Medicine, Amy B. Middleman, MD, Oklahoma City, Oklahoma; Society for Healthcare Epidemiology of America, David Weber, MD, Chapel Hill, North Carolina.

ACIP Japanese Encephalitis Vaccine Work Group

Membership as of February 27, 2019

Chair: Emmanuel Walter, Jr., MD, Duke University School of Medicine, Durham, North Carolina.

Members: Robert L. Atmar, MD, Baylor College of Medicine, Houston, Texas; Elizabeth Barnett, MD, Boston Medical Center, Boston, Massachusetts; Alan Barrett, PhD, University of Texas, Medical Branch, Galveston, Texas; Joseph A. Bocchini, MD, Louisiana State University, Baton Rouge, Louisiana; Lin Chen, MD, Mount Auburn Hospital, Cambridge, Massachusetts; Eric Deussing, MD, Department of Defense, Atlanta, Georgia; Doran Fink, MD, PhD, Food and Drug Administration, Bethesda, Maryland; Michael Holbrook, MD, Battelle Memorial Institute, Frederick, Maryland; Myron Levin, MD, University of Colorado, Denver, Colorado; Anthony Marfin, MD, PATH, Seattle, Washington; Cody Meissner, MD, Tufts University, Boston, Massachusetts; Robert Schechter, MD, California Department of Public Health, California; David Shlim, MD, Jackson Hole Travel and Tropical Medicine, Jackson, Wyoming; Mary Wilson, MD, University of California, San Francisco, California.

CDC Contributors: Marc Fischer, MD, J. Erin Staples, MD, PhD, Fort Collins, Colorado; Steven Waterman, MD, San Juan, Puerto Rico; Mark Gershman, MD, Terri Hyde, MD, Michael M. McNeil, MD, Atlanta, Georgia.

Work Group Secretariat: Susan L. Hills, MBBS, CDC, Fort Collins, Colorado.

**CDC Adoption of ACIP Recommendations for *MMWR* Recommendations and Reports,
MMWR Policy Notes, and Immunization Schedules (Child/Adolescent, Adult)**

Recommendations for routine use of vaccines in children, adolescents, and adults are developed by the Advisory Committee on Immunization Practices (ACIP). ACIP is chartered as a federal advisory committee to provide expert external advice and guidance to the Director of CDC on use of vaccines and related agents for the control of vaccine-preventable diseases in the civilian population of the United States. Recommendations for routine use of vaccines in children and adolescents are harmonized to the greatest extent possible with recommendations made by the American Academy of Pediatrics (AAP), the American Academy of Family Physicians (AAFP), and the American College of Obstetricians and Gynecologists (ACOG). Recommendations for routine use of vaccines in adults are harmonized with recommendations of AAFP, ACOG, and the American College of Physicians (ACP). ACIP recommendations approved by the CDC Director become agency guidelines on the date published in the *Morbidity and Mortality Weekly Report (MMWR)*. Additional information is available at <https://www.cdc.gov/vaccines/acip>.

TABLE 1. Seroprotection rates at 1 month after a 2-dose primary series of inactivated Vero cell culture–derived Japanese encephalitis vaccine administered according to the dose and schedule approved by the Food and Drug Administration, by age group

Age group (yrs)	Study location	Seroprotection rate*			Reference
		Total	No. seroprotected	(%)	
≥18	United States, Europe	361	352	98	203
≥18	Europe	127	126	99	209
≥18	Europe	113	110	97	204
≥18	Europe	31	30	97 [†]	208
≥18	United States	92	88	96	207
18–49	United States	22	21	95	201
18–65	Europe	206	206	100	205
≥64	Europe	197	128	65	206

* Proportion with 50% plaque reduction neutralization test titer ≥10.

[†] Seroprotection measured 4–8 weeks after dose 2.

TABLE 2. Seroprotection rates and geometric mean titers for inactivated Vero cell culture–derived Japanese encephalitis vaccine administered to adults aged 18–65 years in an accelerated schedule with rabies vaccine or standard schedule with and without rabies vaccine*

Measure and time after second JE-VC dose	Primary series schedule								
	JE-VC, 0 and 7 days with rabies vaccine [†]			JE-VC, 0 and 28 days with rabies vaccine [§]			JE-VC, 0 and 28 days alone		
Seroprotection rate [¶]	Total	No. seroprotected	(%)	Total	No. seroprotected	(%)	Total	No. seroprotected	(%)
28 days	206	203	99	157	157	100	49	49	100
>300 days**	199	188	94	154	132	86	48	42	88
GMT ^{††}	GMT (95% CI)			GMT (95% CI)			GMT (95% CI)		
28 days	690 (595–801)			299 (254–352)			337 (252–451)		
>300 days**	117 (100–137)			39 (33–47)			39 (28–54)		

Sources: Food and Drug Administration. Ixiaro: Japanese encephalitis vaccine, inactivated, adsorbed [package insert]. Vienna, Austria: Valneva Austria GmbH; 2018. <https://www.fda.gov/media/75777/download>; Jelinek T, Burchard GD, Dieckmann S, et al. Short-term immunogenicity and safety of an accelerated pre-exposure prophylaxis regimen with Japanese encephalitis vaccine in combination with a rabies vaccine: A phase III, multicenter, observer-blind study. *J Travel Med* 2015;22:225–31; Cramer JP, Jelinek T, Paulke-Korinek M, et al. One-year immunogenicity kinetics and safety of a purified chick embryo cell rabies vaccine and an inactivated Vero cell-derived Japanese encephalitis vaccine administered concomitantly according to a new, 1-week, accelerated primary series. *J Travel Med* 2016;23:1–8.

Abbreviations: CI = confidence interval; GMT = geometric mean titer; JE-VC = Vero cell culture–derived Japanese encephalitis vaccine; PCEC = purified chick embryo cell.

* Per-protocol analysis.

[†] PCEC rabies vaccine administered in a 0-, 3-, 7-day schedule.

[§] PCEC rabies vaccine administered in a 0-, 7-, 28-day schedule.

[¶] Proportion with 50% plaque reduction neutralization test titer ≥10.

** Study ended on day 365.

^{††} PRNT titers <10 were imputed to 5.

TABLE 3. Seroprotection rates and geometric mean titers among adults at intervals after the first dose of a 2-dose primary series of inactivated Vero cell culture–derived Japanese encephalitis vaccine

Measure and study site	6 mos			12–15 mos			24 mos			60 mos		
	Total	No. seroprotected	(%)									
Austria, Germany, Romania	181	172	(95)	181	151	(83)	181	148	(82)	151	124	(82)
Germany, Northern Ireland	116	96	(83)	116	67	(58)	116	56	(48)	—	—	—
Austria, Germany	—	—	—	198	137	(69)	—	—	—	—	—	—
GMT	GMT (95% CI)											
Austria, Germany, Romania	84 (71–98)			41 (34–49)			44 (37–53)			43 (36–53)		
Germany, Northern Ireland	47 (37–59)			18 (14–23)			16 (13–21)			—		
Austria, Germany	—			23 (19–27)			—			—		

Sources: Food and Drug Administration. Ixiaro: Japanese encephalitis vaccine, inactivated, adsorbed [package insert]. Vienna, Austria: Valneva Austria GmbH; 2018. <https://www.fda.gov/media/75777/download>; Dubischar-Kastner K, Eder S, Buerger V, et al. Long-term immunity and immune response to a booster dose following vaccination with the inactivated Japanese encephalitis vaccine IXIARO, IC51. *Vaccine* 2010;28:5197–202. Schuller E, Jilma B, Voicu V, et al. Long-term immunogenicity of the new Vero cell-derived, inactivated Japanese encephalitis virus vaccine IC51 Six and 12 month results of a multicenter follow-up phase 3 study. *Vaccine* 2008;26:4382–6. Dubischar-Kastner K. New clinical data for IXIARO Japanese encephalitis vaccine, inactivated, adsorbed. Presentation to Advisory Committee on Immunization Practices (ACIP), February 24, 2016. Atlanta, GA: US Department of Health and Human Services, CDC; 2016. <https://stacks.cdc.gov/view/cdc/60592>; Eder S, Dubischar-Kastner K, Firbas C, et al. Long term immunity following a booster dose of the inactivated Japanese Encephalitis vaccine IXIARO®, IC51. *Vaccine* 2011;29:2607–12.

Abbreviations: CI = confidence interval; GMT = geometric mean titer.

* Proportion with 50% plaque reduction neutralization test titer ≥10.

TABLE 4. Seroprotection rates and geometric mean titers among adults at intervals after first dose of a 2-dose primary series of inactivated Vero cell culture–derived Japanese encephalitis vaccine, by tickborne encephalitis vaccination status

Measure	6 mos			12 mos			24 mos			60 mos		
	Total	No. seroprotected	(%)	Total	No. seroprotected	(%)	Total	No. seroprotected	(%)	Total	No. seroprotected	(%)
TBE vaccine†	89	86	(97)	89	82	(92) [§]	86	78	(91) [§]	78	67	(86) [§]
No TBE vaccine	92	86	(93)	92	69	(75)	78	53	(68)	47	30	(64)
GMT	GMT			GMT			GMT			GMT		
TBE vaccine†	96			48			56			45		
No TBE vaccine	73			35			33			29		

Sources: Dubischar-Kastner K. New clinical data for IXIARO Japanese encephalitis vaccine, inactivated, adsorbed. Presentation to Advisory Committee on Immunization Practices (ACIP), February 24, 2016. Atlanta, GA: US Department of Health and Human Services, CDC; 2016. <https://stacks.cdc.gov/view/cdc/60592>; Taucher C, Kollaritsch H, Dubischar KL. Persistence of the immune response after vaccination with the Japanese encephalitis vaccine, IXIARO® in healthy adults: A five year follow-up study. *Vaccine* 2019;37:2529–31.

Abbreviations: GMT = geometric mean titer; TBE = tickborne encephalitis.

* Proportion with 50% plaque reduction neutralization test titer ≥10.

† TBE vaccine received before or after Vero cell culture–derived Japanese encephalitis vaccine.

§ Nonoverlapping 95% confidence intervals (calculated according to the method recommended by Altman, developed by Wilson) for seroprotection rates for TBE and no TBE vaccine groups.

TABLE 5. Seroprotection rates and geometric mean titers before and after a booster dose of inactivated Vero cell culture–derived Japanese encephalitis vaccine administered 15 months after the first dose of a 2-dose primary series

Measure	0 days			1 mo			6 mos			12 mos			76 mos		
	Total	No. seroprotected	(%)	Total	No. seroprotected	(%)	Total	No. seroprotected	(%)	Total	No. seroprotected	(%)	Total	No. seroprotected	(%)
Seroprotection rate*	198	137	(69)	198	198	(100)	197	194	(98)	194	191	(98)	67	64	(96)
GMT	GMT (95% CI)			GMT (95% CI)			GMT (95% CI)			GMT (95% CI)			GMT (95% CI)		
	23 (19–27)			900 (742–1,091)			487 (391–608)			361 (295–444)			148 (107–207)		

Sources: Eder S, Dubischar-Kastner K, Firbas C, et al. Long term immunity following a booster dose of the inactivated Japanese Encephalitis vaccine IXIARO®, IC51. *Vaccine* 2011;29:2607–12; Paulke-Korinek M, Kollaritsch H, Kundi M, Zwazl I, Seidl-Friedrich C, Jelinek T. Persistence of antibodies six years after booster vaccination with inactivated vaccine against Japanese encephalitis. *Vaccine* 2015;33:3600–4.

Abbreviations: CI = confidence interval; GMT = geometric mean titer.

* Proportion with 50% plaque reduction neutralization test titer ≥10.

TABLE 6. Seroprotection rates in children at 1 month after a 2-dose primary series of inactivated Vero cell culture–derived Japanese encephalitis vaccine administered according to the dose and schedule approved by the Food and Drug Administration*

Study site	Age group	Seroprotection rate [†]					
		0.25-mL JE-VC dose			0.5-mL JE-VC dose		
		Total	No. seroprotected	(%)	Total	No. seroprotected	(%)
Philippines	2 mos–17 yrs	148	147	(99) [§]	237	237	(100)
India	1–2 yrs	23	22	(96)	—	—	— [¶]
United States, Europe, Australia	2 mos–17 yrs	5	5	(100)	57	57	(100)

Sources: Food and Drug Administration. Ixiaro: Japanese encephalitis vaccine, inactivated, adsorbed [package insert]. Vienna, Austria: Valneva Austria GmbH; 2018. <https://www.fda.gov/media/75777/download>; Dubischar KL, Kadlec V, Sablan JB, et al. Immunogenicity of the inactivated Japanese encephalitis virus vaccine IXIARO in children from a Japanese encephalitis virus-endemic region. *Pediatr Infect Dis J* 2017;36:898–904; Kaltenböck A, Dubischar-Kastner K, Schuller E, Datla M, Klade CS, Kishore TS. Immunogenicity and safety of IXIARO (IC51) in a Phase II study in healthy Indian children between 1 and 3 years of age. *Vaccine* 2010;28:834–9; Jelinek T, Cromer MA, Cramer JP, et al. Safety and immunogenicity of an inactivated Vero cell–derived Japanese encephalitis vaccine (IXIARO®, JESPECT®) in a pediatric population in JE non-endemic countries: An uncontrolled, open-label phase 3 study. *Travel Med Infect Dis* 2018;22:18–24.

Abbreviations: FDA = Food and Drug Administration; JE-VC = Vero cell culture–derived Japanese encephalitis vaccine.

* For children aged 2 months–2 years, 2 doses (0.25 mL each) administered 28 days apart; for children aged 3–17 years, 2 doses (0.5 mL each) administered 28 days apart.

[†] Proportion with 50% plaque reduction neutralization test titer ≥10.

[§] Of an additional 98 children aged 3–11 years who received 2 doses of 0.25 mL, 94 (96%) were seroprotected at 1 month after the second dose.

[¶] Of 21 children aged 1–2 years who received 2 doses of 0.5 mL, 20 (95%) were seroprotected at 1 month after the second dose; the FDA-approved dose for children aged 1–2 years is 0.25 mL.

TABLE 7. Seroprotection rates and geometric mean titers among children aged 14 months–17 years in the Philippines before and after a booster dose of inactivated Vero cell culture–derived Japanese encephalitis vaccine administered 11 months after the second dose of a 2-dose primary series

Measure	0 days		1 mo		12 mos		24 mos	
	No. seroprotected (n = 148)	(%)	No. seroprotected (n = 148)	(%)	No. seroprotected (n = 147)	(%)	No. seroprotected (n = 143)	(%)
Seroprotection rate*	139	(94)	148	(100)	147	(100)	143	(100)
GMT	GMT (95% CI)		GMT (95% CI)		GMT (95% CI)		GMT (95% CI)	
	53 (45–64)		2,067 (1,671–2,556)		428 (335–546)		350 (279–440)	

Source: Kadlec V, Borja-Tabora CF, Eder-Lingelbach S, et al. Antibody persistence up to 3 years after primary immunization with inactivated Japanese encephalitis vaccine IXIARO in Philippine children and effect of a booster dose. *Pediatr Infect Dis J* 2018;37:e233–40.

Abbreviations: CI = confidence interval; GMT = geometric mean titer.

* Proportion with 50% plaque reduction neutralization test titer ≥10.

TABLE 8. Local and systemic adverse events in adults occurring within 7 days after vaccination with inactivated Vero cell culture–derived Japanese encephalitis vaccine or inactivated mouse brain–derived Japanese encephalitis vaccine*

Adverse events	JE-VC [†]	JE-MB [§]
Severe local adverse events	No. (%) (n = 421)	No. (%) (n = 427)
Redness	4 (1)	46 (11)
Swelling	3 (1)	23 (5)
Hardness	4 (1)	25 (5)
Any [¶]	9 (2)	59 (14)
Systemic adverse events	No. (%) (n = 428)	No. (%) (n = 435)
Headache	113 (26)	125 (29)
Myalgia	88 (21)	69 (16)
Influenza-like illness	54 (13)	55 (13)
Fatigue	54 (13)	48 (11)

Source: Tauber E, Kollaritsch H, Korinek M, et al. Safety and immunogenicity of a Vero-cell-derived, inactivated Japanese encephalitis vaccine: a non-inferiority, phase III, randomised controlled trial. *Lancet* 2007;370:1847–53.

Abbreviations: JE-MB = mouse brain–derived Japanese encephalitis vaccine; JE-VC = Vero cell culture–derived Japanese encephalitis vaccine.

* Analysis includes all participants who entered into the study and received ≥1 dose of vaccine.

[†] Two doses administered at days 0 and 28, with one dose of placebo at 7 days.

[§] Three doses administered at days 0, 7, and 28.

[¶] p<0.01 calculated using Fisher’s exact test for difference between two vaccines.

The *Morbidity and Mortality Weekly Report (MMWR)* Series is prepared by the Centers for Disease Control and Prevention (CDC) and is available free of charge in electronic format. To receive an electronic copy each week, visit *MMWR* at <https://www.cdc.gov/mmwr/index.html>.

Readers who have difficulty accessing this PDF file may access the HTML file at https://www.cdc.gov/mmwr/volumes/68/rr/rr6802a1.htm?s_cid=rr6802a1_w. Address all inquiries about the *MMWR* Series, including material to be considered for publication, to Executive Editor, *MMWR* Series, Mailstop E-90, CDC, 1600 Clifton Rd., N.E., Atlanta, GA 30329-4027 or to mmwrq@cdc.gov.

All material in the *MMWR* Series is in the public domain and may be used and reprinted without permission; citation as to source, however, is appreciated. *MMWR* and *Morbidity and Mortality Weekly Report* are service marks of the U.S. Department of Health and Human Services.

Use of trade names and commercial sources is for identification only and does not imply endorsement by the U.S. Department of Health and Human Services.

References to non-CDC sites on the Internet are provided as a service to *MMWR* readers and do not constitute or imply endorsement of these organizations or their programs by CDC or the U.S. Department of Health and Human Services. CDC is not responsible for the content of these sites. URL addresses listed in *MMWR* were current as of the date of publication.

ISSN: 0149-2195 (Print)