Newborn Screening Quality Assurance Program
Second-tier Congenital Adrenal Hyperplasia (CAH)
Proficiency Testing Program (PT)

In co-sponsorship with Association of Public Health Laboratories (APHL)
Provided by the Newborn Screening and Molecular Biology Branch
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Quarterly Report
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Report Authorization

This report has been reviewed and authorized by Dr. Joanne Mei, Laboratory Chief, Newborn Screening Quality Assurance Program.

Confidentiality Statement

NSQAP participant information and evaluations are strictly confidential and shared only with individual participants, unless written authorization for release is received.

Introduction

This report summarizes data collected within the specified reporting period for the Quarter 1, 2019 Second-tier Congenital Adrenal Hyperplasia (CAH) Proficiency Testing (PT) Program event. Reports are distributed to all participants, state laboratory directors, and program colleagues by request. The tables within this report provide certification information for the PT specimen panel, statistical analysis for reported quantitative data, and the frequency distribution summaries for expected interpretations. An evaluation of your submitted data is attached to this summary.

Certification of PT Specimens

The dried blood spot (DBS) PT specimens were prepared at 50% hematocrit, with different enrichments of five biomarkers for congenital adrenal hyperplasia (CAH); 17 α-hydroxyprogesterone (17OHP), 4-androstenedione (4AD), cortisol (Cort), 11-deoxycortisol (11D), 21-deoxycortisol (21D). Expected values (sum of endogenous and enrichment values) were determined by EIA (17OHP only) and LC-MS/MS. For determination of the Clinical Assessment (CA) NSQAP applies the formula: clinical ratio = ([17OHP] + [4AD])/[CORT]. A cutoff of 1.0 is used to assess whether the specimen is Within Normal Limits (1) or Outside Normal Limits (2).
Table 1. Expected Values (ng/mL serum) and Expected Clinical Assessments (CA)

<table>
<thead>
<tr>
<th>Specimen</th>
<th>EIA 17OHP</th>
<th>EIA CA</th>
<th>LC-MS/MS 17OHP</th>
<th>LC-MS/MS 4AD</th>
<th>LC-MS/MS Cort</th>
<th>LC-MS/MS 11D</th>
<th>LC-MS/MS 21D</th>
<th>LC-MS/MS Clinical Rato</th>
<th>LC-MS/MS CA</th>
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</thead>
<tbody>
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<td>119A1</td>
<td>41.2</td>
<td>2</td>
<td>51.5</td>
<td>26.5</td>
<td>101.5</td>
<td>11.5</td>
<td>1.5</td>
<td>0.8</td>
<td>1</td>
</tr>
<tr>
<td>119A2</td>
<td>79.2</td>
<td>2</td>
<td>91.5</td>
<td>37.5</td>
<td>21.5</td>
<td>6.5</td>
<td>41.5</td>
<td>6.0</td>
<td>2</td>
</tr>
<tr>
<td>119A3</td>
<td>81.3</td>
<td>2</td>
<td>91.5</td>
<td>37.5</td>
<td>21.5</td>
<td>6.5</td>
<td>41.5</td>
<td>6.0</td>
<td>2</td>
</tr>
<tr>
<td>119A4</td>
<td>82.4</td>
<td>2</td>
<td>81.5</td>
<td>26.5</td>
<td>121.5</td>
<td>21.5</td>
<td>1.5</td>
<td>0.9</td>
<td>1</td>
</tr>
<tr>
<td>119A5</td>
<td>8.4</td>
<td>1</td>
<td>11.5</td>
<td>21.5</td>
<td>41.5</td>
<td>51.5</td>
<td>11.5</td>
<td>0.8</td>
<td>1</td>
</tr>
</tbody>
</table>

1 = Within Normal Limits  
2 = Outside Normal Limits  
NE = Not Evaluated

Distribution of PT Specimens

On January 15, 2019, a PT panel of five DBS specimens was distributed eight domestic laboratories and 24 international laboratories.

Participant Results

Quantitative Data

We received results from 28 participants by the data reporting deadline. Laboratories were asked to report concentrations of 17OHP, 4AD, Cort, 11D and 21D analyzed by Second-tier tandem mass spectrometry (LC-MS/MS) and enzyme immunoassay (EIA) (optional). For the statistical summary analysis, we did not include data that were outside the 99% confidence interval.

All data are presented in units of ng/mL serum. Participants whose methods yield data in nM whole blood units were asked to multiply by the following factors for conversion to serum concentration: 0.66 (17OHP), 0.57 (4AD), 0.72 (CORT), and 0.69 (11D and 21D). Data that are not submitted in the requested units (ng/mL serum) are not accepted. Conversion factors are provided on the CAHPT Data Report Form.

Twenty-eight laboratories reported results using LC-MS/MS. Twenty laboratories reported EIA results. The expected analyte concentration values were based on CDC expected values. Overall statistics from EIA (Table 2) and LC-MS/MS (Tables 3a-b) methods were combined so as to not identify an individual laboratory.

Table 2. Overall statistics – 17OHP (ng/mL serum) by EIA

<table>
<thead>
<tr>
<th>Specimen</th>
<th>N</th>
<th>Mean</th>
<th>SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>119A1</td>
<td>20</td>
<td>54.7</td>
<td>10.5</td>
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<tr>
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<td>20</td>
<td>97.9</td>
<td>20.4</td>
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<td>119A3</td>
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<td>96.6</td>
<td>20.4</td>
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<td>119A4</td>
<td>20</td>
<td>89.6</td>
<td>19.7</td>
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<tr>
<td>119A5</td>
<td>20</td>
<td>12.3</td>
<td>5.4</td>
</tr>
</tbody>
</table>
Table 3a. Overall statistics – 17OHP, 4AD, Cort, (ng/mL serum) by LC-MS/MS

<table>
<thead>
<tr>
<th>Specimen</th>
<th>17OHP N</th>
<th>17OHP Mean</th>
<th>17OHP SD</th>
<th>4AD N</th>
<th>4AD Mean</th>
<th>4AD SD</th>
<th>Cort N</th>
<th>Cort Mean</th>
<th>Cort SD</th>
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<tbody>
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<td>58.11</td>
<td>16.1</td>
<td>28</td>
<td>26.67</td>
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<td>28</td>
<td>117.59</td>
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<tr>
<td>119A2</td>
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<td>100.63</td>
<td>20.0</td>
<td>28</td>
<td>44.81</td>
<td>12.0</td>
<td>28</td>
<td>25.09</td>
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<td>119A3</td>
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<td>101.87</td>
<td>20.8</td>
<td>28</td>
<td>45.37</td>
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<td>22.5</td>
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<td>7.7</td>
<td>28</td>
<td>144.94</td>
<td>42.2</td>
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<td>24</td>
<td>13.51</td>
<td>5.9</td>
<td>24</td>
<td>21.03</td>
<td>5.1</td>
<td>24</td>
<td>45.56</td>
<td>13.8</td>
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</table>

Table 3b. Overall statistics –11D, 12D (ng/mL serum) by LC-MS/MS

<table>
<thead>
<tr>
<th>Specimen</th>
<th>11D N</th>
<th>11D Mean</th>
<th>11D SD</th>
<th>21D N</th>
<th>21D Mean</th>
<th>21D SD</th>
</tr>
</thead>
<tbody>
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<td>3.1</td>
<td>17</td>
<td>0.87</td>
<td>1.2</td>
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<td>4.1</td>
<td>20</td>
<td>45.52</td>
<td>7.4</td>
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<td>45.90</td>
<td>6.6</td>
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<td>25.37</td>
<td>6.3</td>
<td>17</td>
<td>0.58</td>
<td>0.6</td>
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<tr>
<td>119A5</td>
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<td>63.95</td>
<td>17.8</td>
<td>17</td>
<td>11.02</td>
<td>2.2</td>
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</table>

Qualitative Clinical Assessments

Qualitative assessments may differ by participant because of specific assessment practices. The frequency distribution of participants’ Clinical Assessments for screening results is shown in Table 4.

Most programs use a clinical ratio to determine if samples are normal or abnormal. Specimens with a calculated ratio less than the cutoff are considered “within normal limits”; those specimens with a calculated ratio greater than the cutoff are evaluated as “outside normal limits”. LC-MS/MS cutoff values are summarized in Table 5.

Table 4. Frequency Distribution of Participants’ Clinical Assessments (LC-MS/MS)

<table>
<thead>
<tr>
<th>Specimen</th>
<th>Within Normal Limits (WNL)</th>
<th>Outside Normal Limits (ONL)</th>
<th>Not Reported</th>
</tr>
</thead>
<tbody>
<tr>
<td>119A1</td>
<td>25</td>
<td>3</td>
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</tr>
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<td>119A2</td>
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<td>0</td>
</tr>
<tr>
<td>119A3</td>
<td>0</td>
<td>28</td>
<td>0</td>
</tr>
<tr>
<td>119A4</td>
<td>24</td>
<td>4</td>
<td>0</td>
</tr>
<tr>
<td>119A5*</td>
<td>21</td>
<td>6</td>
<td>1</td>
</tr>
</tbody>
</table>

*Specimen 119A5 was not evaluated due to lack of 80% consensus among reporting laboratories.
Table 5. LC-MS/MS Clinical Ratio Cutoff Values

<table>
<thead>
<tr>
<th>Specimen</th>
<th>All Laboratories</th>
<th>Domestic</th>
<th>International</th>
</tr>
</thead>
<tbody>
<tr>
<td>MEAN</td>
<td>1.68</td>
<td>1.05</td>
<td>1.96</td>
</tr>
<tr>
<td>MODE</td>
<td>1.00</td>
<td>1.00</td>
<td>1.00</td>
</tr>
<tr>
<td>MIN</td>
<td>0.10</td>
<td>0.90</td>
<td>0.10</td>
</tr>
<tr>
<td>MAX</td>
<td>9.00</td>
<td>1.40</td>
<td>9.00</td>
</tr>
</tbody>
</table>

Evaluations
Of the evaluated specimens, participants reported seven False-positive results and no False-negative results based on the LC-MS/MS final Clinical Assessment.

Specimen 119A5 was considered “not-evaluated” due to lack of 80% consensus among reporting laboratories.

Future Shipments
The Newborn Screening Quality Assurance Program will ship next quarter’s PT specimens for CAHPT on June 25, 2019.

Direct Inquiries
If you have any comments or questions about CAHPT MS/MS analysis, contact Dr. Joanne V. Mei at 770-488-7945 or by email at jvm0@cdc.gov
For data reporting questions, contact Irene Williams at nsqapdmt@cdc.gov

The content of this report may also be located on our website at: https://www.cdc.gov/labstandards/nsqap_reports.html
This NEWBORN SCREENING QUALITY ASSURANCE PROGRAM report is an internal publication distributed to program participants and selected program colleagues. The laboratory quality assurance program is a project cosponsored by the Centers for Disease Control and Prevention (CDC) and the Association of Public Health Laboratories.

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