Prenatal Screening for Group B *Streptococcus*  
(Non-pigmented broth, plate subculture only for isolation and identification)

**Annual Review:**

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**Revision History:**

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Approval Signatures:

Approved by: _______________________________ Date: ________________
Author

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Print Name and Title

Approved by: _______________________________ Date: ________________
Supervisor/Technical Specialist-Director

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Print Name and Title

Approved by: _______________________________ Date: ________________
Quality Manager

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Print Name and Title
Purpose/Principle:

The purpose of this document is to provide instructions for performing prenatal screening for Group B *Streptococcus* from appropriately collected vaginal/rectal specimens using enriched non-pigmented broth followed by plate culture.

Scope:

This document applies to laboratory assistants, technicians and technologists within the Division of Microbiology at XXXX.

Related documents:

1. Provider collection procedure for Prenatal Screening for Group B *Streptococcus* - # xxxx. Note: Procedure should specify the acceptable specimen is a vaginal-rectal swab and that swabs from other sites (e.g. cervical or perianal swabs only) are not acceptable. Procedure should list acceptable type of swab and transport medium.

2. Quality Control procedure for tube and plated media - # xxxx.


4. Notification procedure for positive Prenatal Screening for Group B *Streptococcus* - # xxxx.

5. Procedure for susceptibility testing of Group B *Streptococcus* - # xxxx.
Responsibility:

1. It is the responsibility of the technical staff of the microbiology laboratory to review and assure they are familiar with the processes outlined in this document which are within their job duties.

2. It is the responsibility of the technical specialist to keep this procedure updated and current with national standards/guidelines/recommendations.

3. It is the responsibility of the microbiology supervisor to accurately train and assess the competency of all microbiology staff involved in performing this procedure.

4. It is the responsibility of the microbiology data technologist to assure that reporting of all results is appropriate in all computer systems.

5. It is the responsibility of the technical director to approve this procedure on an annual basis.

Definitions:

N/A

Equipment/Materials:

1. Trans-Vag broth supplemented with 5% defibrinated sheep blood, LIM broth (or equivalent)

2. Agar plates for subculture (available from a variety of manufacturers): Sheep blood agar, neomycin-nalidixic acid (NNA) agar, Columbia colistin and naladixic acid (CNA) agar with 5% sheep blood, or other selective Streptococcus agar; GBS Detect™ agar (Hardy Diagnostics, Santa Maria, CA); CHROMagar™ StrepB
Prenatal Screening for Group B *Streptococcus*: Non-pigmented broth, subculture only

(distributed by Gibson Laboratories, LLC, Lexington, KY), ChromID Strepto B agar (bioMérieux, Inc., Durham, NC), or other chromogenic medium designed for detection of Group B *Streptococcus*, including non-ß-hemolytic strains.

3. Sterile loops
4. Incinerator
5. Incubator

Quality Control:

Follow Quality Control procedures for tube and plated media - # xxxx.

Safety Precautions:

Use standard precautions when performing this procedure.

Procedure: (Appendix includes one-page flowchart)

1. Remove patient specimen (swab; swabs) from transport medium. Specimens should be processed within 4 days of collection. Specimens requiring more than 24 hr of transport time should be refrigerated until received and processed by the laboratory.

2. Inoculate swab(s) into a recommended selective enrichment broth medium, such as Trans-Vag broth supplemented with 5% defibrinated sheep blood, LIM broth or equivalent. [Before inoculating into broth, laboratories may choose to direct plate by rolling the swab(s) on a sheep blood agar plate, NNA agar, CNA agar, other selective *Streptococcus* agar. If Group B *Streptococcus* is identified from the primary plate, report Group B streptococcus positive on the culture report. If Group B
Streptococcus is not identified from the primary plate, plate should be discarded, and enriched broth processed as per protocol below and results reported out from enriched broth.

3. Incubate inoculated selective broth for a minimum of 18-24 hours at 35-37°C in ambient air or 5% CO₂ as recommended by the manufacturer. The broth should display visible turbidity as an indicator of adequate specimen collection. A clear broth after 24 hours of incubation indicates failure to obtain a proper sample and a new specimen should be requested.

4. Subculture the incubated broth to a sheep blood agar plate, NNA agar, CNA agar, other selective Streptococcus agar; GBS Detect™ agar, or chromogenic medium such as CHROMagar™ StrepB, ChromID Strepto B agar that will detect non-hemolytic Group B Streptococcus and incubate a minimum of 18-24 hour at 35-37°C in 5% CO₂ or as recommended by the manufacturer.

5. Inspect and identify colonies characteristic of Group B Streptococcus (i.e., gram-positive cocci, catalase negative, narrow zone of β-hemolysis or no hemolysis) using routine laboratory identification protocols. (Refer to procedure # xxxx – Procedure for the identification of streptococci).

6. If Group B Streptococcus is not identified, reincubate the agar plate for an additional overnight incubation to identify suspected Group B Streptococcus organisms.

7. Report as negative for Group B Streptococcus when no Group B Streptococcus has been recovered.

8. If Group B Streptococcus is identified, report Group B streptococcus positive on the culture report.
9. If the positive specimen is from a penicillin-allergic patient at high risk for anaphylaxis, perform susceptibilities for clindamycin and erythromycin, including inducible clindamycin resistance, using appropriate laboratory susceptibility procedures. (Refer to protocol # xxxx – Procedure for susceptibility testing of Group B Streptococcus.)

Format:

N/A

Reference:

Flowchart #1: Using non-pigmented broth and only subculture for isolation and identification of GBS

**Direct plate & incubate 24-48 hrs at 35-37°C**
(if negative, discard plate and report results from enriched specimen)

- **Vaginal-rectal swab**
  - Place in non-pigmented enrichment broth
  - Incubate in enrichment broth x 18-24 hrs at 35-37°C
  - Subculture to plate & incubate 18-24 hrs at 35-37°C

**Identify GBS by recommended method**

- **GBS+**
  - Report as GBS+
  - If PCN-allergic & at high risk for anaphylaxis, subculture to plate and incubate 18-24 hrs at 35-37°C to conduct susceptibility testing
- **GBS−**
  - Re-incubate overnight
  - **GBS+**
  - **GBS−**
  - Report as GBS−

Without enrichment, the false negative rate can be as high as 50%

Subcultures that are negative after the 1st incubation should be incubated again overnight and re-examined

*Typical characteristics include: β-hemolysis, gram stain (+, catalase −, GBS serology, CAMP+, hippurate,+.*

For more information, see: http://www.cdc.gov/groupBstrep/cib