60

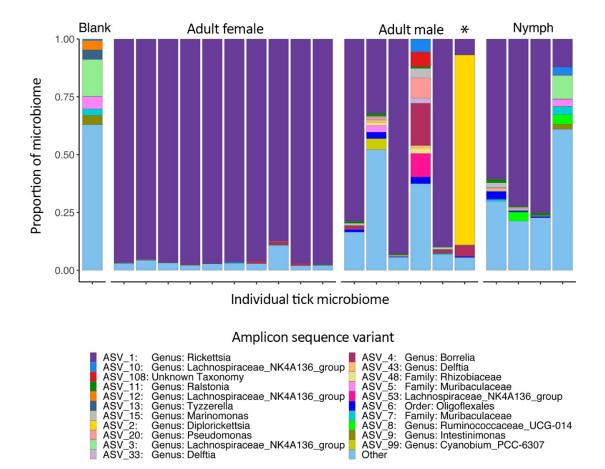
Diplorickettsia Bacteria in an Ixodes scapularis Tick, Vermont, USA

Appendix

parC reverse

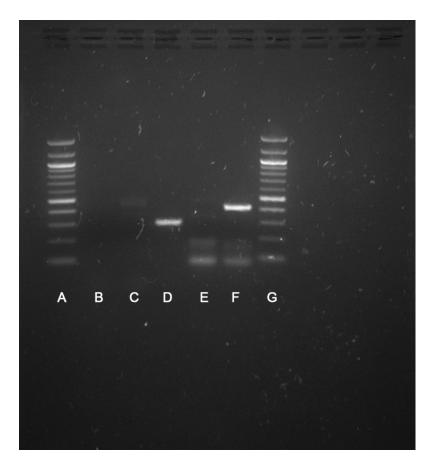
Site	Latitude, °	Longitude, °	Mean elevation, m
Sunny Hollow Colchester	44.518353	-73.17112	93
Shelbourne Town Forest	44.369066	-73.143851	122
New Haven	44.113699	-73.156602	119
Chipman Hill	44.023385	-73.162146	221
Chipman Hill (Backside)	44.026342	-73.159889	213
Orwell	43.818445	-73.264537	140
Appendix Table 2. Primers Primer	s used for PCR of novel <i>Diploricke</i> Melting temperature, °C	ttsia bacteria found in <i>Ixodes scaularis</i> tick, Primer sequence	Vermont, USA Amplicon length, bt
fts Y forward	60	TCATCGATGGCCAAGCTGTT	386
fts Y reverse	60	TTTCACCTTCGCGAGCTCTT	
parC forward	60	ACTCGACCATCCAAAAGCGT	272
paro ioiwaru	00		

TCACCAAACGTGTCGGTTGT



Appendix Figure 1. Microbiome of each tick via 16s RNA sequencing. Each vertical bar represents an individual tick, colored by the proportion of the microbiome made up of each amplicon sequence variant (ASV). For the top 22 ASVs, taxa information is shown as the lowest order to which the ASV can be

identified. Asterisk indicates the tick in which Diplorickettsia was identified.



Appendix Figure 2. Presence of *Diplorickettsia* is confirmed by PCR for the *parC* and *ftsY* gene. A) New England Biolabs (https://www.neb.com) 100bp ladder; B) negative control, using the *parC* primers but no DNA; C) *Diplorickettsia*-negative tick with *parC*; D) *Diplorickettsia*-positive tick with *parC* (272 bp amplicon); E) *Diplorickettsia*-negative tick with *ftsY*; F) *Diplorickettsia*-positive tick with *ftsY* (386 bp amplicon); G) NEB 100 bp ladder. We used NEB Phusion 2X master mix for PCR, using 25 µL reactions, 1 µL of tick prep DNA (unquantified); final primer concentration for each sample was 0.4 µM. PCR temperatures were 98°C for 30 sec, followed by 35 cycles of 98°C for 10 sec, 61°C for 15 sec, 72°C for 20 sec, followed by 72°C for 10 min.