Invasive Serotype 35B Pneumococci Including an Expanding Serotype Switch Lineage, United States, 2015–2016

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We used whole-genome sequencing to characterize 199 nonvaccine serotype 35B pneumococcal strains that caused invasive pneumococcal disease (IPD) in the United States during 2015-2016 and related these findings to previous serotype 35B IPD data obtained by Active Bacterial Core surveillance. Penicillin-nonsusceptible 35B IPD increased during post-pneumococcal 7-valent conjugate vaccine years (2001-2009) and increased further after implementation of pneumococcal 13-valent conjugate vaccine in 2010. This increase was caused primarily by the 35B/sequence type (ST) 558 lineage. 35B/ST558 and vaccine serotype 9V/ST156 lineages were implicated as cps35B donor and recipient, respectively, for a single capsular switch event that generated emergent 35B/ST156 progeny in 6 states during 2015-2016. Three additional capsular switch 35B variants were identified, 2 of which also involved 35B/ST558 as cps35B donor. Spread of 35B/ ST156 is of concern in view of past global predominance of pathogenic ST156 vaccine serotype strains. Protection against serotype 35B should be considered in next-generation pneumococcal vaccines.

A lthough the dramatic protective effect of the pneumococcal 7-valent conjugate vaccine (PCV7) against invasive pneumococcal disease (IPD) persisted a full decade after its introduction in the United States in 2000, the emergence of 19A and other non-PCV7 serotypes reduced the overall benefit (1,2). Before PCV7 implementation, we observed only 2 different 35B lineages within Active Bacterial Core surveillance (ABCs) (3), a populationbased, multistate program that assesses the effect of invasive bacterial infections and is part of the Emerging Infections Program network of the Centers for Disease Control and Prevention (CDC; Atlanta, GA, USA) (http://www. cdc.gov/abcs/index.html). Both lineages were relatively

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rare causes of IPD but were geographically widespread in the United States before and after PCV7 introduction (3). One 35B lineage was antimicrobial-susceptible and multilocus sequence type (MLST) 452 (35B/ST452), and the second strain was penicillin-nonsusceptible 35B/ST558. During 1995–2001, the penicillin-nonsusceptible 35B/ ST558 lineage, which had resistant MICs of 0.25–2.0 μ g/ mL, accounted for 69% of serotype 35B ABCs isolates (3). During 1999–2007 in the United States, the proportion of penicillin-nonsusceptible IPD isolates within serotype 35B increased to 84%; 35B/ST558 accounted for this increase (4). Consistent with this observation was a 9-fold increase in carriage of 35B/ST558 in young children in the Atlanta, Georgia, area (5).

After introduction of the 13-valent conjugate vaccine (PCV13) in 2010, 35B became the most common serotype in ABCs, associated with MICs $\geq 2 \mu g/mL$ for penicillin and amoxicillin in pediatric isolates (6) and in the adult population (B. Beall, unpub. data). Consistent with its status as a major cause of IPD in the post-PCV13 era, the 35B/ST558 lineage is currently commonly found in disease and asymptomatic pneumococcal carriage in many countries (7–11).

We provide a whole-genome sequence (WGS) pipeline–based resolution and description of current 35B lineages within current ABCs surveillance (6,12), including an invasive 35B variant of the historically successful ST156 lineage. Recently, we identified 2 different 35B isolates recovered during 2009 and 2012 that each appeared to have arisen through a unique capsular switch event involving the same 2 parental strains. This observation was made on the basis of the penicillin-binding protein (PBP) gene types flanking the 35B biosynthetic locus (*cps35B*) in each of the variants (6).

Only the 35B/ST156 variant detected during 2012 has emerged and has been detected within 6 states. ST156 has a remarkable history of conjugate vaccine evasion. Formerly the primary genotype of PCV7

serotype 9V in the United States during the preconjugate vaccine era (13), 9V greatly decreased after PCV7 implementation (1) and was partially replaced by resistant 19A/ST156 (14). To verify that these isolates originated from a single recombinational serotype switch event involving 35B/ST558 and 9V/ST156 parental strains, we analyzed regions flanking the *cps35B* locus during 2015–2016, 35B/ST156 progeny and the original strain detected during 2012.

Methods

Isolates

The surveillance population of ABCs is \approx 32 million persons in 10 states (http://www.cdc.gov/pneumococcal/ surveillance.html). Serotype 35B IPD isolates described include 132 recovered during 2015 and 67 recovered during ABCs in 2016 (Table 1, https://wwwnc.cdc.gov/EID/ article/23/6/17-0071-T1.htm). The listing of 35B isolates from 2016 is incomplete because we typically receive all ABCs isolates recovered during a given year by the following summer. Relevant ST156 lineage isolates of other serotypes recovered during this and previous periods are shown in Table 2. Total numbers of ABCs 35B isolates recovered during 1999–2015 and categorized by patient age, penicillin MIC, and IPD incidence are shown in Table 3.

WGS and WGS-Based Predictions

Library construction and sequencing was performed as described (*12*). WGS accessions for all 199 serotype 35B isolates from 2015–2016, two previous 35B switch strains from previous years, and relevant strains of other sero-types of ST156 from previous years are provided (online Technical Appendix Table, https://wwwnc.cdc.gov/EID/article/23/6/17-0071-Techapp1.pdf). WGS pipeline data and quality metrics for all isolates are also provided (online Technical Appendix Table). Capsular serotypes, antimicrobial genotypes/phenotypes, MLST, sequence type (ST), and pili (presence or absence) for year 2015–2016 isolates were deduced through our bioinformatics pipeline (*6*,*12*,*15*).

Phylogeny

Paired-end fastq files were trimmed with Cutadapt version 1.8.1 (17), and draft genome assemblies were

 Table 2.
 Nonserotype 35B isolates of ST156 lineage included in study of penicillin-nonsusceptible 35B pneumococcal isolates causing

 IPD.
 United States.
 1998–2015*

] =	,													
Serotype/														
MLST type			Non-PBP resistance								Cli +			State (year
(no.)†	No.	PBP type‡	determinants§	Pen	Amo	Tax	Cft	Cfx	Mer	Ery	Tet	Cot	Fq	isolated)
9V/156 (25)	12	15:12:18	folAl100L, folPins178	4	2	1	2	>2	0.5	S	S	R	S	CA, GA, MD, MN, NY, TN (1998–1999)
	9	15:12:18	mef, folAl100, folPins178	4	2	1	2	>2	0.5	R	S	R	S	ČA, CT, MD, MN, OR, TN (1998,1999, 2015)
	2	15:12:18	<i>ermB, tetM</i> , folAl100L, folAins178	4	2	1	2	>2	0.5	R	R	R	S	CA (2015)
	1	15:12:18	<i>mef, tetM, folA</i> I100L, <i>folPins178</i>	4	2	1	2	>2	0.5	R	S	R	S	CT (2015)
	1	15:12:228	Mef, folAI100L, folPins178	4	2	1	2	>2	0.5	R	S	R	S	MD (2016)
19A/156 (4)	3	29:12:26	mef, folAI100L, folAins189	4	2	8	4	>2	0.5	R	S	R	S	CA,GA (2009)
	1	8:12:36	mef, folAI100L, folAins189	1	2	0.5	0.5	2	0.25	R	S	R	S	GA (2015)
13/156 (1)	1	15:12:173	folAl100L, folPins178	1	1	0.25	0.25	1	0.5	S	S	R	S	TN (2015)
31/156 (1)	1	15:12:18	mef, folAl100L, folPins178	4	2	1	2	>2	0.5	R	S	R	S	MN (2015)

*All isolates were positive for pilus PI-type 1 and negative for pilus PI-type 2. IPD, invasive pneumococcal disease; MLST, multilocus sequence type; PBP, penicillin-binding protein; R, resistant; S, susceptible; ST, sequence type.

†Types that probably arose through serotype switching are indicated in bold.

‡See Li et al. (15) and MIC correlates for PBP types (http://www.cdc.gov/streplab/mic-tables.html).

\$For a description of WGS-based bioinformatic pipeline for deduction of all features shown, see Metcalf et al. (6,12). For a description of folP insertions (folPins178, folP189), see Figure 1 in Metcalf et al. (12).

Predicted MICs for β-lactam antimicrobial drugs were based on transpeptidase domain sequences of PBPs 1a, 2b, and 2x

(http://www.cdc.gov/streplab/mic-tables.html). For penicillin (meningitis only), nonsusceptible is considered ≥0.12 µg/mL (*16*). Currently applied clinical cutoffs are also provided for the other 5 β-lactams shown (*16*). Where shown, R and S correspond to breakpoint MIC values (*16*). Amo, amoxicillin; Cft, ceftriaxone; Cfx, cefuroxime; Cli, clindamycin; Cot, cotrimoxazole; Ery, erythromycin; Fq, fluoroquinolones levofloxacin and ciprofloxacin; Mer, meropenem; Pen, penicillin; Tax, cefotaxime.

			No. 35B isc	plates from					penNS 35B
	Surveillance		patients t	by age, y	Relative incidence	Pe	en MIC, μg/	mL	_ isolates/ <i>penS</i> 35B
Year	population	% CIS	<5	≥5	of 35B IPD†	<u>></u> 2	0.12–1	<u><</u> 0.06	isolates
1998	17,383,935	86.1	3	18	1.40	9	5	7	2.0
1999	18,550,681	87.0	2	18	1.24	8	3	9	1.2
2000	19,821,607	86.3	4	21	1.46	7	9	9	1.8
2001	22,479,308	88.1	2	40	2.12	14	18	10	3.2
2002	25,051,246	87.6	2	34	1.64	20	8	8	3.5
2003	25,264,246	91.4	11	49	2.56	22	26	11	4.4
2004	27,419,898	87.9	15	69	3.49	44	25	15	4.6
2005	27,816,784	89.5	11	57	2.73	35	18	15	3.5
2006	28,204,455	86.7	1	65	2.70	37	15	14	3.7
2007	28,579,312	87.5	5	83	3.52	51	23	14	5.3
2008	28,856,774	86.7	12	80	3.68	62	16	14	5.6
2009	29,206,528	89.8	8	70	2.97	45	16	17	3.6
2010	29,757,552	90.2	4	67	2.65	52	14	5	13.2
2011	30,075,050	90.3	11	77	3.28	71	7	10	7.8
2012	30,356,544	90.6	13	101	4.14	94	14	6	18.0
2013	30,604,240	88.7	15	114	4.75	94	25	10	11.9
2014	31,328,211	88.2	16	116	4.78	104	22	6	21.0
2015	31,977,800	92.0	10	121	4.45	101	24	6	20.8
*CIS, cas	e isolates serotyped;	IPD, invasive	pneumococcal	disease.					
+Estimate	ed cases/million = tot	al 35Bs x 100/	% CIS surveilla	nce nonulati	n/1 000 000				

Table 3. Annual incidence and proportions of penicillin-nonsusceptible 35B pneumococcal isolates causing IPD, United States, 1999-2015*

constructed by using VelvetOptimiser version 2.2.5 with an optimal kmer value calculated by using VelvetK (18). Core genome single-nucleotide polymorphism (SNP) identification and alignment were performed by using kSNP3.0 (19). A maximum-likelihood phylogenetic tree was generated from the core SNP alignment by using RaxML version 7.3.0 (20). RaxML was run with an ASC GTRGAMMA DNA substitution model and used the Lewis method for ascertainment bias correction. Node support was assessed by using 500 bootstrap replicates.

Conventional MIC Testing and Serotyping

Serotype 35B isolates recovered during 2015 were subjected to conventional broth dilution testing for determination of antimicrobial MICs. A selection of these isolates were also subjected to conventional serotyping by using CDC typing antisera as described (6).

Statistical Analyses

A χ^2 test was performed to evaluate differences among groups. This test was performed by using OpenEpi Version 3.01 (http://www.openepi.com/Menu/OE Menu.htm).

Results

Increase in Penicillin-Nonsusceptible 35B during the Conjugate Vaccine Era

During 1998–2001, penicillin-nonsusceptible 35B accounted for 67.6% (108) of serotype 35B ABCs isolates (Table 3). During 2002-2015, the proportion of penicillinnonsusceptible IPD isolates with serotype 35B increased to 87.7% (1,237; p<0.001).

Population Snapshot of Ongoing ABCs for 35B IPD, 2015-2016

Among 2,710 IPD isolates obtained during 2015 and subjected to WGS, 132 (4.9%) were serotype 35B. Of 1,528 IPD isolates recovered from partial year 2016 IPD surveillance, 67 (4.4%) were serotype 35B. Most (168/199) of these isolates belonged to penicillin-nonsusceptible clonal complex (CC) 558 (168 isolates) and CC156 (21 isolates) (Figure 1; Table 1). Serotype 35B CC558 and CC156 isolates of all serotypes discussed were uniformly positive for the rrgA gene (Tables 1, 2), which encodes a pilus subunit that functions in epithelial adhesion (22). Ten isolates of long-standing penicillin-susceptible 35B/ST452 (3) were also recovered. Single 35B isolates were identified of ST1092, a lineage of conjugate vaccine serotypes 6A and 6B (http://pubmlst.org/spneumoniae/) and of ST11818 (highly related to 15A/ST63), an antimicrobial-resistant nonvaccine serotype lineage that has increased in the postconjugate vaccine era (4).

CC558 (35B/CC558)

Of 168 35B/CC558 isolates obtained, 147 were ST558, 20 were single-locus variants (SLVs) of ST558 corresponding to 13 STs, and 1 was a double-locus variant (Figure 1; Table 1). Within CC558, only ST558 had SLVs, which is consistent with initial successful establishment of 35B/ ST558 in its ecologic niche and subsequent rare shedding of closely related SLVs (21).

The increased incidence of 35B IPD during the post-PCV7 period (2.1-3.7 cases/million population during 2001-2009 vs. 1.2-1.3 cases/million population during 1998-1999) and the post-PCV13 period (3.3-4.8 cases/million



Figure 1. Population snapshot of 199 serotype 35B pneumococcal isolates obtained by ongoing Active Bacterial Core surveillance, United States, 2015–2016, configured by using eBURST (*21*). Diameters are proportional to number of isolates. Solid lines indicate single-locus variants, and the single dashed line indicates a double-locus variant of ST558. ST, sequence type.

population during 2011–2015), combined with the consistent trend of markedly increased proportions of penicillinnonsusceptible 35B IPD isolates throughout the conjugate vaccine era (Table 2), is consistent with reported increased 35B/ST558 in IPD and carriage (4–11). ABCs surveillance sites increased after 2000, but 35B IPD incidence calculations did not vary whether including the expanded surveillance sites or by using only continuously participating ABCs sites during 1998–2015.

CC156 (35B/CC156)

We analyzed PBP types (6,12,15) of 35B/ST558 (4:7:7) and 35B/ST156 (4:12:7) isolates. These PBP amino acid sequence types are used for predicting β -lactam MICs and correspond to PBP transpeptidase domains from PBP1a, PBP2b and PB-P2x, respectively. PBP genes *pbp1a* and *pbp2x* flank opposite ends of the capsular biosynthetic locus and are sometimes co-transferred during serotype switching events (6,23-25). The 35B/ST558 lineage has been nearly exclusively associated with PBP type 4:7:7 among isolates obtained since 1998, and the serotype 9V/ST156 lineage is similarly highly associated with PBP type 15:12:18 (6,15). However, 9V/ST156 is rare among IPD isolates in the post-PCV7 period.

In addition to the PBP2b-12 marker, *mef* gene, and FolA-I100L substitution, candidate *cps35B* recipient 9V/ST156 strains contain the 2-codon insertion designated *fol-Pins178* (Table 2; Figures 2, 3). Such 1–2 codon *folP* insertions, which together with FolA-I100L confer cotrimoxazole resistance, are categorized by specific location of the insertion and specific sequence flanking the insertions (*12*). These genomic features are also found within the 35B/ST156 lineage isolates described (Table 1; Figures 2, 3), which are consistent with a 9V/ST156 (*mef*, FolA-I100L,

folPins178) strain serving as the recipient strain for a 35B/ ST558 *cps35B* donor strain (Figure 3). Another potential recipient strain present before and after PCV13 introduction was 19A/ST156 (*6*,*14*). However, this lineage is associated with the *folPins189* insertion (Table 2; Figure 2).

Both flanking *pbp* loci from 35B/ST558 were cotransferred with the *cps35B* locus to replace the *cps9V*, *pbp1a*-15 and *pbp2x*-18 determinants in the putative 9V/ST156 recipient, which resulted in PBP type 4:12:7 (Figure 3). This serotype switch progeny strain was obtained from a 4-yearold child during 2012 and is a potential progenitor of the current invasive 35B/CC156 lineage (isolate 2012221165) (Figures 2, 3). Antimicrobial resistance markers PBP2b-12, *mef*, FoIA-I100L, and *folPins178*, combined with the close phylogenetic relatedness of the 9V/ST156 isolates (Figure 2), suggest that a member of this lineage served as the recipient parental strain for the 35B/ST156 clade isolated in ongoing ABCs during 2015–2016 (Figure 3).

Analysis of the regions flanking the *cps35B* locus for all 35B/ST156 lineage isolates obtained during 2015–2016 showed identical recombinational sites at bases 6,453 (left coordinate of progeny reference) (Figure 3) and 10,836 (right coordinate), which is clearly indicative of a single event within a 35B/ST156 ancestral strain of the 19 progeny shown (Figure 3). Thus, a double-crossover event replaced the recipient strain *cps9V* locus and flanking PBP markers (2x-18 and 1a-15) with the *cps35B* locus and its flanking PBP markers (2x-7 and 1a-4). On the basis of available strain data, the original progeny strain is predicted to have been an ST156 strain with the PBP type 4:12:7; a total of 14 of the 19 35B/ST156 lineage strains still shared these characteristics (Table 1). Five isolates are SLVs or differ in PBP2b type.

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Figure 2. Phylogenetic analysis of potential recipient and serotype switch pneumococcal progeny strains within the ST56 lineage based upon a total alignment of 10,409 core single-nucleotide Serotype 35B/ST156 lineage progeny polymorphisms, United States, 2015-2016. All 20 serotype 35B isolated during 2015-2016 progeny shown were recovered through Active Bacterial Core PBP-4:12:7, mef+, FolA-I100L, folPins178 surveillance, and all but 2 indicated strains were obtained during 2015-2016. Isolate features are depicted for the 2 major nodes, with exceptions indicated by asterisks within the tree. Bootstrap values are indicated at key nodes. The serotype 9V isolate that was used for the reference recipient sequences described in First 35B/ST156 from ABCs collected during 2012 (isolate 2012221165) Figure 3 is indicated as the third isolate from bottom. All isolates above the 9V recipient reference within this major cluster, except where indicated, are also 9V/ST156. Three additional serotype switch ST156 strain types detected by Active Bacterial Core ero 13/ST156, PBP-15:12:18, mefA+, FoIA-I100L, folPins178 surveillance during 2015-2016 are indicated by asterisks (single isolates of serotypes 31 and 13 and 4 serotype 19A isolates). Scale bar indicates nucleotide substitutions per site. Sero 19A/ST156, PBP-8:12:36, mef+, FoIA-1100L, folPins189, recovered during 2009 and 2015 Serotype 9V/ST156 recipient lineage collected during 1998-2016 * Single 35B/ST156 lineage (ST10174) ro 31/ST156.PBP-15:12:18 progeny isolate recovered in 2009 PBP-4:7:18, mefA, FoIA-I100L, folPi nce strain recovered during 2016 otype 35B/ST558 donor reference strain recovered in 2015

We detected the small (491 bp) segment (bases 11349– 11839 of progeny) (Figure 3) of clearly recipient lineage origin within the left side of the major recombinational fragment. During a single–double crossover event that facilitates a serotype switch, additional independent doublecrossover events appear to occur concurrently (27). However, these events probably do not occur simultaneously. It appears that the actual serotype switch event involved a shorter donor fragment bordering upon the right side of this small recipient lineage segment (base 11839), followed by a second double-crossover event that bordered upon the left side of the recipient lineage fragment (base 11349).

A single penicillin-susceptible 35B/ST162 (SLV of ST156) has the completely sensitive PBP type 0:0:0 (6,12) (Table 1). This strain arose through an independent serotype switch event that involved a penicillin-susceptible recipient strain.

Postserotype Switch Event Diversification of 35B/ST156 Progeny

Five of the 19 35B/ST156 lineage progeny showed indications of genetic diversification that occurred after the capsular switch event. Four of these isolates have 1 of 3 SLV MLSTs of ST156 (ST9910, ST11584, and ST12921) (Table 1). Although 18 of the 19 strains were PBP type 4:12:7, the SLV ST12921 variant had PBP type 4:11:7. For this particular strain, it is probable that recombination with a highly penicillin-resistant ST320 strain, prevalent during the post-PCV7 era and having PBP type 13:11:16 (6), simultaneously replaced the *pbp2b* locus and flanking *ddl* sequence to change the PBP type to 4:11:7 and the MLST type to ST12921 through transfer of *ddl* with the resistanceconferring selectable *pbp2b* allele (*28*). We also observed an increased MIC for amoxicillin for this PBP type 4:11:7 strain compared with MICs for PBP type 4:12:7 strains (Table 1).

Two isolates (20152877 and 20161763) underwent a postswitch intra-cps35B gene deletion event within the wciG gene (Figure 3), which is predicted to encode an acetyltransferase (26). Although these 2 isolates were serotyped as 35B by using CDC typing antisera, they differed in reactivity with serologic factor 35a compared with the other 17 isolates of this lineage (Table 4). Typing antisera factors 29b and 35c are the CDC Quellung reagents definitive for serotype 35B. The original protocol (29,30) that CDC first followed for serogroup 35 resolution also used factor 35a along with factors 29b and 35c for identification of serotype 35B. We found that the 2 wciG deletion strains did not react with factor 35a, but the other 17 serotype 35B strains reacted strongly with factor 35a. These preliminary data is suggestive of a new serotype within serogroup 35 because this specific factor reactivity pattern has not been observed for serogroup 35 (31).

Invasive Pneumococci and Serotype Switch Lineage



Figure 3. Diagrammatic representation of *cps* loci and adjacent regions from donor, recipient and progeny strains depicting serotype switch event for pneumococcal isolates, United States, 2015–2016. Red and green lines in progeny indicate regions of sequence identity or near identity (<2 single-nucleotide polymorphisms/10,000 bp) to the above corresponding donor and recipient sequences, respectively. Rectangles indicate relative locations of PBP gene types for *pbp2x* and *pbp1a*. Below each *cps* locus, a representative reference strain is indicated along with relevant features determined through a bioinformatics pipeline (MLST, PBP type, resistance markers). Junctions between donor and recipient sequences involved in the 2 single recombinational crossovers in the gene replacement event are indicated with blue arrowheads above the progeny diagram, although a single short internal region with sequence identity to the recipient nested within the donor fragment (left coordinates 11349–11839) is also present. Below each green or red segment of the progeny, the level of sequence identity to donor and recipient is provided. The list of each progeny strain, date of isolation, and state is provided. Where MLST is not ST156, its single locus variants (ST9910, ST11584, and ST12921) are included. Two exceptions indicating flanking post-switch recombination within left coordinates 1–6453 are indicated in isolates 20152884 and 20161413 (asterisks): isolate 20152884 had only 99.3%–99.5% identity to recipient and donor over bases 1–3715, and isolate 20161413 had only 99.5%–99.7% identity to recipient and donor over bases 1–3715, and isolate 20161413 had only 99.5%–99.7% identity to recipient and donor over bases 1–2143. Two strains on the right indicate a post-switch deletion event within the wciG putative acetyltransferase gene, which putatively contributes to the acetylation pattern of the serotype 35B polysaccharide (26). MLST, multilocus sequence type; PBP, penicillin-binding protein; ST, sequence type.

Further indication of chromosome-wide postswitch diversification of this single clade was shown in the left-flanking region of the *cps35B* locus. Two progeny strains showed diversification within the first 2–3.7 kb when compared with the other 17 progeny. The 6-kb region immediately to the left of base 1 in all of the progeny strains had \leq 99.2% sequence identity with the most similar potential ST156 parental recipient strains that we analyzed. However, beyond this segment, progeny had sequence identity with the parental strain for \geq 8 kb.

35B/ST156 Variant Lineages Arising through Separate Serotype Switch Events

The 9V/ST156 clade also appears likely to have served as the recipient for an independent serotype switch from the same 2 parental strains (Figure 3) which resulted in 35B/ST10174, an SLV of ST156 (Figure 2) obtained from an infant during 2009 (isolate 2009219987). This isolate differs in the flanking *pbp2x* marker and distal *pbp2b* marker (PBP type 4:7:18). Features of this variant have been described (6), and we have not obtained additional 35B isolates with these distinguishing features. A third sero-type switch event involving a CC156 recipient strain is intuitive from the pipeline data, which indicate that the 35B SLV of ST162 is featured by the β -lactam–susceptible PBP type 0:0:0 (Table 1). ST162 has long been associated with penicillin-susceptible 9V strains (*13*) and more recently with PI-1 positive and penicillin-susceptible 23B, 15B, and 15C strains (*6*).

Nonserotype 35B Variants of ST156

Single ST156 isolates of serotypes 31 and 13 were obtained during 2015 (Figure 2) and showed high relatedness to different 9V subdivisions. Again, the likely recipient background for the presumed capsular switch does not appear likely to involve the 19A/ST156 lineage (Figure 2), which

serotype switch lineage, United States, 2015–2016*												
		Que	llung fao	ctor								
Strain	35b	29b	35c	42a	35a							
cps35B†	-	+	+	-	+							
<i>cps35B</i> (<i>wciG</i> deletion)‡	-	+	+	-	-							
*CDC, Centers for Disease Co -, negative; +, positive.	ntrol and	Preventio	on; ST, se	equence	type;							
†Refers to 17 progeny resultin	g from re	combinati	ion indica	ited in lef	ft							
+Pofore to 2 indicated waiC do	column under progeny lineage diagram in Figure 3.											
rogeny lineage diagram in Figure 3												
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Table 4. Serologic comparison of 35B/ST156 lineage strainswith CDC Quellung reagents for resolution of serogroup 35Binvasive serotype pneumococci including an expandingserotype switch lineage, United States, 2015–2016*

was well-represented during the 2000s after PCV7 implementation (6, 14, 32).

CCs of Remaining 35B Isolates Obtained during 2015–2016

Ten of the 12 isolates other than penicillin-nonsusceptible CC558 and CC156 (together composed of 187 isolates) were of the long-established penicillin-susceptible 35B/ ST452 lineage (3), which decreased in proportion during the 2000s (4) (Figure 1; Table 3). The 2 remaining 35B isolates obtained during 2015-2016 also appear to have originated through serotype switching events involving a 35B/ST558 cps35B donor, as implicated by the presence of the PBP1a-4 or PBP2x-7 determinants flanking the cps loci of these 2 progeny strains (35B/ST11818 and 35B/ ST1092) (Table 1). The 35B/ST11818 variant is an SLV of ST63 and has the same resistance features and accessory resistance genes (ermB and tetM) as the currently common 15A/ST63 clone (4,6,32). The 35B/ST1092 isolate is likely to have originated through serotype switching with a serogroup 6 recipient strain (6). The 14 remaining ST1092 isolates obtained during surveillance in 2015-2016 were serotype 6C. The 6 previously collected ST1092 IPD isolates (1999-2013) represented in our WGS collection are from serotype 6A, 6B, and 6C strains (6; B. Beall, unpub. data).

Discussion

An increase of penicillin-nonsusceptible serotype 35B IPD and carriage caused by 35B/ST558 has been apparent in the United States since the introduction of PCV7 in 2000, and it has shown a major increase after PCV13 implementation (4–8). This finding is of concern because even strains that are rarely detected in IPD sometimes rapidly emerge. For example, the 19A/ST320 strain was not detected during extensive characterization of pre-PCV7 ABCs isolates (13), yet it became the predominant invasive pneumococcal strain during 2005–2009 (14,32). We have performed comprehensive strain characterization (MLST and WGS) of pediatric (from children \leq 5 years of age) ABCs isolates obtained during 1999, 2001, 2002, 2008, 2009, and 2011– 2013 (6,13) and WGS-based characterization of a large sampling of isolates from all age groups during 1998–2013 (4,6,13; B. Beall, unpub. data).

Before 2015, we detected only 1 isolate of the 35B/ ST156 lineage that was recovered during 2012 (6). Thus, we feel justified in describing it as newly emergent isolate. A smaller study was recently published (during peer review of this article) that described 78 invasive and 48 noninvasive serotype 35B isolates obtained during 1994– 2014 from 8 hospitals in 8 states (33). Our data, which included a population-based sampling of 199 35B isolates obtained during 2015–2016, clearly shows the current national predominance of 35B/ST558 and does not support the observation that 35B/ST156 is the major contributor to post-PCV13 antimicrobial-resistant 35B. Both studies noted the initial appearance and emergence of 35B/ST156 in the post-PCV13 period.

This recent identification of the antimicrobial-resistant 35B/ST156 lineage and its subsequent detection within 6 ABCs sites is a cause for concern. The ST156 lineage has shown a remarkable propensity to persist through undergoing serotype-switch events (12,23,25,32). The penicillin-resistant 9V/ST156 lineage was the predominant serotype 9V cause of IPD in the United States during the pre-PCV7 era (13,32). Soon after introduction of PCV7, serotype 9V IPD became rare (1,2), and 19A became the predominant representative of the ST156 lineage within ABCs (14,32). After introduction of PCV13, 35B has become the predominant serotype of the ST156 lineage within the United States (B. Beall, unpub. data).

A distinct antimicrobial-susceptible serotype 35B SLV of ST156 (35B/ST162) is included among 35B ABCs of 35B during 2015–2016 by the β -lactam–susceptible PBP type 0:0:0. Thus, our data indicates that ≥ 3 independent serotype switches involving the nonvaccine type cps35B locus and the broad ST156 clonal complex serving as recipient strains have previously occurred. In this study, we demonstrated that all penicillin-nonsusceptible 35B/ST156 lineage isolates obtained during current ABCs (2015–2016) arose through a single ancestral recombination event. This event was facilitated through detailed analyses of crossover points and comparisons of corresponding regions of all progeny isolates with likely parental 35B/ST558 and 9V/ ST156 strains. The genetic plasticity of the ST156 lineage is also highlighted in this study by detection of postserotype switch changes affecting β-lactam resistance (PBP1a type) and capsular serotype (wciZ deletion), which is potentially reflective of recent antimicrobial drug pressure and immunologic selection pressure.

An additional 35B variant within a vaccine serotype lineage is shown with ST1092 that is typically associated with serogroup 6 strains (Figure 1). Because these putative 35B switch variants were not detected during extensive strain surveillance before and shortly after conjugate vaccine implementation (3,4,13), it is plausible that these serotype switches occurred after implementation of conjugate vaccine. The observation of a 35B variant within the antimicrobial 15A/ST63 lineage brings the number of serotype switch events generating 35B strains described in this study to 5; (35B/ST11818, 35B/ST156, 35B/ST162 from 2015–2016, and 35B/ST10174 from 2009). Except for the 35B/ST162 variant, these serotype switch events were predicted on the basis of progeny PBP type to involve the 35B/ST58 strain as the *cps35B* donor (online Technical Appendix Table).

Although conjugate vaccines have a history of providing effective and durable protection against IPD (1,2), the continued emergence and expansion of serotype 35B into different clonal complexes supports continued development of wider spectrum pneumococcal vaccines. Serotype 19A IPD, although relatively uncommon in the pre-PCV7 era, rapidly became the predominant invasive serotype in the post-PCV7 period (14,32,34). Serotype 35B strains have several of the same features that were found among serotype 19A strains before implementation of PCV7 in 2000. These features that could predispose for serotype 35B to continue its increasing trend as a cause of IPD include its lack of inclusion within conjugate vaccine, high carriage rates within children, antimicrobial resistance, clonal expansion, and serotype switching. An experimental 15-valent conjugate vaccine in development includes serotypes 22F and 33F (35), which have increased as causes of IPD in the postconjugate vaccine era. Serotypes 15A, 15B, and 23A are expressed by moderately antimicrobial-resistant clones and are not uncommon causes of IPD (4,32). Although less resistant to β -lactam antimicrobial drugs than 35B/ST558 and 35B/ST156, these strains also present a challenge to address through more encompassing pneumococcal vaccines.

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References

- Pilishvili T, Lexau C, Farley MM, Hadler J, Harrison LH, Bennett NM, et al.; Active Bacterial Core Surveillance/Emerging Infections Program Network. Sustained reductions in invasive pneumococcal disease in the era of conjugate vaccine. J Infect Dis. 2010;201:32–41. http://dx.doi.org/10.1086/648593
- Moore MR, Link-Gelles R, Schaffner W, Lynfield R, Lexau C, Bennett NM, et al. Effect of use of 13-valent pneumococcal conjugate vaccine in children on invasive pneumococcal disease in children and adults in the USA: analysis of multisite, populationbased surveillance. Lancet Infect Dis. 2015;15:301–9. http://dx.doi.org/10.1016/S1473-3099(14)71081-3
- Beall B, McEllistrem MC, Gertz RE Jr, Boxrud DJ, Besser JM, Harrison LH, et al.; Active Bacterial Core Surveillance/Emerging Infections Program Network. Emergence of a novel penicillinnonsusceptible, invasive serotype 35B clone of *Streptococcus pneumoniae* within the United States. J Infect Dis. 2002;186:118– 22. http://dx.doi.org/10.1086/341072
- Gertz RE Jr, Li Z, Pimenta FC, Jackson D, Juni BA, Lynfield R, et al. Increased penicillin-nonsusceptibility of nonvaccine serotype (other than 19A and 6A) invasive pneumococci in post 7 valent conjugate vaccine era. J Infect Dis. 2010;201:770–5. http://dx.doi.org/10.1086/650496
- Sharma D, Baughman W, Holst A, Thomas S, Jackson D, da Gloria Carvalho M, et al. Pneumococcal carriage and invasive disease in children before introduction of the 13-valent conjugate vaccine: comparison with the era before 7-valent conjugate vaccine. Pediatr Infect Dis J. 2013;32:e45–53. http://dx.doi.org/10.1097/INF.0b013e3182788fdd
- Metcalf BJ, Gertz RE Jr, Gladstone RA, Walker H, Sherwood LK, Jackson D, et al.; Active Bacterial Core surveillance team. Strain features and distributions in pneumococci from children with invasive disease before and after 13-valent conjugate vaccine implementation in the USA. Clin Microbiol Infect. 2016;22:60. e9–29. http://dx.doi.org/10.1016/j.cmi.2015.08.027
- Desai AP, Sharma D, Crispell EK, Baughman W, Thomas S, Tunali A, et al. Decline in pneumococcal nasopharyngeal carriage of vaccine serotypes after the introduction of the 13-valent pneumococcal conjugate vaccine in children in Atlanta, Georgia. Pediatr Infect Dis J. 2015;34:1168–74. http://dx.doi.org/10.1097/ INF.000000000000849

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- Kaur R, Casey JR, Pichichero ME. Emerging *Streptococcus* pneumoniae strains colonizing the nasopharynx in children after 13-valent pneumococcal conjugate vaccination in comparison to the 7-valent era, 2006–2015. Pediatr Infect Dis J. 2016;35:901–6. http://dx.doi.org/10.1097/INF.000000000001206
- Kawaguchiya M, Urushibara N, Kobayashi N. Multidrug resistance in non-PCV13 serotypes of *Streptococcus pneumoniae* in northern Japan, 2014. Microb Drug Resist. 2017;23:206–14. http://dx.doi.org/ 10.1089/mdr.2016.0054
- McElligott M, Vickers I, Meehan M, Cafferkey M, Cunney R, Humphreys H. Noninvasive pneumococcal clones associated with antimicrobial nonsusceptibility isolated from children in the era of conjugate vaccines. Antimicrob Agents Chemother. 2015;59:5761– 7. http://dx.doi.org/10.1128/AAC.00990-15
- Golden AR, Adam HJ, Gilmour MW, Baxter MR, Martin I, Nichol KA, et al. Assessment of multidrug resistance, clonality and virulence in non–PCV-13 *Streptococcus pneumoniae* serotypes in Canada, 2011–13. J Antimicrob Chemother. 2015;70:1960–4.
- Metcalf BJ, Chochua S, Gertz RE Jr, Li Z, Walker H, Tran T, et al.; Active Bacterial Core surveillance team. Using whole genome sequencing to identify resistance determinants and predict antimicrobial resistance phenotypes for year 2015 invasive pneumococcal disease isolates recovered in the United States. Clin Microbiol Infect. 2016;22:1002.e1–8. http://dx.doi.org/ 10.1016/j.cmi.2016.08.001
- Beall B, McEllistrem MC, Gertz RE Jr, Wedel S, Boxrud DJ, Gonzalez AL, et al.; Active Bacterial Core Surveillance Team. Pre- and postvaccination clonal compositions of invasive pneumococcal serotypes for isolates collected in the United States in 1999, 2001, and 2002. J Clin Microbiol. 2006;44:999–1017. http://dx.doi.org/10.1128/JCM.44.3.999-1017.2006
- Beall BW, Gertz RE, Hulkower RL, Whitney CG, Moore MR, Brueggemann AB. Shifting genetic structure of invasive serotype 19A pneumococci in the United States. J Infect Dis. 2011;203: 1360–8. http://dx.doi.org/10.1093/infdis/jir052
- Li Y, Metcalf BJ, Chochua S, Li Z, Gertz RE Jr, Walker H, et al. Penicillin-binding protein transpeptidase signatures for tracking and predicting β-lactam resistance levels in *Streptococcus pneumoniae*. MBio. 2016;7:e00756–16. http://dx.doi.org/10.1128/ mBio.00756-16
- Clinical and Laboratory Standards Institute (CLSI). Performance standards for antimicrobial susceptibility testing. Twenty-third informational supplement (M100–S22.CLSI). Wayne (PA): The Institute; 2013.
- Martin M. Cutadapt removes adapter sequences from highthroughput sequencing reads. EMBnet. 2011;17:10–2. http://dx.doi.org/10.14806/ej.17.1.200
- Zerbino DR, Birney E. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. Genome Res. 2008;18:821–9. http://dx.doi.org/10.1101/gr.074492.107
- Gardner SN, Slezak T, Hall BG. kSNP3.0: SNP detection and phylogenetic analysis of genomes without genome alignment or reference genome. Bioinformatics. 2015;31:2877–8. http://dx.doi.org/10.1093/bioinformatics/btv271
- Stamatakis A. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. Bioinformatics. 2014;30:1312–3. http://dx.doi.org/10.1093/bioinformatics/btu033
- Feil EJ, Li BC, Aanensen DM, Hanage WP, Spratt BG. eBURST: inferring patterns of evolutionary descent among clusters of related bacterial genotypes from multilocus sequence typing data. J Bacteriol. 2004;186:1518–30. http://dx.doi.org/10.1128/ JB.186.5.1518-1530.2004

- Barocchi MA, Ries J, Zogaj X, Hemsley C, Albiger B, Kanth A, et al. A pneumococcal pilus influences virulence and host inflammatory responses. Proc Natl Acad Sci U S A. 2006;103:2857–62. http://dx.doi.org/10.1073/pnas.0511017103
- Coffey TJ, Daniels M, Enright MC, Spratt BG. Serotype 14 variants of the Spanish penicillin-resistant serotype 9V clone of *Streptococcus pneumoniae* arose by large recombinational replacements of the *cpsA-pbp1a* region. Microbiology. 1999; 145:2023–31. http://dx.doi.org/10.1099/13500872-145-8-2023
- Brueggemann AB, Pai R, Crook DW, Beall B. Vaccine escape recombinants emerge after pneumococcal vaccination in the United States. PLoS Pathog. 2007;3:e168. http://dx.doi.org/ 10.1371/journal.ppat.0030168
- Wyres KL, Lambertsen LM, Croucher NJ, McGee L, von Gottberg A, Liñares J, et al. Pneumococcal capsular switching: a historical perspective. J Infect Dis. 2013;207:439–49. http://dx.doi.org/10.1093/infdis/jis703
- Mavroidi A, Aanensen DM, Godoy D, Skovsted IC, Kaltoft MS, Reeves PR, et al. Genetic relatedness of the *Streptococcus pneumoniae* capsular biosynthetic loci. J Bacteriol. 2007; 189:7841–55. http://dx.doi.org/10.1128/JB.00836-07
- Golubchik T, Brueggemann AB, Street T, Gertz RE Jr, Spencer CC, Ho T, et al. Pneumococcal genome sequencing tracks a vaccine escape variant formed through a multi-fragment recombination event. Nat Genet. 2012;44:352–5. http://dx.doi.org/10.1038/ng.1072
- Enright MC, Spratt BG. Extensive variation in the *ddl* gene of penicillin-resistant *Streptococcus pneumoniae* results from a hitchhiking effect driven by the penicillin-binding protein 2b gene. Mol Biol Evol. 1999;16:1687–95. http://dx.doi.org/ 10.1093/oxfordjournals.molbev.a026082
- Kauffmann F, Mørch E, Schmith K. On the serology of the pneumococcus-group. J Immunol. 1940;39:397–426.
- Lund E. On the nomenclature of the pneumococcal types. Int J Syst Bacteriol. 1970;20:321–3. http://dx.doi.org/10.1099/ 00207713-20-3-321
- 31. Henrichsen J. Six newly recognized types of *Streptococcus pneumoniae*. J Clin Microbiol. 1995;33:2759–62.
- 32. Kim L, McGee L, Tomczyk S, Beall B. Biological and epidemiological features of antibiotic-resistant *Streptococcus pneumoniae* in pre- and post-conjugate vaccine eras: a United States perspective. Clin Microbiol Rev. 2016;29:525–52. http://dx.doi.org/10.1128/CMR.00058-15
- Olarte L, Kaplan SL, Barson WJ, Romero JR, Lin PL, Tan TQ, et al. Emergence of multidrug-resistant pneumococcal serotype 35B among children in the United States. J Clin Microbiol. 2017;55:724–34. http://dx.doi.org/10.1128/JCM.01778-16
- Moore MR, Gertz RE Jr, Woodbury RL, Barkocy-Gallagher GA, Schaffner W, Lexau C, et al. Population snapshot of emergent *Streptococcus pneumoniae* serotype 19A in the United States, 2005. J Infect Dis. 2008;197:1016–27. http://dx.doi.org/10.1086/ 528996
- 35. Sobanjo-ter Meulen A, Vesikari T, Malacaman EA, Shapiro SA, Dallas MJ, Hoover PA, et al. Safety, tolerability and immunogenicity of 15-valent pneumococcal conjugate vaccine in toddlers previously vaccinated with 7-valent pneumococcal conjugate vaccine. Pediatr Infect Dis J. 2015;34:186–94. http://dx.doi.org/10.1097/INF.000000000000516

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Invasive Serotype 35B Pneumococci Including an Expanding Serotype Switch Lineage, United States, 2015–2016

Technical Appendix

Technical Appendix Table. Accession numbers, whole-genome sequencing pipeline features, and quality metrics of 207 invasive pneumococcal isolates, United States, 2015–2016*

												Length of			
									Non-PBP			longest		No.	No. bases
SRA accession			Year of					PBP type	resistance	No.	N50,	contig,	No. bases	contigs	in contigs
no.	Isolate name	State [†]	isolation	Serotype	pl-1‡	pl-2‡	MLST	1a:2b:2x§	determinants¶	contigs	bases	bases	in contigs	>1K	>1K
ERR586423#	2010200750	CA	2009	19A	Yes	No	156	29:12:26	mef, foIAI100L, foIPins189	84	105,726	341,189	2,087,641	39	2,073,476
SAMN05220851	20151623	GA	2015	35B	Yes	No	558	4:7:7	mef	345	11,534	52,098	2,066,482	299	2,028,530
SAMN05220873	20151893	GA	2015	35B	Yes	No	558	4:7:7	mef	69	104,614	165,383	2,022,450	31	2,010,218
SAMN05220875	20151895	GA	2015	35B	Yes	No	558	4:7:7	mef	50	78,901	206,084	2,098,334	40	2,091,749
SAMN05220883	20152130	GA	2015	35B	Yes	No	558	4:7:7	Negative	31	120,620	268,659	2,021,152	25	2,017,441
SAMN05220884	20152131	GA	2015	35B	Yes	No	156	4:12:7	mef, foIAI100L, foIPins180	63	61,342	238,796	2,101,100	57	2,097,051
SAMN05220924	20152247	GA	2015	35B	Yes	No	558	4:7:7	mef	32	165,555	329,660	2,023,444	25	2,019,140
SAMN05220932	20152255	GA	2015	35B	Yes	No	558	4:7:7	mef	45	91,096	208,104	2,060,658	38	2,056,189
SAMN05220978	20152635	GA	2015	35B	Yes	No	558	4:7:7	mef	45	105,615	158,315	2,022,574	36	2,017,138
SAMN05220988	20152649	GA	2015	35B	Yes	No	558	4:7:7	mef	63	74,766	267,777	2,055,552	49	2,047,804
SAMN05220990	20152651	GA	2015	35B	Yes	No	558	4:7:7	mef	36	158,343	260,315	2,048,689	26	2,042,220
SAMN05220991	20152652	GA	2015	35B	Yes	No	11250	4:14:7	folPins169	70	66,332	152,334	2,066,222	60	2,060,214
SAMN05221020	20152694	NM	2015	35B	Yes	No	558	4:7:7	folPins195	2255	3,153	35,263	3,471,381	770	2,623,239
SAMN05221036	20152712	NM	2015	35B	Yes	No	558	4:7:7	Negative	51	165,485	264,926	2,021,941	28	2,010,850
SAMN05221054	20152805	GA	2015	35B	Yes	No	558	4:7:7	mef	153	28,000	77,453	2,024,387	125	2,008,430
SAMN05221068	20152819	GA	2015	19A	Yes	No	156	8:12:36	mef, folAI100L, folPins189	54	79,725	171,901	2,107,451	47	2,103,268
SAMN05221081	20152877	ΤN	2015	35B	Yes	No	156	4:12:7	mef, folAI100L, folPins178	67	69,960	240,680	2,142,873	54	2,134,812
SAMN05221087	20152884	TN	2015	35B	Yes	No	9910	4:12:7	mef, folAl100L, folPins180	60	78,563	427,552	2,141,711	45	2,133,089
SAMN05221099	20152896	ΤN	2015	35B	No	No	558	4:7:7	mef	40	105,973	253,714	2,049,350	29	2,042,292
SAMN05221130	20152953	СТ	2015	35B	Yes	No	558	4:7:7	mef	41	104,752	359,235	2,048,113	33	2,043,373
SAMN05221131	20152954	СТ	2015	35B	Yes	No	558	4:7:7	mef, ParC-S79F	43	99,836	267,811	2,048,448	34	2,042,394
SAMN05221137	20152960	СТ	2015	35B	Yes	No	558	4:7:7	mef	50	93,353	225,698	2,052,543	39	2,045,342
SAMN05221153	20153004	NY	2015	35B	Yes	No	558	4:7:7	mef	43	106,157	256,689	2,015,996	35	2,011,436
SAMN05221170	20153021	GA	2015	35B	Yes	No	558	4:7:133	mef	61	64,424	171,124	2,056,356	55	2,052,821
SAMN05221230	20153206	CO	2015	35B	Yes	No	558	4:7:7	Negative	61	69,265	185,639	2,020,833	53	2,015,716
SAMN05221253	20153229	CO	2015	35B	Yes	No	11254	4:7:7	foIAI100L, foIPins169	54	69,587	141,164	2,067,296	48	2,063,425
SAMN05221268	20153245	NY	2015	35B	Yes	No	558	4:7:7	mef	48	100,363	186,047	2,043,861	37	2,038,001
SAMN05221290	20153268	TN	2015	35B	Yes	No	558	4:7:112	mef	36	110,217	306,204	2,022,381	30	2,019,060
SAMN05221329	20153308	TN	2015	35B	Yes	No	558	4:7:7	mef	56	75,409	139,907	2,025,258	49	2,020,002

												Length of			.
									Non-PBP			longest		No.	No. bases
SRA accession		<u>.</u>	Year of	o ,			NH OT	PBP type	resistance	No.	N50,	contig,	No. bases	contigs	in contigs
no.	Isolate name	State	isolation	Serotype	pi-1‡	pi-2‡	MLST	1a:2b:2x§	determinants	contigs	bases	bases	in contigs	>1K	>1K
SAMN05221338	20153394	CA	2015	35B	No	No	452	0:0:0	Negative	//	71,399	123,897	2,089,273	50	2,074,421
SAMN05221366	20153422	CA	2015	9V	Yes	No	156	15:12:18	ermB, foIAI100L, foIPins178, tetM	58	74,719	243,646	2,067,218	47	2,061,016
SAMN05221369	20153425	CA	2015	35B	No	No	452	0:0:0	Negative	71	101,781	195,615	2,161,796	37	2,143,676
SAMN05221410	20153478	NM	2015	35B	Yes	No	10493	4:7:7	mef	53	78,074	174,938	2,040,592	51	2,039,384
SAMN05221412	20153526	GA	2015	35B	No	No	558	4:7:7	mef	48	93,085	210,845	2,051,947	38	2,046,086
SAMN05221447	20153901	MN	2015	35B	Yes	No	558	4:7:7	mef	75	51,030	133,733	2,018,673	70	2,015,621
SAMN05221448	20153902	MN	2015	35B	Yes	No	558	4:7:7	mef	52	92,130	159,730	2,014,917	42	2,007,904
SAMN05221483	20153937	MN	2015	35B	Yes	No	558	4:7:7	mef	45	110,339	267,781	2,022,658	32	2,015,024
SAMN05221491	20153945	MN	2015	35B	Yes	No	558	4:7:7	mef	33	165,574	276,997	2,021,333	26	2,017,038
SAMN05221520	20153974	MN	2015	35B	Yes	No	558	4:7:7	Negative	69	74,571	156,076	2,022,222	58	2,014,941
SAMN05221873	20154264	NM	2015	35B	Yes	No	10493	4:7:7	mef, folAI100L	38	106,125	297,817	2,051,274	30	2,046,287
SAMN05221920	20154350	NY	2015	35B	Yes	No	558	4:7:7	mef	83	48,103	154,090	2,027,137	74	2,020,997
SAMN05221961	20154457	ΤN	2015	35B	Yes	No	558	4:7:7	folPins173	70	75,849	112,748	2,022,694	56	2,016,001
SAMN05221992	20154489	СТ	2015	35B	Yes	No	7487	4:49:7	Negative	49	92,283	165,554	2,026,826	40	2,021,564
SAMN05222034	20154575	GA	2015	35B	Yes	No	558	4:7:7	mef	29	123,759	268,656	2,016,212	24	2,012,931
SAMN05222063	20154698	MN	2015	35B	Yes	No	558	4:7:7	mef	65	71,564	158,172	2,012,692	54	2,005,300
SAMN05222074	20154709	MN	2015	35B	Yes	No	558	4:7:7	mef	30	110.276	313,438	2.006.123	23	2.002.203
SAMN05222102	20154740	MN	2015	35B	Yes	No	558	4:7:7	mef	49	82.837	206,129	2.007.751	40	2.002.868
SAMN05222113	20154753	MN	2015	31	Yes	No	156	15:12:18	mef, folAI100L, folPins178	56	78,904	241,592	2,091,629	43	2,083,767
SAMN05222122	20154762	MN	2015	35B	Yes	No	558	4:7:7	Negative	115	34,829	145,244	2,060,779	98	2,049,243
SAMN05222124	20154764	MN	2015	35B	Yes	No	558	4:7:7	mef	40	102.412	344.853	2.001.315	35	1.998.325
SAMN05222138	20154861	CO	2015	35B	Yes	No	558	4:7:7	mef	50	91,885	174,021	2,017,466	42	2,012,697
SAMN05222147	20154871	CO	2015	35B	Yes	No	558	4:7:7	Negative	48	86.836	146.223	2.013.425	43	2.010.386
SAMN05222149	20154873	CO	2015	35B	No	No	452	0:0:148	Negative	71	69,780	195.635	2.085.785	50	2.072.719
SAMN05222152	20154876	CO	2015	35B	Yes	No	558	4:7:28	mef	47	88.654	221.878	2.017.098	41	2.013.130
SAMN05222170	20154894	CO	2015	35B	Yes	No	558	4:7:7	Negative	56	80,744	260.302	2.045.935	43	2.037.405
SAMN05222174	20154898	CO	2015	35B	Yes	No	558	4:7:7	mef	43	120.528	223.043	2.014.603	33	2.008.712
SAMN05222180	20154904	CO	2015	35B	Yes	No	558	4:7:7	Negative	95	71.997	136,148	2.009.843	57	1.995.835
SAMN05222199	20154923	CO	2015	35B	Yes	No	11606	4:7:7	Negative	60	67.755	198.530	2.015.960	55	2.012.635
SAMN05222217	20154941	NY	2015	35B	Yes	No	558	4:7:7	Negative	64	61.235	159,717	2.014.227	58	2.010.063
SAMN05222220	20154980	TN	2015	35B	Yes	No	11584	4:12:7	mef, folAl100L, folPins178	62	68,761	174,251	2,138,313	52	2,131,321
SAMN05222224	20154985	ΤN	2015	35B	Yes	No	7486	4:7:7	mef	61	77.881	175.856	2.026.726	53	2.022.345
SAMN05222284	20155252	NM	2015	35B	Yes	No	558	4:7:7	mef	55	86,578	170,908	2.038.123	43	2,030,908
SAMN05222300	20155285	CT	2015	35B	Yes	No	558	4:7:7	mef	96	48,836	105,501	2.036.455	61	2,025,412
SAMN05222316	20155301	CT	2015	35B	Yes	No	558	4.7.7	mef	36	123 831	275 657	2 008 754	27	2 002 993
SAMN05222339	20155324	CT	2015	35B	Yes	No	558	4.7.7	mef	126	29 682	73,368	2 027 346	113	2 018 266
SAMN05222355	20155341	NY	2015	35B	Yes	No	558	4.7.7	mef	39	105 629	166 148	2,008,199	32	2 003 865
SAMN05222369	20155422	MD	2015	35B	Yes	No	558	4.7.7	mef	58	62 697	106 398	2,000,100	55	2,000,000
SAMN05222370	20155424	MD	2015	35B	Yes	No	558	4.7.7	mef	98	42 200	117 380	2,010,041	87	2,010,070
SAMN05222399	20155454	MD	2015	35B	Yes	No	11254	4:7:7	folAl100L, folPins169	78	48,396	116,466	2,035,836	71	2,031,364
SAMN05222426	20155483	MD	2015	35B	Yes	No	558	4:7:7	mef. ParC-D83Y	61	90.884	267.659	2.015.042	31	2.003.664
SAMN05222453	20155510	MD	2015	35B	Yes	No	558	4.7.7	folPins189	42	113,849	211,223	2.014.392	28	2,007,669
SAMN05222454	20155511	MD	2015	35B	Yes	No	558	4:7:7	mef	59	97,746	220.585	2.019.599	40	2.010.752
SAMN05222473	20155530	MD	2015	35B	Yes	No	558	4:7:7	Negative	99	40,708	108 537	2,058,817	90	2,052,352
SAMN05222480	20155537	MD	2015	35B	Yes	No	558	4:7:7	mef	41	93,125	177,099	2,013,666	33	2,009,209

												Length of			
									Non-PBP			longest		No.	No. bases
SRA accession	lestere enco	0	Year of	0			NU OT	PBP type	resistance	No.	N50,	contig,	No. bases	contigs	in contigs
no.	Isolate name	StateT	Isolation	Serotype	pi-1‡	pi-2‡	MLSI	1a:2b:2x§	determinants	contigs	bases	bases	In contigs	>1K	>1K
SAMN05222504	20155562	IN	2015	13	Yes	No	156	15:12:173	folPins178	43	111,285	255,659	2,045,236	31	2,039,152
SAMN05222517	20155901	GA	2015	35B	Yes	No	558	4:7:7	mef	41	112,540	165,695	2,013,797	30	2,007,676
SAMN05596826	20155922	MN	2015	35B	Yes	No	558	4:7:7	mef	32	106,109	325,049	1,999,846	26	1,996,090
SAMN05596827	20155926	MN	2015	35B	Yes	No	156	4:12:7	mef, folAI100L, folPins178	49	78,775	279,019	2,126,279	39	2,120,858
SAMN05596828	20155951	MN	2015	35B	Yes	No	558	4:7:7	mef	59	73,152	196,224	2,010,083	49	2,003,658
SAMN05222556	20156078	ΤN	2015	35B	Yes	No	558	4:7:7	Negative	59	82,107	221,409	2,025,411	49	2,020,007
SAMN05222599	20156298	GA	2015	35B	Yes	No	558	4:7:7	mef	47	93,019	223,438	2,017,585	33	2,010,494
SAMN05222602	20156301	GA	2015	35B	Yes	No	558	4:7:7	mef	45	105,889	262,369	2,011,956	33	2,006,448
SAMN05222619	20156540	CA	2015	35B	Yes	No	558	4:7:7	Negative	42	93,474	260,132	2,013,813	34	2,008,687
SAMN05222660	20156581	CA	2015	35B	No	No	558	4:7:7	mef	43	85762	301,048	2,002,644	40	2,000,453
SAMN05222676	20156649	СТ	2015	35B	Yes	No	558	4:7:7	mef	41	104,825	223,465	2,042,864	32	2,038,004
SAMN05222686	20156659	СТ	2015	35B	Yes	No	558	4:7:7	Negative	64	59.402	158.272	2.050.784	57	2.046.464
SAMN05222693	20156666	ĊТ	2015	35B	Yes	No	11603	4:7:7	mef	50	91.521	256.674	2.017.334	39	2.011.233
SAMN05222697	20156670	CT	2015	35B	Yes	No	558	4:7:7	mef	51	90,896	165,366	2.014.227	34	2,007,629
SAMN05222704	20156677	MD	2015	35B	Yes	No	558	4:7:7	Negative	65	66,752	162,146	2.019.702	58	2.015.338
SAMN05222705	20156678	MD	2015	35B	Yes	No	558	4.7.7	mef	48	105 515	221 519	2 018 478	42	2 014 793
SAMN05222711	20156684	MD	2015	35B	No	No	452	0.0.0	mef	68	69 921	161 942	2 087 079	47	2 075 378
SAMN05222715	20156689	MD	2015	35B	Yes	No	558	4.7.7	mef	37	91 052	279 457	2,007,073	30	2,012,047
SAMN05506820	20156033	MNI	2010	35B	Vec	No	558	4.7.7	mof	17	77 165	157 740	1 007 705	30	1 002 311
SAMN05596830	20156036	MNI	2015	35B	Vec	No	558	4.7.7	Negative	56	70 321	204 607	2 051 711	33 45	2 045 286
SAMN05506821	20156930	MNI	2015	35B	Voc	No	156	4.7.7	mof fold 1100	50 60	65 782	204,037	2,001,711	4J 52	2,043,200
SAMIN03390831	20130940		2015	356	165	NU	150	4.12.7	folPins178		05,762	210,473	2,129,301	52	2,124,334
SAMN05222800	20160119	GA	2015	35B	Yes	No	156	4:12:7	mef, folAI100L, folPins178	74	58,812	117,153	2,134,217	66	2,129,337
SAMN05222828	20160273	TN	2015	35B	Yes	No	558	4:7:7	mef,parC-D83Y, gyrA-S81Y	138	27,871	91,211	2,035,655	128	2,028,330
SAMN05222863	20160308	GA	2015	35B	Yes	No	558	4:7:7	Negative	46	101,661	165,518	2,045,468	38	2,040,619
SAMN05222880	20160528	MD	2015	35B	Yes	No	558	4:7:7	mef	85	40,578	102,802	2,055,955	78	2,051,459
SAMN05222885	20160534	MD	2015	35B	Yes	No	11604	4:7:7	mef	50	89,698	171,807	2,014,102	37	2,008,156
SAMN05222910	20160559	ΤN	2015	35B	Yes	No	558	4:7:7	mef	63	80,228	167,817	2,004,803	47	1,995,742
SAMN05222922	20160571	СТ	2015	35B	Yes	No	558	4:7:7	mef	51	76,272	227,736	2,017,530	43	2,013,125
SAMN05222929	20160578	СТ	2015	35B	Yes	No	10493	4:7:7	mef	74	81,944	305,405	2,071,800	47	2,060,517
SAMN05222937	20160586	СТ	2015	35B	Yes	No	558	4:7:7	mef	71	63,833	139,228	2,013,553	59	2,006,086
SAMN05222969	20160886	GA	2015	35B	Yes	No	558	4:7:7	mef	114	35.631	159,040	2.069.781	103	2.061.803
SAMN05222989	20160906	NM	2015	35B	Yes	No	558	4:7:7	Negative	47	87.665	172.854	2.013.642	40	2.009.199
SAMN05222997	20160914	NM	2015	35B	Yes	No	10493	4:7:7	mef	54	82.019	179.360	2.080.791	51	2.078.481
SAMN05223002	20160919	NM	2015	35B	Yes	No	1092	6:7:36	folAI100L.	115	36,416	114,401	2.161.903	104	2.154.336
									folPins195		,	,	_,,		_,,
SAMN05223093	20161195	NY	2015	35B	Yes	No	156	4:12:7	mef, folAI100L, folPins178	67	60,715	149,428	2,132,963	55	2,126,325
SAMN05596832	20161250	MN	2015	35B	Yes	No	558	4:7:7	mef	63	74.782	134.247	2.022.806	53	2.017.060
SAMN05596833	20161257	MN	2015	35B	Yes	No	558	4:7:7	mef	60	78,703	113,175	2.011.397	53	2.006.497
SAMN05596835	20161304	MN	2015	35B	Yes	No	558	4:7:7	mef	58	83,268	230,802	2.009.412	47	2.002.330
SAMN05596836	20161312	MN	2015	35B	Yes	No	558	4.7.7	mef	168	25,368	105 651	2 070 391	150	2 058 010
SAMN05596837	20161318	MN	2015	35B	Yes	No	558	4:7.7	mef	62	68,201	194 581	2.010 256	53	2,004,559
SAMN05223133	20161390	GA	2015	35B	Yes	No	558	4.7.7	mef	39	112 692	232 100	2 016 345	31	2 011 820
SAMN05223145	20161402	GA	2015	35B	Yes	No	558	4:7.7	mef	43	90,976	173 428	2.048 722	36	2.044 540
2	20.01102	<i></i>	_0.0							.0	00,010		_,		_,,

												Length of			
									Non-PBP			longest		No.	No. bases
SRA accession			Year of					PBP type	resistance	No.	N50.	contia.	No. bases	contias	in contias
no.	Isolate name	Statet	isolation	Serotype	pl-1t	pl-2t	MI ST	1a:2b:2x8	determinants¶	contias	bases	bases	in contias	>1K	>1K
SAMN05223156	20161413	GA	2015	35B	Yes	No	156	4.12.7	mef folAl100l	55	70 259	279 680	2 130 702	47	2 125 944
0/10100220100	20101410	0A	2010	000	103	NO	100	7.12.1	folDine178	55	10,200	210,000	2,100,702	77	2,120,044
SAMN05222181	20161606	СТ	2015	0\/	Voc	No	156	15.12.19	mof folA11001	55	60 206	229 710	2 117 177	46	2 111 212
3AMIN03223101	20101000	01	2015	90	165	INU	150	15.12.10	falDing 170 toth	55	09,390	230,719	2,117,477	40	2,111,010
0.1.1.105000.00.0	00404040	от	0045						TOIPINS 178, TETIVI	07	400 074	470.004	0 044 007	~~	0 000 400
SAMN05223194	20161619	CI	2015	35B	Yes	NO	558	4:7:7	met	37	120,271	173,384	2,014,607	28	2,009,100
SAMN05223196	20161621	СТ	2015	35B	Yes	No	558	4:7:7	mef	52	110,264	221,047	2,053,911	35	2,045,003
SAMN05223197	20161622	СТ	2015	35B	Yes	No	558	4:7:7	mef	38	93,266	268,577	2,007,538	29	2,002,683
SAMN05223202	20161627	СТ	2015	35B	Yes	No	558	4:7:7	mef	37	109,078	329,557	2,008,734	31	2,005,184
SAMN05223206	20161631	СТ	2015	35B	Yes	No	558	4:120:7	mef	38	112,348	283,017	2,051,468	29	2,045,940
SAMN05223222	20161647	MD	2015	35B	No	No	11818	1.31.111	ormB folDins105	56	117 080	273 832	2,003,564	35	2 083 044
OAIWIN0J22J222	20101047		2015	550	NO	INU	11010	4.51.114		50	117,300	210,002	2,035,504	55	2,003,044
CANINOCO00044	00404000		0045		Vee	Nia	0004	4.7.7	Negative	40	00 770	470.005	0 000 077	20	0.077.000
SAMIN05223244	20161669	IVID	2015	35B	res	INO	6961	4:7:7	Negative	46	89,773	173,235	2,082,077	30	2,077,396
SAMN05223249	20161674	MD	2015	35B	Yes	No	558	4:7:7	met	52	93,157	157,673	2,021,717	41	2,016,287
SAMN05596838	20161697	MD	2015	35B	Yes	No	558	4:7:7	mef	76	49,448	128,234	2,023,535	69	2,018,951
SAMN05596839	20161763	TN	2015	35B	Yes	No	156	4:12:7	mef, folAI100L,	74	61,347	118,236	2,139,347	61	2,130,933
									folPins178						
SAMN05596840	20161772	ΤN	2015	35B	Yes	No	558	4:7:7	mef	35	94,121	214,007	2.007.894	30	2.004.814
SAMN055968/1	20161706	TN	2015	35B	Vos	No	558	1.7.7	mof	50	104 680	260,238	2 015 030	34	2,000,670
CAMNO5530041	20101730		2015	350	Vee	No	550	4.7.7	men	50	70,000	200,230	2,013,353	42	2,003,073
SAMIN05596842	20161806		2015	358	res	INO	558	4:7:7	mer	53	70,235	170,203	2,017,465	43	2,011,036
SAMN05596843	20161992	CA	2015	35B	NO	NO	452	0:0:0	Negative	93	68,407	129,046	2,125,843	61	2,109,508
SAMN05596848	20162335	TN	2015	35B	Yes	No	558	4:7:7	mef	79	81,310	165,420	2,040,006	51	2,031,050
SAMN05596849	20162342	ΤN	2015	35B	Yes	No	156	4:12:7	mef, folAI100L,	164	38,112	116,941	2,132,394	90	2,109,381
									folPins178						
SAMN05596853	20162573	NM	2015	35B	Yes	No	558	4:7:7	mef	46	95,570	165,406	2,005,856	35	2,000,856
SAMN05751705	20162860	TN	2015	35B	Yes	No	558	102.7.7	mef	52	92 709	202 610	1 998 583	34	1 990 417
SAMN05596864	20163113	MN	2015	35B	Yes	No	558	4.7.7	mef	50	89,836	221 579	2 017 514	40	2 012 326
CAMNOEE00072	20100110	TN	2015	250	Vee	No	11504	4.40.7	mof fold 11001	40	70,110	400 475	2,017,014	40	2,012,020
SAIWINU5596673	20103079	I IN	2015	300	res	INO	11564	4.12.7	folPins178	40	79,119	492,175	2,132,330	40	2,127,526
SAMN05617281	20164527	CO	2015	9V	Yes	No	156	15:12:18	mef, folAl100L,	76	105,812	265,417	2,096,832	45	2,082,189
	20162014	C A	2016	250	Vee	No	FFO	4.7.7	IUIF II IS I 70	25	150 017	277 400	2 012 205	26	2 007 696
SAMINU5590044	20162014	CA	2016	330	res	INO	220	4.7.7	mei, ioiPins i 60	30	150,617	377,102	2,013,395	20	2,007,000
SAMIN05596845	20162040	GA	2016	35B	Yes	INO	558	4:7:7	mer	44	106,033	260,232	2,016,925	30	2,008,717
SAMN05596846	20162045	GA	2016	35B	Yes	No	558	4:7:7	mef	60	84,502	146,825	2,013,691	45	2,006,344
SAMN05596847	20162312	NY	2016	35B	Yes	No	558	4:7:7	mef	34	108,278	267,785	2,008,023	26	2,003,188
SAMN05596850	20162366	GA	2016	35B	Yes	No	11866	4:7:7	mef	42	105,288	165,485	2,012,415	34	2,007,634
SAMN05596851	20162388	GA	2016	35B	Yes	No	558	4:7:7	mef	32	123.819	268.582	2.014.483	23	2.009.884
SAMN05596852	20162390	GA	2016	35B	Yes	No	156	4.15.2	mef folAl100l	53	69 911	361 463	2 129 840	42	2 123 871
0, 101100000002	20102000	<u>O</u>	2010	000	100	110	100	7.12.7	folPins178	00	00,011	001,400	2,120,040	72	2,120,071
SAMN05596854	20162889	ΤN	2016	35B	Yes	No	156	4:12:7	mef. folAI100L	61	63.143	149.031	2.092.555	50	2.086.601
									folPins178	•	,	,	_,,		_,,
SAMN05596856	20162912	СТ	2016	35B	Yes	No	156	4:12:7	mef, folAI100L,	49	90,635	216,786	2,130,046	43	2,126,635
0 4 M NOT 500057	00400047	0T	0040	055	Ver	N -		4.7.7	toiPins178	70	00 400	005 050	0.040.040	50	0.040.400
SAMINU5596857	20162917		2016	35B	Yes	NO	558	4:/:/	Negative	70	63,408	205,652	2,049,848	58	2,042,132
SAMN05596858	20162920	CT	2016	35B	No	No	452	0:0:0	Negative	78	73,710	186,612	2,085,665	50	2,071,103
SAMN05596859	20162941	MD	2016	35B	Yes	No	558	4:7:7	mef	44	106,088	253,376	2,043,264	34	2,037,582
SAMN05596860	20162948	MD	2016	35B	Yes	No	558	4:7:7	Negative	75	63,449	150,746	2,043,703	55	2,035,188
SAMN05596861	20162969	MD	2016	35B	Yes	No	558	4:7:7	mef	57	75,713	175.536	2,015.950	49	2,011.333
SAMN05596862	20162972	MD	2016	35B	Yes	No	558	106:7:7	Negative	54	77145	178,983	2,014,160	42	2,007,683
2	_0.0 _ 0.2		-0.0							•••			_, • · ·, · • •		_,,

												Length of			
									Non-PBP			longest		No.	No. bases
SRA accession			Year of					PBP type	resistance	No.	N50,	contig,	No. bases	contigs	in contigs
no.	Isolate name	State [†]	isolation	Serotype	pl-1‡	pl-2‡	MLST	1a:2b:2x§	determinants¶	contigs	bases	bases	in contigs	>1K	>1K
SAMN05596863	20163008	NY	2016	35B	Yes	No	558	4:123:7	mef	38	112,805	313,544	2,006,543	26	2,001,014
SAMN05596865	20163194	MN	2016	35B	Yes	No	558	4:7:7	mef	56	89.735	262.271	2.012.685	38	2.004.614
SAMN05596866	20163213	MN	2016	35B	Yes	No	558	4:7:7	mef	58	91,575	256,172	2,057,438	41	2,048,656
SAMN05596867	20163215	MN	2016	35B	Yes	No	558	4.7.7	mef	38	104 755	221 865	2 008 017	.32	2 004 469
SAMN05596868	20163410	NM	2016	35B	No	No	452	0.0.0	Negative	68	92 976	206 750	2 128 641	42	2 115 150
SAMN05596869	20163480	GA	2016	35B	Vec	No	558	0.0.0 1.7.7	mof	33	106 001	313 /10	2,120,041	27	2,110,100
SAMN05596009	20163500	CA CA	2010	35B	Voc	No	550	4.7.7	folA11001	43	86 577	222 544	2,013,072	27	2,003,500
SAMM05590070	20103309		2010	35D 25D	Vee	No	156	4.7.7	mof folAl100L	43	00,377	223,344	2,049,007	20	2,040,110
SAMIN05590671	20103042	IIN	2010	330	res	INU	150	4.12.7	folPins178	52	90,405	210,435	2,127,197	39	2,120,744
SAMN05596872	20163669	ΤN	2016	35B	Yes	No	558	4:7:7	mef	34	120,612	268,637	2,015,492	28	2,011,925
SAMN05596874	20163701	OR	2016	35B	No	No	452	0:0:0	Negative	66	94,,801	195,539	2,081,512	40	2,068,754
SAMN05596875	20163728	NY	2016	35B	Yes	No	558	4:7:7	mef	478	26,166	115,452	2,054,312	137	1,963,751
SAMN05596876	20164142	NM	2016	35B	Yes	No	162	0:0:0	Negative	115	56,676	265,920	2,102,021	59	2,082,245
SAMN05596877	20164157	GA	2016	35B	Yes	No	10493	4:7:7	mef	44	93,170	294,685	2.038.238	33	2.033.518
SAMN06215793	20164170	GA	2016	35B	Yes	No	12854	4:7:7	mef	76	63,877	173,194	2,058,373	61	2.051.342
SAMN06215794	20164318	MN	2016	35B	Yes	No	558	4.7.7	mef	46	104 615	205 294	2 010 923	33	2 004 779
SAMN06215795	20164323	MN	2016	35B	Yes	No	558	4.7.7	mef	88	66 808	116 805	2,010,020	52	2,004,770
SAMN06215796	20164323	MD	2010	25B	No	No	10/02	4.7.7	mof	74	69 197	116 219	2,001,000	52	2,010,000
SAMN06215755	20104002		2010	35D	Voo	No	10493	4.7.7	mof	120	46 901	10,510	2,020,003	00	2,013,100
SAIVIN00213797	20104372		2016	330	Vee	No	10493	4.7.7	niei mof folklagol	120	40,001	123,721	2,039,734	00	2,023,041
SAIWIN00215796	20104307	IVID	2010	300	165	INU	150	4.12.7	folPins178	137	40,001	150,191	2,090,001	04	2,000,150
SAMN06215799	20164405	ΤN	2016	35B	Yes	No	2082189	4:7:7	mef	3217	3,817	33,659	2,279,413	359	1,568,218
SAMN06215800	20164408	ΤN	2016	35B	Yes	No	12921	4:11:7	mef, folAI100L, folPins178	97	69,811	147,008	2,126,987	57	2,111,689
SAMN06215801	20164453	СТ	2016	35B	No	No	452	0:0:0	Negative	123	50.059	120.756	2.083.262	71	2.062.939
SAMN06215802	20164455	ĊТ	2016	35B	Yes	No	558	4:7:7	Negative	78	78.527	160.641	2.017.615	50	2.005.477
SAMN06215803	20164456	CT	2016	35B	Yes	No	558	4.7.7	mef	103	45 263	95 304	2 020 183	79	2 010 278
SAMN06215804	20164465	CT	2016	35B	Yes	No	558	4.7.7	mef	70	63,940	147 951	2 019 143	54	2 012 182
SAMN06215805	20164476	CT	2016	35B	Yes	No	558	4.7.7	mef	03	63 483	182 215	2,056,675	56	2 043 534
SAMN06215806	20164485	CT	2010	35B	Yes	No	558	4.7.7	Negative	87	63 417	112 701	2,000,070	60	2,040,004
SAMN06215807	20104403		2010	35B	Vee	No	558	4.135.7	folA11001	111	17 830	125 040	2,013,204	70	2,002,000
SAMINU0213007	20104330	00	2010	336	165	NU	550	4.155.7	folPins169		47,009	125,940	2,013,033	19	2,003,477
SAMN06215808	20164579	CO	2016	35B	Yes	No	558	4:7:7	folAl100L, folPins180	108	54,780	107,696	2,011,176	66	1,998,980
SAMN06215809	20164584	СО	2016	35B	Yes	No	558	4:7:7	folAl100L, folPins169	53	94,840	237,216	2,014,730	36	2,007,411
SAMN06215810	20165166	OR	2016	35B	Yes	No	558	4.7.7	mef	167	35 168	85 353	2 013 744	95	1 993 804
SAMN06215811	20165435	NY	2016	35B	Yes	No	558	4.7.7	mef	114	49 620	165 048	1 997 966	59	1 984 526
SAMN06215812	20165446	TN	2010	35B	Vec	No	558	4.7.7 A·7·7	mof	52	106 037	230 738	2 021 1/8	31	2 012 234
SAMN06215912	20165460	TN	2010	25B	Voc	No	550	4.7.7	Nogotivo	91	79 / 21	140 820	2,021,140	51	2,012,204
SAIVIN00213013	20103400		2016	30D 25D	Vee	No	550	4.7.7	megative	0 I 5 0	10,421	140,020	2,010,704	42	2,000,303
SAIVIN00213014	20100010	GA	2010	300	Vee	NO No	556	4.7.7	mel	30	91,400	222,902	2,031,473	43	2,043,757
SAIVINU6215815	20165562	IVIIN	2016	358	res	INO	558	4:7:7	mer	70	89,761	201,707	2,016,051	40	2,003,973
SAIVINU6215816	20165585	IVIN	2016	358	res	INO	558	4:7:7	mer	99	51,117	165,382	2,013,853	62	2,000,883
SAMN06215817	20165590	MN	2016	35B	Yes	No	558	4:7:7	met	116	49,676	165,343	2,019,643	76	2,005,318
SAMN06215818	20165613	MN	2016	35B	Yes	No	558	4:139:7	met	61	81,988	172,386	2,039,499	46	2,031,802
SAMN06215819	20165637	MN	2016	35B	Yes	No	558	4:7:7	mef	51	82,907	202,647	2,043,980	40	2,038,852
SAMN06215820	20165685	MN	2016	35B	Yes	No	12883	4:7:7	foIAI100L	57	68,109	253,901	2,018,465	43	2,012,245
SAMN06215821	20165915	СТ	2016	35B	Yes	No	12885	4:7:7	mef	67	80,988	261,406	2,019,225	46	2,011,232
SAMN06215822	20165986	NY	2016	35B	Yes	No	558	4:7:7	mef	84	78,417	165,496	2,086,489	51	2,072,015
SAMN06215823	20165996	СТ	2016	35B	Yes	No	558	4:120:7	mef	65	76,906	162,547	2,051,848	46	2,043,512

												Length of			
									Non-PBP			longest		No.	No. bases
SRA accession			Year of					PBP type	resistance	No.	N50,	contig,	No. bases	contigs	in contigs
no.	Isolate name	State [†]	isolation	Serotype	pl-1‡	pl-2‡	MLST	1a:2b:2x§	determinants¶	contigs	bases	bases	in contigs	>1K	>1K
SAMN06215824	20166006	MD	2016	9V	Yes	No	156	15:12:228	mef, folAI100L,	49	104,699	567,749	2,048,506	32	2,038,541
									folPins178						
SAMN06215825	20166007	MD	2016	35B	Yes	No	558	4:7:7	foIAI100L	34	144,034	329,701	2,048,856	25	2,043,137
SAMN06215826	20166027	MD	2016	35B	Yes	No	558	4:7:7	Negative	40	108,014	257,659	2,002,985	31	1,998,164
SAMN06215827	20166030	MD	2016	35B	Yes	No	12922	4:142:7	Negative	44	105,529	221,286	2,011,129	33	2,005,703
SAMN06215828	20166031	MD	2016	35B	Yes	No	558	4:7:7	mef	41	98,102	268,599	2,024,426	33	2,019,673
SAMN06215829	20166040	MD	2016	35B	Yes	No	558	4:7:7	Negative	33	120,367	282,949	2,012,657	28	2,009,526
SAMN06215830	20166079	GA	2016	35B	Yes	No	558	4:7:7	mef	32	105,548	237,002	2,010,391	28	2,008,033
SAMN06215831	20166343	OR	2016	35B	No	No	452	0:0:0	Negative	65	64,765	181,330	2,073,513	47	2,064,230
SAMN06555337	20166603	MN	2016	35B	Yes	No	558	4:7:7	mef	70	88,319	173,091	2,004,297	39	1,994,204

*MLST, multilocus sequence type; N50, average length of contigs; PBP, penicillin-binding protein; SRA, sequence read archive.

†CA, California; CO, Colorado; CT, Connecticut; GA, Georgia; MD, Maryland; MN, Minnesota; NM, New Mexico; NY, New York; OR, Oregon; TN, Tennessee.

‡Pilus subunit pl-1 and pl-2 genes.

§See Li et al. (1) and MIC correlates for PBP types (http://www.cdc.gov/streplab/mic-tables.html).

¶For a description of whole-genome sequence—based bioinformatic pipeline for deduction of all features shown, see Li et al. (1) and Metcalf et al. (2,3). See Figure 1 in Metcalf et al. (3) for a description of folP insertions (folPins178, folP189).

#European Nucleotide Archive accession number.

References

- Li Y, Metcalf BJ, Chochua S, Li Z, Gertz RE Jr, Walker H, et al. Penicillin-binding protein transpeptidase signatures for tracking and predicting βlactam resistance levels in *Streptococcus pneumoniae*. MBio. 2016;7:e00756–16. <u>PubMed http://dx.doi.org/10.1128/mBio.00756-16</u>
- Metcalf BJ, Gertz RE Jr, Gladstone RA, Walker H, Sherwood LK, Jackson D, et al.; Active Bacterial Core surveillance team. Strain features and distributions in pneumococci from children with invasive disease before and after 13-valent conjugate vaccine implementation in the USA. Clin Microbiol Infect. 2016;22:60.e9–29. <u>PubMed http://dx.doi.org/10.1016/j.cmi.2015.08.027</u>
- <jrn>3. Metcalf BJ, Chochua S, Gertz RE Jr, Li Z, Walker H, Tran T, et al.; Active Bacterial Core surveillance team. Using whole genome sequencing to identify resistance determinants and predict antimicrobial resistance phenotypes for year 2015 invasive pneumococcal disease isolates recovered in the United States. Clin Microbiol Infect. 2016;22:1002.e1–8. PubMed http://dx.doi.org/10.1016/j.cmi.2016.08.001