

Table of Contents

NARMS Working Group, 2001

On-line Materials

Part I. Narrative Report

| | |
|---|----|
| Summary of 2001 data | 9 |
| Surveillance and Laboratory Testing Methods | 11 |
| Results | 14 |
| Summary of Long Term Changes..... | 24 |
| References | 27 |
| Publications and Abstracts, 2001 | 28 |

Part II. Summary Tables and Graphs

Tables

| | |
|--|----------|
| Population size and number of isolates received and tested, by site, 2001 | Table 1 |
| Antimicrobial agents used for susceptibility testing for <i>Salmonella</i> , <i>Shigella</i> , <i>E. coli</i> O157, and <i>Campylobacter</i> isolates, 2001 | Table 2 |
| Results of ceftriaxone confirmatory testing for non-Typhi <i>Salmonella</i> isolates meeting screening criteria, 2001 | Table 3 |
| Antimicrobial resistance of <i>Salmonella</i> , <i>Shigella</i> , and <i>E. coli</i> O157 isolates, 2001 | Table 4 |
| Additional resistance by antimicrobial agent for non-Typhi <i>Salmonella</i> , 2001..... | Table 5a |
| Additional resistance by antimicrobial agent for <i>Salmonella</i> Typhi, 2001..... | Table 5b |
| Additional resistance by antimicrobial agent for <i>Shigella</i> , 2001..... | Table 5c |
| Additional resistance by antimicrobial agent for <i>E. coli</i> O157, 2001 | Table 5d |
| Additional resistance by antimicrobial agent for <i>Campylobacter</i> , 2001 | Table 5e |
| The 16 most common non-Typhi <i>Salmonella</i> serotypes, 2001..... | Table 6 |

| | |
|--|----------|
| Frequency of pansusceptibility, and resistance to one or more antimicrobial agents, among the 16 most common non-Typhi <i>Salmonella</i> serotypes, 2001 | Table 7 |
| Frequency of resistance and multidrug resistance among the 16 most common non-Typhi <i>Salmonella</i> serotypes, 2001..... | Table 8 |
| Frequency of multidrug resistance among the most common non-Typhi <i>Salmonella</i> serotypes in each site, 2001 | Table 9 |
| <i>Salmonella</i> Typhimurium isolates with at least ACSSuT, ACKSSuT, or AKSSuT resistance patterns, by site, 2001 | Table 10 |
| Additional antimicrobial resistance for <i>Salmonella</i> Typhimurium isolates with at least ACSSuT, ACKSSuT, or AKSSuT resistance patterns, 2001 | Table 11 |
| <i>Salmonella</i> Newport isolates with at least MDR-AmpC resistance pattern, by site, 2001 | Table 12 |
| Additional antimicrobial resistance for <i>Salmonella</i> Newport Isolates with at least MDR-AmpC resistance pattern, 2001 | Table 13 |
| Most common multidrug resistance patterns among non-Typhi <i>Salmonella</i> , 2001 | Table 14 |
| Clinical source of non-Typhi <i>Salmonella</i> isolates, 2001 | Table 15 |
| Proportion of non-Typhi <i>Salmonella</i> isolates (N=1419) with clinically important resistance: decreased susceptibility or resistance to ciprofloxacin or ceftriaxone, 2001 | Table 16 |
| Proportion of non-Typhi <i>Salmonella</i> isolates submitted, by site, with decreased susceptibility to ciprofloxacin ($\text{MIC} \geq 0.25 \mu\text{g/ml}$), 2001 | Table 17 |
| Proportion of non-Typhi <i>Salmonella</i> isolates submitted, by site, with decreased susceptibility to ceftriaxone ($\text{MIC} \geq 16 \mu\text{g/ml}$), 2001 | Table 18 |
| Antimicrobial resistance of non-Typhi <i>Salmonella</i> isolates, 1996-2001..... | Table 19 |
| Multidrug resistance of non-Typhi <i>Salmonella</i> isolates, 1996-2001 | Table 20 |
| Antimicrobial resistance of <i>Salmonella</i> Typhi isolates, 1999-2001 | Table 21 |
| Antimicrobial resistance of <i>E. coli</i> O157 isolates, 1996-2001 | Table 22 |
| Frequency of <i>Shigella</i> species, 2001 | Table 23 |
| Antimicrobial resistance of <i>Shigella</i> isolates, by <i>Shigella</i> species, 2001 | Table 24 |

| | |
|---|----------|
| Antimicrobial resistance of <i>Shigella</i> isolates, 1999-2001 | Table 25 |
| Antimicrobial resistance of <i>Shigella sonnei</i> isolates, 1999-2001 | Table 26 |
| Antimicrobial resistance of <i>Shigella flexneri</i> isolates, 1999-2001 | Table 27 |
| Frequency of <i>Campylobacter</i> species, 2001 | Table 28 |
| Antimicrobial resistance of <i>Campylobacter</i> isolates, by <i>Campylobacter</i> species, 2001 | Table 29 |
| Antimicrobial resistance of <i>Campylobacter jejuni</i> isolates, by site, 2001 | Table 30 |
| Antimicrobial resistance of <i>Campylobacter</i> isolates, 1997-2001 | Table 31 |
| Antimicrobial resistance of <i>Campylobacter jejuni</i> isolates, 1997-2001 | Table 32 |
| Antimicrobial resistance of <i>Campylobacter coli</i> isolates, 1997-2001 | Table 33 |
| Frequency of resistance and multidrug resistance among <i>Salmonella</i> Typhi, <i>Shigella</i> , <i>E. coli</i> O157, and <i>Campylobacter</i> isolates, 2001 | Table 34 |

Graphs

| | |
|--|---------------|
| Number of isolates submitted, by site, 2001 | Figure 1 |
| MICs among non-Typhi <i>Salmonella</i> isolates, by antimicrobial agent, 1996-2001 | Figures 2a-2q |
| Percent of non-Typhi <i>Salmonella</i> isolates (N=1419) that were pansusceptible, and percent resistant to ≥ 1 , ≥ 2 , ≥ 5 , and ≥ 8 of the 18 antimicrobial agents tested, 2001 | Figure 3 |
| Resistance among non-Typhi <i>Salmonella</i> isolates, 1996-2001 | Figure 4 |
| Resistance among the most common non-Typhi <i>Salmonella</i> serotypes, 1996-2001 | Figures 5a-5p |
| Percent of non-Typhi <i>Salmonella</i> isolates that are serotype Typhimurium, by site, 1996-2001 | Figure 6 |
| Percent of <i>Salmonella</i> Typhimurium isolates that are resistant to at least ampicillin, chloramphenicol, streptomycin, sulfamethoxazole, and tetracycline (ACSSuT), by site, 1996-2001 | Figure 7 |
| Percent of <i>Salmonella</i> Typhimurium isolates that are resistant to at least ampicillin, kanamycin, streptomycin, sulfamethoxazole, and tetracycline (AKSSuT), by site, 1996-2001 | Figure 8 |
| Percent of non-Typhi <i>Salmonella</i> isolates that are serotype Newport, by site, 1996-2001 | Figure 9 |

| | |
|---|-----------------|
| Percent of <i>Salmonella</i> Newport isolates that are at least MDR-AmpC [†] , by site, 1996-2001 | Figure 10 |
| Resistance among <i>Salmonella</i> Typhi isolates, 1999-2001 | Figure 11 |
| MICs among <i>Salmonella</i> Typhi isolates, by antimicrobial agent, 1999-2001 | Figures 12a-12q |
| Resistance among <i>Shigella</i> isolates, 1999-2001 | Figure 13 |
| Resistance among <i>Shigella sonnei</i> isolates, 1999-2001 | Figure 14a |
| Resistance among <i>Shigella flexneri</i> isolates, 1999-2001 | Figure 14b |
| MICs among <i>Shigella sonnei</i> isolates, by antimicrobial agent, 1999-2001..... | Figures 15a-15q |
| MICs among <i>Shigella flexneri</i> isolates, by antimicrobial agent, 1999-2001..... | Figures 16a-16q |
| Resistance among <i>E. coli</i> O157 isolates, 1996-2001 | Figure 17 |
| MICs among <i>E. coli</i> O157 isolates, by antimicrobial agent, 1996-2001 | Figures 18a-18q |
| Resistance among <i>Campylobacter</i> isolates, 1997-2001 | Figure 19 |
| Resistance among <i>Campylobacter jejuni</i> isolates, 1997-2001 | Figure 20a |
| Resistance among <i>Campylobacter coli</i> isolates, 1997-2001 | Figure 20b |
| MICs among <i>Campylobacter jejuni</i> isolates, by antimicrobial agent, 1997-2001 | Figures 21a-21h |
| MICs among <i>Campylobacter coli</i> isolates, by antimicrobial agent, 1997-2001 | Figures 22a-22h |

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Materials Available On-Line

All CDC NARMS Annual Reports are posted on the CDC NARMS Website. The address is www.cdc.gov/narms

Salmonella Annual Summaries are posted on the PHLIS website. The address is www.cdc.gov/ncidod/dbmd/phlisdata/salmonella.htm

Shigella Annual Summaries are also posted on the PHLIS website. The address is www.cdc.gov/ncidod/dbmd/phlisdata/shigella.htm

Information on animal isolates in NARMS is available on the USDA-ARS website at:
<http://www.ars-grin.gov/ars/SoAtlantic/Athens/arru/narms.html>

General information about the NARMS surveillance program is posted on the FDA Center for Veterinary Medicine website at:

http://www.fda.gov/cvm/index/narms/narms_pg.html

Part I

Narrative Report

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Human Isolates Final Report, 2001

Summary of 2001 Data

There were 17 health departments, representing approximately 109 million persons (40% of the United States population), participating in the National Antimicrobial Resistance Monitoring System (NARMS) for Enteric Bacteria in 2001. Antimicrobial resistance among *Campylobacter* isolates was monitored in nine NARMS participating states; resistance among the other bacteria was monitored in all NARMS participating states.

The following key findings were observed in 2001:

Multidrug Resistance

Three multidrug-resistant strains accounted for 10% (142/1419) of non-Typhi *Salmonella* isolates tested by NARMS in 2001.

- In 2001, 7% (96/1419) of non-Typhi *Salmonella* isolates were *Salmonella* Typhimurium ACSSuT (resistant to at least ampicillin, chloramphenicol, streptomycin, sulfamethoxazole, and tetracycline), compared with 8% (103/1326) in 1996. These resistant isolates comprised 30% of *Salmonella* Typhimurium in 2001.
- In 2001, 1% (15/1419) of non-Typhi *Salmonella* isolates were *Salmonella* Typhimurium AKSSuT (resistant to at least ampicillin, kanamycin, streptomycin, sulfamethoxazole, and tetracycline), compared with 2% (27/1326) in 1996. These resistant isolates comprised 5% of *Salmonella* Typhimurium in 2001.
- In 2001, 2% (31/1419) of non-Typhi *Salmonella* isolates were *Salmonella* Newport MDR-AmpC (resistant to at least amoxicillin/clavulanic acid, ampicillin, cephalothin, ceftiofur, cefoxitin, chloramphenicol, streptomycin, sulfamethoxazole, tetracycline, and with

decreased susceptibility [minimum inhibitory concentration (MIC) $\geq 16\mu\text{g/ml}$] to ceftriaxone. In 1996, none were detected. These resistant isolates comprised 25% of *Salmonella* Newport in 2001.

Clinically Important Resistance

Fluoroquinolones (e.g., ciprofloxacin) and third generation cephalosporins (e.g., ceftriaxone) are commonly used antimicrobial agents for the treatment of severe *Campylobacter* and *Salmonella* infections, including *Salmonella* serotype Typhi. Nalidixic acid is an elementary quinolone; resistance to nalidixic acid correlates with decreased susceptibility to ciprofloxacin. An important proportion of isolates tested by NARMS in 2001 demonstrated resistance or decreased susceptibility to these clinically important antimicrobials.

- In 2001, 19% (75/384) of *Campylobacter* isolates were resistant to ciprofloxacin, compared with 13% (28/217) in 1997.
- In 2001, 1% (15/1419) of non-Typhi *Salmonella* isolates had decreased susceptibility (MIC $\geq 0.25\mu\text{g/ml}$) to ciprofloxacin and 0.2% (3/1419) were resistant to ciprofloxacin. In 1996, 0.4% (5/1326) of non-Typhi *Salmonella* isolates had decreased susceptibility to ciprofloxacin and none were resistant.
- In 2001, 3% (48/1419) of non-Typhi *Salmonella* isolates had decreased susceptibility (MIC $\geq 16\mu\text{g/ml}$) to ceftriaxone and 2% (34/1419) were resistant to ceftriaxone. In 1996, 0.1% (1/1326) had decreased susceptibility and 0.1% (1/1326) were resistant.
- In 2001, 18% (35/197) of *Salmonella* Typhi isolates were resistant to ampicillin, chloramphenicol, and trimethoprim-sulfamethoxazole, compared with 12% (20/166) in 1999.
- In 2001, 30% (59/197) of *Salmonella* Typhi isolates were nalidixic acid-resistant, compared with 23% (41/177) in 2000 and 19% (31/166) in 1999.

Surveillance and Laboratory Testing Methods

NARMS is a collaboration between the Centers for Disease Control and Prevention (CDC) National Center for Infectious Diseases, the U.S. Food and Drug Administration (FDA) Center for Veterinary Medicine, the U.S. Department of Agriculture (USDA) Food Safety and Inspection Service, the USDA Agricultural Research Service, and 17 state and local health departments. CDC monitors antimicrobial resistance in enteric bacteria isolated from humans, USDA monitors antimicrobial resistance in enteric bacteria isolated from animals, and FDA monitors antimicrobial resistance in enteric bacteria isolated from meats.

Many NARMS activities are conducted within the framework of CDC's Emerging Infections Program's Epidemiology and Laboratory Capacity Program and the Foodborne Disease Active Surveillance Network (FoodNet). The primary purpose of NARMS is to monitor antimicrobial resistance among foodborne enteric bacteria isolated from humans. NARMS data are also used to provide platforms for additional studies including field investigations and molecular characterization of resistance determinants, and to guide efforts to mitigate antimicrobial resistance.

NARMS began in 1996 to monitor prospectively the antimicrobial resistance of human non-Typhi *Salmonella* and *Escherichia coli* O157 isolates. Testing of human *Campylobacter* isolates was added in 1997, and testing of human *Salmonella* Typhi and *Shigella* isolates was added in 1999. Before NARMS was established, CDC monitored antimicrobial resistance in *Salmonella*, *Shigella*, and *Campylobacter* using periodic surveys of isolates from a panel of sentinel counties. In 2001, there were 17 NARMS health department participants (California, Colorado, Connecticut, Florida, Georgia, Kansas, Los Angeles County, Massachusetts, Maryland, Minnesota, New Jersey, New York City, New York State, Oregon, Tennessee, Washington, and West Virginia), representing approximately 109 million persons (40% of the United States population) [Table 1]. In 2001, the nine NARMS participating state health departments (California, Colorado, Connecticut, Georgia, Maryland, Minnesota, New York, Oregon, and Tennessee) that also participate in FoodNet monitored antimicrobial resistance among human *Campylobacter* isolates [Table 1].

In 2001, NARMS participating public health laboratories systematically selected every tenth non-Typhi *Salmonella*, every *Salmonella* Typhi, every tenth *Shigella*, and every fifth *E. coli* O157 isolate received at their laboratory, and forwarded the isolates to CDC for susceptibility testing. Non-Typhi *Salmonella* refers to all *Salmonella* serotypes except serotype Typhi. At CDC, *Salmonella*, *Shigella*, and *E. coli* O157 isolates are tested with a semi-automated system (Sensititre[®], Trek Diagnostics, Westlake, OH) to determine the partial range MIC for 17 antimicrobial agents: amikacin, ampicillin, amoxicillin-clavulanic acid, apramycin, cefoxitin, ceftiofur, ceftriaxone, cephalothin, chloramphenicol, ciprofloxacin, gentamicin, kanamycin, nalidixic acid, streptomycin, sulfamethoxazole, tetracycline, and trimethoprim-sulfamethoxazole [Table 2]. Isolates meeting screening criteria for possible ceftriaxone resistance (ceftiofur MIC \geq 4 $\mu\text{g}/\text{ml}$ and / or ceftriaxone MIC \geq 2 $\mu\text{g}/\text{ml}$, via Sensititre[®]), were tested for ceftriaxone susceptibility with by-hand broth microdilution. Before 1999, isolates meeting the screening

criteria were additionally tested to confirm ceftriaxone susceptibility results with the E-test system (AB BIODISK, Solna, Sweden). In this report, with the exception of the Sensititre® MIC data reported in Figures 2g, 12g, 15g, 16g, and 18g, ceftriaxone results are based upon the confirmatory testing (by-hand broth microdilution or E-test system).

The nine state public health departments that participate in FoodNet forwarded *Campylobacter* isolates to CDC for susceptibility testing. Selection of *Campylobacter* isolates to submit to NARMS was conducted by either of two ways. In Georgia, Maryland, Minnesota, New York, and Tennessee, one isolate a week was selected (usually the first isolate received each week is selected, but otherwise isolates were randomly selected) from the collection of isolates sent to the state health department laboratory from almost all clinical laboratories in a geographical area (metro Atlanta area in Georgia, statewide in Maryland and Minnesota, metro Albany and Rochester areas in New York, and Gallatin, Knoxville, and Nashville areas in Tennessee). In California, Colorado, Connecticut, and Oregon, the first isolate isolated each week was selected at one sentinel clinical laboratory. Sentinel clinical laboratories followed routine isolation practices for *Campylobacter*. At CDC, isolates were confirmed as *Campylobacter* by dark field microscopy and oxidase test. Identification to species level was performed using the hippurate hydrolysis test. Hippurate-positive isolates were identified as *C. jejuni*. Hippurate-negative isolates were identified as *C. jejuni* by the hippuricase gene-based PCR assay or as *C. coli* based on polymerase chain reaction (PCR) test for the coli-specific *ceuE* gene. Isolates that were determined not to be *C. jejuni* or *C. coli* were referred to the National *Campylobacter* Reference Laboratory at CDC for identification using genotypic and phenotypic methods. The E-test system was used to determine the MICs for 8 antimicrobial agents: azithromycin, chloramphenicol, ciprofloxacin, clindamycin, erythromycin, gentamicin, nalidixic acid, and tetracycline [Table 2]. No more than 53 *Campylobacter* isolates, per state in 2001, were included in NARMS analyses; if more than 53 isolates were received, only the first 4 or 5 isolates received each month, depending on the number of Mondays in the month, were included.

For all pathogens in this report, MIC results were dichotomized: isolates with intermediate susceptibility were categorized as susceptible. Analysis was restricted to one isolate (per pathogen) per patient. When established, National Committee for Clinical Laboratory Standards (NCCLS) interpretive criteria were used; apramycin resistance was defined as $\text{MIC} \geq 64 \mu\text{g/ml}$ and ceftiofur resistance was defined as $\text{MIC} \geq 8 \mu\text{g/ml}$ [Table 2]. Multidrug resistance was defined as resistance to two or more antimicrobial agents.

A review of NARMS *Campylobacter* data showed a higher than expected proportion of isolates that were resistant to erythromycin and gentamicin in 1997. Therefore, all NARMS *Campylobacter* isolates with any resistance in 1997 were re-tested. This report reflects these more recent test results. Furthermore, it was observed that more than 53 *Campylobacter* isolates were submitted to CDC NARMS from Georgia (N=81) and Minnesota (N=59) in 1998, and New York (N=54) in 1999. The additional isolates, from those states for those years, were excluded from analyses until no more than the maximum number of isolates (N=53) per year, in each state, were included in NARMS. This report also reflects these changes.

Logistic regression analyses were performed to assess the change in antimicrobial resistance among *Salmonella* isolates tested in NARMS in 1996 and 2001. These included 1)

ceftriaxone, ciprofloxacin, and nalidixic acid resistance among non-Typhi *Salmonella*, 2) nalidixic acid resistance among *Salmonella* Typhi, 3) multidrug resistance among *Salmonella* serotype Newport, and 4) multidrug resistance among *Salmonella* serotype Typhimurium. Multivariate logistic regression analyses were performed to assess the change in ciprofloxacin resistance among *Campylobacter* spp. and *Campylobacter jejuni* isolates tested in NARMS in 1997 and 2001. All final multivariate models took into account both population variation and site variation.

When describing results from several years, multidrug resistance for *Salmonella*, *E. coli* O157, and *Shigella* isolates was limited to the 15 agents tested in all years from 1996 to 2001 (amoxicillin-clavulanic acid, ampicillin, apramycin, ceftiofur, ceftriaxone, cephalothin, chloramphenicol, ciprofloxacin, gentamicin, kanamycin, nalidixic acid, streptomycin, sulfamethoxazole, tetracycline, trimethoprim-sulfamethoxazole). Similarly, when describing multidrug resistance for several years for *Campylobacter* isolates, multidrug resistance was limited to the 6 agents tested in all years from 1997 to 2001 (chloramphenicol, ciprofloxacin, clindamycin, erythromycin, nalidixic acid, tetracycline).

Results

Non-Typhi *Salmonella*

Results for 2001

A total of 1469 non-Typhi *Salmonella* isolates were received at CDC in 2001 [Figure 1]; of these isolates, 1462 (99%) were viable and tested for antimicrobial susceptibility. Of these 1462 isolates, 43 isolates were eliminated from analysis because they were duplicate submissions (30 isolates) or the county of residence was outside the catchment area (13 isolates), leaving 1419 isolates tested for susceptibility to 17 antimicrobial agents for analysis.

Figures 2a-2q provide Sensititre® MIC results for the non-Typhi *Salmonella* isolates for each of the 17 antimicrobial agents tested in 2001. Thirty-four (57%) of the 60 non-Typhi *Salmonella* isolates meeting the screening criteria for confirmatory testing were resistant to ceftriaxone on confirmatory testing [Table 3]. Among the 1419 non-Typhi *Salmonella* isolates, 394 (28%) were resistant to one or more antimicrobial agents, 315 (22%) were resistant to two or more agents, 176 (12%) were resistant to five or more agents, and 40 (3%) were resistant to eight or more agents [Figure 3]. Of the 176 isolates resistant to five or more agents, 114 (65%) were serotype Typhimurium. Of the 40 isolates resistant to eight or more agents, 31 (77%) were serotype Newport. The antimicrobial agents to which the 1419 *Salmonella* demonstrated the highest prevalence of resistance were tetracycline, sulfamethoxazole, streptomycin, and ampicillin: 280 (20%) were resistant to tetracycline, 251 (18%) were resistant to sulfamethoxazole, 241 (17%) were resistant to streptomycin, and 247 (17%) were resistant to ampicillin [Table 4, Figure 4]. Tables 5a through 5e show the correlation of resistance among non-Typhi *Salmonella*, *Salmonella* Typhi, *Shigella*, *E. coli* O157, and *Campylobacter* isolates, respectively. These correlation tables are useful to determine the co-resistance between and to antimicrobial agents. For example, among the 34 non-Typhi *Salmonella* isolates resistant to ceftriaxone, 34 (100%) were additionally resistant to amoxicillin-clavulanic acid, ampicillin, cefoxitin, ceftiofur, and cephalothin, 32 (94%) were resistant to streptomycin, 31 (91%) were resistant to sulfamethoxazole and tetracycline, and 30 (88%) were resistant to chloramphenicol.

Among 1419 *Salmonella* isolates tested, 1399 (99%) could be serotyped. Among the 1399 serotyped isolates, 325 (23%) were serotype Typhimurium (includes serotype Typhimurium var. Copenhagen), 282 (20%) were serotype Enteritidis, 124 (9%) were serotype Newport, 100 (7%) were serotype Heidelberg, and 51 (4%) were serotype Javiana; the 16 most common serotypes accounted for 80% (1125/1399) of isolates that were serotyped [Table 6]. The serotypes with the highest proportion of isolates that were pansusceptible (among the 16 most common serotypes) were Javiana (100%), Oranienburg (97%), and Thompson (96%); the serotypes with the highest proportion of isolates that were resistant to one or more antimicrobial agents were Agona (53%), Typhimurium (51%), and Stanley (47%) [Table 7, Table 8]. Figures 5a-5p provide susceptibility results for each of the antimicrobial agents tested, for the 16 most

common serotypes. Among the 325 S. Typhimurium isolates, 165 (51%) were resistant to one or more antimicrobial agents and 156 (48%) were multidrug resistant. Among the 282 S. Enteritidis isolates, 40 (14%) were resistant to one or more antimicrobial agents and 16 (6%) were multidrug resistant [Table 8]. Among the 124 S. Newport isolates, 43 (35%) were resistant to one or more antimicrobial agents and 39 (31%) were multidrug resistant; 79% (31/39) of the multidrug resistant S. Newport were resistant to 8 or more antimicrobials. The serotypes with the highest proportion of multidrug resistance were Typhimurium (48%), Stanley (47%), and Newport (31%). Tables 9a-9i provide data on multidrug resistance among the most common serotypes in each site.

Figure 6 shows the number of S. Typhimurium isolates submitted by site, by year. In recent years, two multidrug-resistant phenotypes of S. Typhimurium have been frequently identified. The most prominent strain is resistant to at least ampicillin, chloramphenicol, streptomycin, sulfamethoxazole, and tetracycline (ACSSuT), a phenotype commonly associated with Definitive Type 104 (DT104) [Table 10]. Among 325 S. Typhimurium isolates tested in 2001, 96 (29%) had at least the ACSSuT resistance pattern [Figure 7]. Of the 96 S. Typhimurium isolates with at least the ACSSuT resistance pattern, 13 (13%) were also resistant to amoxicillin-clavulanic acid, 4 (4%) to cephalothin, 4 (4%) to cefoxitin, 4 (4%) to ceftiofur, 4 (4%) to kanamycin, 3 (3%) to trimethoprim-sulfamethoxazole, 2 (2%) to gentamicin, 1 (1%) to ceftriaxone, 1 (1%) to ciprofloxacin, and 1 (1%) to nalidixic acid [Table 11]. A second multidrug resistant strain, resistant to at least ampicillin, kanamycin, streptomycin, sulfamethoxazole, and tetracycline (AKSSuT), was also common among S. Typhimurium [Table 10]. Among 325 S. Typhimurium isolates tested in 2001, 15 (5%) had at least the AKSSuT resistance pattern [Figure 8]. Of the 15 S. Typhimurium isolates with at least the AKSSuT resistance pattern, 4 (27%) were also resistant to chloramphenicol, 1 (7%) to amoxicillin-clavulanic acid, 1 (7%) to cefoxitin, 1 (7%) to ceftiofur, 1 (7%) to ceftriaxone, 1 (7%) to cephalothin, 1 (7%) to gentamicin, and 1 (7%) to trimethoprim-sulfamethoxazole [Table 11].

Figure 9 shows the number of S. Newport isolates submitted by site, by year. The most common multidrug-resistant phenotype among S. Newport was resistance to amoxicillin/clavulanic acid, ampicillin, cephalothin, cefoxitin, ceftiofur, chloramphenicol, streptomycin, sulfamethoxazole, tetracycline, and decreased susceptibility to ceftriaxone (MDR-AmpC). Among the 124 S. Newport isolates tested in 2001, 31 (25%) had at least the MDR-AmpC resistance pattern [Table 12, Figure 10]. Of the 31 S. Newport with at least the MDR-AmpC resistance pattern, 29 (93%) were also resistant to ceftriaxone, 6 (19%) to kanamycin, 2 (6%) to gentamicin, and 1 (3%) to trimethoprim-sulfamethoxazole [Table 13].

The three most common multidrug resistant phenotypes in 2001 were S. Typhimurium with at least the ACSSuT resistance pattern, S. Newport with at least the MDR-AmpC resistance pattern, and S. Typhimurium with at least the AKSSuT resistance pattern [Table 14]. These multidrug resistant phenotypes, respectively, accounted for 7%, 2% and 1% of all non-Typhi *Salmonella* isolates and 30%, 10%, and 5% of the multidrug resistant non-Typhi *Salmonella* isolates in 2001. Table 15 describes the clinical source of all non-Typhi *Salmonella* isolates tested in 2001. Of the 1419 non-Typhi *Salmonella* isolates, 1218 (86%) were collected from stool specimens, 71 (5%) were from blood specimens, 87 (6%) were from other sources, and 43 (3%) were from an unknown source.

Table 16 shows the proportion of non-Typhi *Salmonella* isolates that showed decreased susceptibility or resistance to ciprofloxacin or ceftriaxone. Table 17 shows the proportion of non-Typhi *Salmonella* isolates with a decreased susceptibility to ciprofloxacin by site. Fifteen non-Typhi *Salmonella* isolates (1%) had a decreased susceptibility to ciprofloxacin; of which, 3 (0.2%) were resistant to ciprofloxacin. Table 18 shows the proportion of non-Typhi *Salmonella* isolates with a decreased susceptibility to ceftriaxone by site. Forty-eight non-Typhi *Salmonella* isolates (3%) had a decreased susceptibility to ceftriaxone; of which 34 (71%) were resistant to ceftriaxone.

Changes in Antimicrobial Resistance since 1996

In 1996, 34% (103/306) of *S. Typhimurium* isolates tested had at least the ACSSuT resistance pattern [Table 19]. This proportion increased to 35% (115/326) in 1997; after which, it decreased to 32% (120/377) in 1998, 28% in both 1999 (102/362) and 2000 (84/303), and 29% (96/325) in 2001 [Figure 7]. Although the proportion of *S. Typhimurium* isolates with at least the ACSSuT resistance pattern decreased, there is no significant difference in the proportion of *S. Typhimurium* isolates with at least the ACSSuT resistance pattern in 1996 and 2001 (95% CI [0.5, 1.3]). A similar pattern was seen among *S. Typhimurium* isolates with at least the AKSSuT resistance pattern. In 1996, the prevalence of AKSSuT among *S. Typhimurium* was 9% (27/306). This proportion rose to 13% (41/326) in 1997. In subsequent years, there was a decline in prevalence of AKSSuT among *S. Typhimurium* isolates: 12% (47/377) in 1998, 11% (39/362) in 1999, 9% (28/303) in 2000 and 5% (15/325) in 2001 [Figure 8]. Although the proportion of *S. Typhimurium* isolates with at least the AKSSuT resistance pattern decreased, there is no significant difference in the proportion of *S. Typhimurium* isolates with at least the AKSSuT resistance pattern in 1996 and 2001 (95% CI [0.2, 1.1]).

In 1996 and 1997, none of the *S. Newport* isolates had at least the MDR-AmpC resistance pattern. This proportion increased to 1% (1/77) in 1998, 17% (17/98) in 1999, 22% (27/124) in 2000, and 25% (31/124) in 2001 [Figure 10]. Using an exact logistic regression model, *S. Newport* isolates in 2001 were more likely to have at least the MDR-AmpC resistance pattern ($p < .01$) than isolates in 1996.

The percentage of non-Typhi *Salmonella* isolates with decreased susceptibility to ciprofloxacin increased from 0.4% (5/1326) in 1996 to 1% (15/1419) in 2001; the percentage resistant to ciprofloxacin increased from 0% in 1996 to 0.2% in 2001. Using a logistic regression model, non-Typhi *Salmonella* isolates in 2001 were 2.8 times more likely to have a decreased susceptibility to ciprofloxacin (95% CI [1.0, 7.8]) than isolates in 1996. Additionally, the percentage of *Salmonella* isolates with decreased susceptibility to ceftriaxone increased from 0.1% (1/1326) in 1996 to 3% (48/1419) in 2001; the percentage resistant to ceftriaxone increased from 0.1% in 1996 to 2% in 2001 [Table 19]. Using a logistic regression model, non-Typhi *Salmonella* isolates were 47.7 times more likely to have a decreased susceptibility to ceftriaxone (95% CI [4.6, 492.9]) than isolates in 1996.

Limitations

The major limitations in NARMS non-Typhi *Salmonella* surveillance are the sampling scheme and the limited geographic area under surveillance. It is important to evaluate whether these limitations might be associated with an increased or decreased likelihood of antimicrobial resistance among non-Typhi *Salmonella* in NARMS.

In 2001, NARMS participating public health laboratories systematically selected every tenth non-Typhi *Salmonella* isolate received at their laboratory, and forwarded those isolates to CDC for antimicrobial susceptibility testing. NARMS participating public health laboratories are instructed to maintain a running count of non-Typhi *Salmonella* isolates received and to systematically select every tenth isolate for NARMS; when the isolates are selected, the antimicrobial resistance pattern of the isolates is not known. It is therefore unlikely that the antimicrobial resistance pattern of an isolate would influence submission of the isolate to NARMS.

There were 17 NARMS public health department participants in NARMS non-Typhi *Salmonella* surveillance in 2001, representing approximately 109 million persons or 40% of the United States population. Because NARMS 2001 non-Typhi *Salmonella* surveillance was not nationwide, generalization to the United States population should be done with caution due to potential regional differences in the prevalence of antimicrobial resistance among non-Typhi *Salmonella*. However, analyses of changes in the prevalence of resistance from 1996 to 2001 are not subject to the above limitation within the population under surveillance.

***Salmonella* Typhi**

Results in 2001

A total of 219 S. Typhi isolates were received at CDC in 2001 [Figure 1]; 211 (96%) were viable upon receipt and tested for antimicrobial susceptibility. Of these 211 isolates, 14 isolates were eliminated from analysis because they were duplicate submissions (12 isolates) or the county of residence was outside the catchment area (2 isolates), leaving 197 isolates for analysis. Among the 197 S. Typhi isolates, 81 (41%) were resistant to one or more antimicrobial agents and 45 (23%) were resistant to two or more agents [Table 34]. The most common resistances among the 197 S. Typhi isolates were to nalidixic acid (30%), chloramphenicol (21%), tetracycline (21%), trimethoprim-sulfamethoxazole (21%), ampicillin (20%), and streptomycin (20%) [Table 4, Figure 11]. Figures 12a-12q provide data on *Salmonella* Typhi MICs by antimicrobial agent. None of the S. Typhi isolates tested were resistant to amoxicillin-clavulanic acid, ceftriaxone, ciprofloxacin, or imipenem.

Changes in Antimicrobial Resistance in NARMS since 1999

In 1999, 12% (20/166) of *S. Typhi* isolates tested were resistant to at least ampicillin, chloramphenicol, and sulfamethoxazole-trimethoprim (ACSuTm). This proportion decreased to 9% (16/177) in 2000; after which, it rose to 18% (35/197) in 2001. The percentage of *S. Typhi* isolates resistant to nalidixic acid increased from 19% (31/166) in 1999 to 30% (59/197) in 2001. Using a logistic regression model, *S. Typhi* isolates in 2001 were 1.9 times more likely to be resistant to nalidixic acid (95% CI [1.1, 3.1]) than isolates in 1999.

Limitations

One limitation is evident in NARMS 2001 *Salmonella* Typhi surveillance, the limited geographic area under surveillance.

There were 17 NARMS public health department participants in NARMS *Salmonella* Typhi surveillance in 2001, representing approximately 109 million persons or 40% of the United States population. NARMS participating public health laboratories forwarded every *Salmonella* Typhi isolate received at their laboratory to CDC for antimicrobial susceptibility testing. Because NARMS 2001 *Salmonella* Typhi surveillance was not nationwide, generalization to the United States population should be done with caution due to potential regional differences in the prevalence of antimicrobial resistance among *Salmonella* Typhi.

Shigella

Results of NARMS in 2001

A total of 360 *Shigella* isolates were received at CDC in 2001 [Figure 1]; 351 (97%) were viable upon receipt and tested for antimicrobial susceptibility. Seven isolates were eliminated from analysis because they were duplicate submissions (4 isolates) or the county of residence was outside the catchment area (3 isolates), leaving 344 isolates for analysis. Of the 344 isolates analyzed, 239 (69%) were *S. sonnei*, 91 (26%) were *S. flexneri*, 5 (1%) were *S. boydii*, 3 (1%) were *S. dysenteriae*, and 6 (2%) were provisional *Shigella* species [Table 23]. Among the 344 *Shigella* isolates, 327 (95%) were resistant to one or more antimicrobial agents and 244 (71%) were multidrug resistant [Table 34]. The most common resistances among the 344 *Shigella* isolates were to ampicillin (80%), tetracycline (59%), sulfamethoxazole (56%),

streptomycin (53%), and trimethoprim-sulfamethoxazole (47%) [Table 25, Figure 13]. The 239 *Shigella sonnei* isolates were most frequently resistant to ampicillin (83%), streptomycin (54%), sulfamethoxazole (54%), and trimethoprim-sulfamethoxazole (51%) [Figure 14a]. The most common resistances among the 91 *Shigella flexneri* isolates were to tetracycline (94%), chloramphenicol (75%), and ampicillin (72%) [Figure 14b]. Figures 15a-15q and 16a-16q provide data on *Shigella sonnei* and *Shigella flexneri* MICs by antimicrobial agent. None of the *Shigella* isolates tested were resistant to amikacin, apramycin, ceftiofur, ceftriaxone, gentamicin, or imipenem. One (0.3%) *Shigella* isolate, a *Shigella flexneri* isolate, was resistant to ciprofloxacin and none had a decreased susceptibility to ceftriaxone.

Changes in Antimicrobial Resistance in NARMS since 1999

In 1999, 78% of *Shigella* isolates tested were resistant to ampicillin; this percentage increased to 79% in 2000 and 80% in 2001. From 1999 to 2001 there was a decrease in the percentage of *Shigella* isolates with the ACSuTm (ampicillin, chloramphenicol, sulfamethoxazole-trimethoprim) resistance pattern, representing 10% (37/375), 7% (31/451), and 7% (24/344) of isolates in 1999, 2000, and 2001, respectively. No *Shigella* isolates tested in 1999 and 2000 were resistant to ciprofloxacin; one isolate was resistant to ciprofloxacin in 2001.

Limitations

Two limitations are evident in NARMS *Shigella* surveillance, the sampling scheme and the limited geographic area under surveillance. It is important to evaluate whether these limitations might be associated with an increased or decreased likelihood of antimicrobial resistance among *Shigella* in NARMS.

In 2001, NARMS participating public health laboratories systematically selected every tenth *Shigella* isolate received at their laboratory, and forwarded those isolates to CDC for antimicrobial susceptibility testing. NARMS participating public health laboratories are instructed to maintain a running count of *Shigella* isolates received and to systematically select every tenth isolate for NARMS; when the isolates are selected, the antimicrobial resistance pattern of the isolates is not known. It is therefore unlikely that the antimicrobial resistance pattern of an isolate would influence submission of the isolate to NARMS.

There were 17 NARMS public health department participants in NARMS *Shigella* surveillance in 2001, representing approximately 109 million persons or 40% of the United States population. Because NARMS 2001 *Shigella* surveillance was not nationwide,

generalization to the United States population should be done with caution due to potential regional differences in the prevalence of antimicrobial resistance among *Shigella*.

E. coli O157

Results in 2001

A total of 294 *E. coli* O157 isolates were received at CDC in 2001 [Figure 1]; 284 (97%) were viable and tested for antimicrobial susceptibility. Of these 284 isolates, 7 isolates were eliminated from analysis because they were duplicate submissions (4 isolates) or the county of residence was outside the catchment area (3 isolates), leaving 277 isolates for analysis. Among the 277 *E. coli* O157 isolates, 24 (9%) were resistant to one or more antimicrobial agents and 15 (5%) were multidrug resistant [Table 34]. The most common resistances among the 277 *E. coli* O157 isolates were to sulfamethoxazole (5%), tetracycline (5%), ampicillin (2%), and streptomycin (2%) [Table 4, Figure 17]. Figures 18a-18q provide data on *E. coli* O157 MICs by antimicrobial agent. None of the *E. coli* O157 isolates tested were resistant to amikacin, apramycin, ceftriaxone, ciprofloxacin, imipenem or kanamycin. Two (1%) had a decreased susceptibility to ceftriaxone and 2 (1%) had a decreased susceptibility to ciprofloxacin.

Changes in Antimicrobial Resistance in NARMS since 1996

No *E. coli* O157 isolates were nalidixic acid-resistant from 1996 to 1998. The number of isolates resistant to nalidixic acid was 2 (1%) isolates in 1999, 2 (0.5%) in 2000, and 3 (1%) in 2001.

Limitations

Two limitations are evident in NARMS *E. coli* O157 surveillance, the sampling scheme and the limited geographic area under surveillance. It is important to evaluate whether these limitations might be associated with an increased or decreased likelihood of antimicrobial resistance among *E. coli* O157 in NARMS.

In 2001, NARMS participating public health laboratories systematically selected every fifth *E. coli* O157 isolate received at their laboratory, and forwarded those isolates to CDC for antimicrobial susceptibility testing. NARMS participating public health laboratories are instructed to maintain a running count of *E. coli* O157 isolates received and to systematically select every fifth isolate for NARMS; when the isolates are selected, the antimicrobial resistance pattern of the isolates is not known. It is therefore unlikely that the antimicrobial resistance pattern of an isolate would influence submission of the isolate to NARMS.

There were 17 NARMS health department participants in NARMS *E. coli* O157 surveillance in 2001, representing approximately 109 million persons or 40% of the United States population. Because NARMS 2001 *E. coli* O157 surveillance was not nationwide, generalization to the United States population should be done with caution due to potential regional differences in the prevalence of antimicrobial resistance among *E. coli*.

Campylobacter

Results in 2001

A total of 497 presumptive *Campylobacter* isolates were received at CDC in 2001; of these isolates, 425 (85%) were viable upon receipt. Of these 425 isolates, 41 isolates were eliminated from analysis because they were duplicate submissions (22 isolates) or were not *Campylobacter* (19 isolates), leaving 384 isolates tested for antimicrobial susceptibility to 8 antimicrobial agents for analysis. Of the 384 isolates tested, 365 (95%) were *C. jejuni*, 17 (4%) were *C. coli*, 1 (0.3%) was *C. fetus*, and 1 (0.3%) was "other" species [Table 28].

Among the 384 *Campylobacter* isolates tested, 193 (50%) were resistant to one or more antimicrobial agents and 81 (21%) were resistant to two or more agents [Table 34]. The most common resistances among the 384 *Campylobacter* isolates were to tetracycline (41%), nalidixic acid (20%), ciprofloxacin (19%), and erythromycin (2%) [Table 31, Figure 19]. None of the *Campylobacter* isolates tested were resistant to chloramphenicol or gentamicin.

Among the 365 *Campylobacter jejuni* isolates, 181 (50%) were resistant to one or more antimicrobial agents and 73 (20%) were resistant to two or more agents [Table 34]. The most common resistances among the 365 *Campylobacter jejuni* isolates were to tetracycline (40%) followed by nalidixic acid (19%), ciprofloxacin (18%), and erythromycin (2%) [Table 32, Figure 20a]. Figures 21a-21h provide data on *C. jejuni* MICs by antimicrobial agent. The proportion of ciprofloxacin-resistance *C. jejuni* varied by site: Connecticut 12/43 (28%), Georgia 14/52 (27%), California 11/46 (24%), Maryland 7/36 (19%), Tennessee 5/31 (16%), Minnesota 7/47 (15%), New York 5/44 (11%), Colorado 4/40 (10%), and Oregon 2/26 (8%) [Table 30].

Among the 17 *Campylobacter coli* isolates, 11 (65%) were resistant to one or more antimicrobial agents and 8 (47%) were resistant to or more agents [Table 34]. The most common resistances among the 17 *Campylobacter coli* isolates were to tetracycline (59%), ciprofloxacin (47%), nalidixic acid (41%), and erythromycin (6%) [Table 33, Figure 20b]. Figures 22a-22h provide data on *C. coli* MICs by antimicrobial agent.

Changes in Antimicrobial Resistance in NARMS since 1997

The percentage of *Campylobacter* isolates resistant to ciprofloxacin was 13% (28/217) in 1997, 13% (42/311) in 1998, 18% (58/318) in 1999, 14% (46/324) in 2000, and 19% (75/384) in 2001. The percentage of *Campylobacter jejuni* isolates resistant to ciprofloxacin was 12% (26/209) in 1997, 14% (41/298) in 1998, 18% (52/294) in 1999, 14% (43/306) in 2000, and 18% (67/365) in 2001. The percentage of *Campylobacter coli* isolates resistant to ciprofloxacin was 33% (2/6) in 1997, 0% (0/8) in 1998, 30% (6/20) in 1999, 25% (3/12) in 2000, and 47% (8/17) in 2001. During the same time period, the percentage of *Campylobacter* isolates resistant to nalidixic acid increased from 14% (31/217) in 1997 to 20% (77/384) in 2001; the percentage of *Campylobacter jejuni* isolates resistant to nalidixic acid increased from 13% (28/209) in 1997 to 19% (69/365) in 2001; whereas, the percentage of *Campylobacter coli* isolates resistant to nalidixic acid decreased from 50% (3/6) in 1997 to 41% (7/17) in 2001.

In the multivariate logistic regression model, the proportion of *Campylobacter* isolates resistant to ciprofloxacin in 2001, controlling for site variation and age, was 2.4 times higher (95% CI [1.4, 4.0]) than in 1997. This increase was relatively consistent, with the proportion of ciprofloxacin-resistant *Campylobacter* increasing every year compared with the previous year, except in 2000. When restricting the multivariate logistic regression analysis to *Campylobacter jejuni* isolates only, the proportion of *C. jejuni* isolates was 2.1 times more likely (95% CI [1.2, 3.6]) to be resistant to ciprofloxacin in 2001 than in 1997. This increasing trend also was relatively consistent over the years.

Limitations

Three limitations are evident in NARMS 2001 *Campylobacter* surveillance, the use of sentinel clinical laboratories in some states, the sampling scheme, and the limited geographic area under surveillance.

In four states that participated in NARMS *Campylobacter* surveillance in 2001 (California, Colorado, Connecticut, and Oregon), *Campylobacter* isolates were submitted to NARMS from one sentinel clinical laboratory. In Georgia, Maryland, Minnesota, New York, and Tennessee, the *Campylobacter* isolates submitted to NARMS were selected (usually the first

isolate received each week is selected, but otherwise isolates were randomly selected) from all *Campylobacter* isolates from most clinical laboratories within a specific geographical area (metro Atlanta area in Georgia, statewide in Maryland and Minnesota, the metro Albany and Rochester areas in New York, and the metro Gallatin, Knoxville, and Nashville areas in Tennessee). In California, Colorado, Connecticut, and Oregon, the sentinel clinical laboratory selected the first *Campylobacter* isolate isolated each week for submission to NARMS; if no isolate was isolated in a week, then no isolate was submitted from that laboratory. Since none of the sentinel clinical laboratories used an isolation procedure that was more or less likely to yield antimicrobial-resistant *Campylobacter* isolates than other clinical laboratories in their respective states, it is unlikely that the use of a sentinel clinical laboratory would be associated with an increased or decreased likelihood of antimicrobial resistance among *Campylobacter* isolates submitted to NARMS.

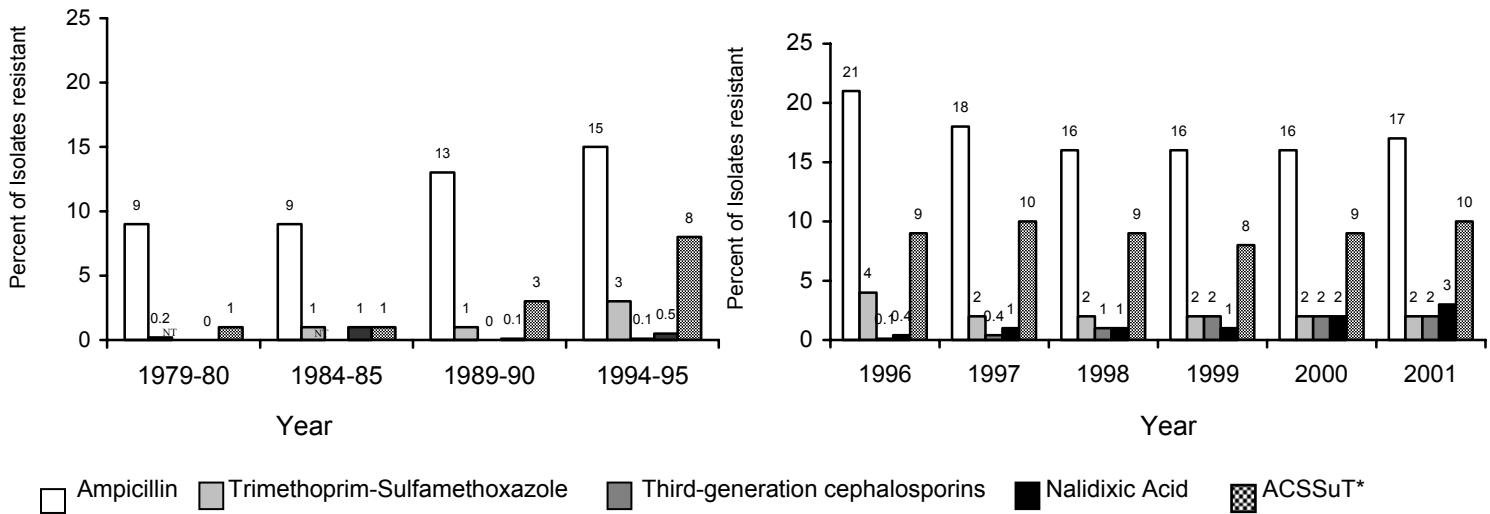
In 2001, the NARMS participating public health laboratory in Maryland, Minnesota, New York, and Tennessee, FoodNet personnel in Georgia, and sentinel clinical laboratories in all other FoodNet sites, selected one *Campylobacter* isolate each week and forwarded the isolate to CDC. When the isolates were selected, the antimicrobial resistance pattern of the isolates was not known. Therefore, it is unlikely that the antimicrobial resistance pattern of an isolate would influence submission of the isolate to NARMS. However, the one-a-week sampling scheme could result in over- or under-sampling of antimicrobial-resistant isolates if the prevalence of such resistance is not uniform throughout the year. The impact of the over- or under-sampling may be variable among states.

Campylobacter isolates were forwarded to CDC by nine FoodNet participating states in 2001, representing approximately 34 million persons or 12% of the United States population. Because NARMS 2001 *Campylobacter* surveillance was not nationwide, generalization to the United States population should be done with caution due to potential regional differences in the prevalence of antimicrobial resistance among *Campylobacter*. However, analyses of changes in the prevalence of resistance from 1997 to 2001 are not subject to the above limitation within the population under surveillance.

Summary of Long Term Changes in Antimicrobial Resistance in non-Typhi *Salmonella* Isolates, 1979-2001

Sentinel county studies: 1979-1980, 1984-1985,
1989-1990, and 1994-1995

NARMS: 1996-2001



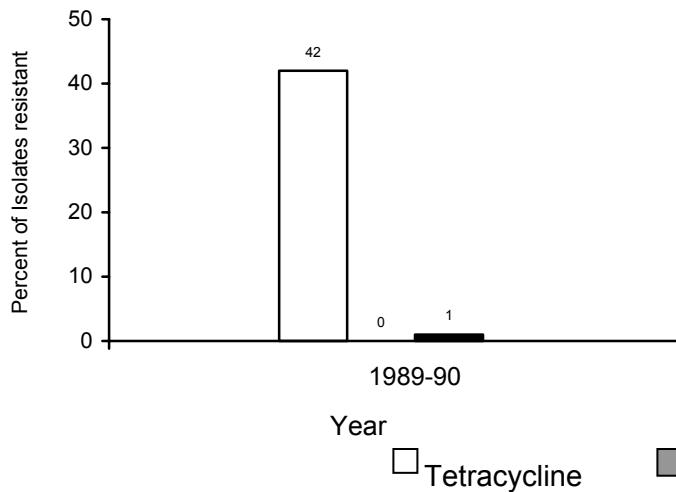
*ACSSuT = resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole, and tetracycline; NT=Not Tested

For non-Typhi *Salmonella*, sentinel county surveys were conducted in 1979-80, 1984-85, 1989-90, and 1994-95 (1,2,3,4). Isolates were tested at CDC by disk diffusion. The National Antimicrobial Resistance Monitoring System (NARMS) for Enteric Bacteria began testing *Salmonella* in 1996. In NARMS, every 10th non-Typhi *Salmonella* isolate received at participating state public health laboratories is forwarded to CDC and tested by broth microdilution to determine partial range MICs to 17 antimicrobial agents.

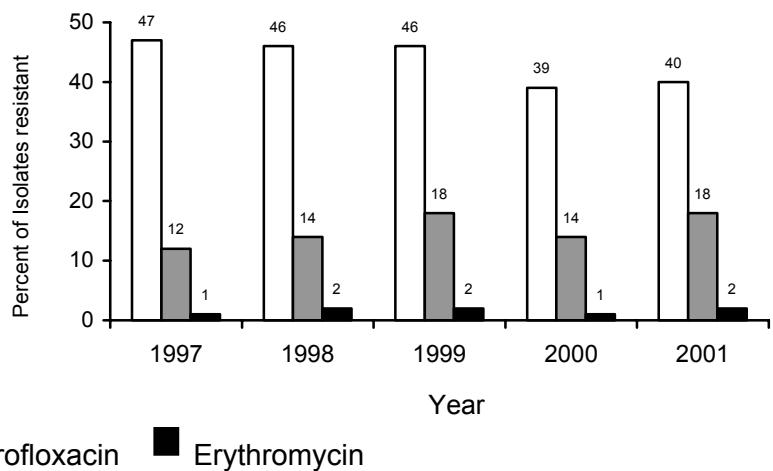
Over the last quarter century, resistance among non-Typhi *Salmonella* has increased to a number of clinically important antimicrobial agents. Resistance to ampicillin and trimethoprim-sulfamethoxazole increased first, reaching 21% and 4%, respectively, in 1996. Resistance to third-generation cephalosporins (e.g., ceftriaxone), quinolones (e.g., nalidixic acid), and the ACSSuT resistance pattern increased more recently. A public health concern raised by this resistance is the loss of efficacious agents to treat serious *Salmonella* infections, especially in children. The clinical implications of current resistance levels are potential treatment failure, increased duration of illness, and increased length of hospitalization (3,5,6). For more information on treatment of *Salmonella* see [Diagnosis and Management of Foodborne Illness: A Primer for Physicians](#) (7).

Summary of Long Term Changes in Antimicrobial Resistance in *Campylobacter jejuni* Isolates, 1989-2001

Sentinel county study: 1989-1990



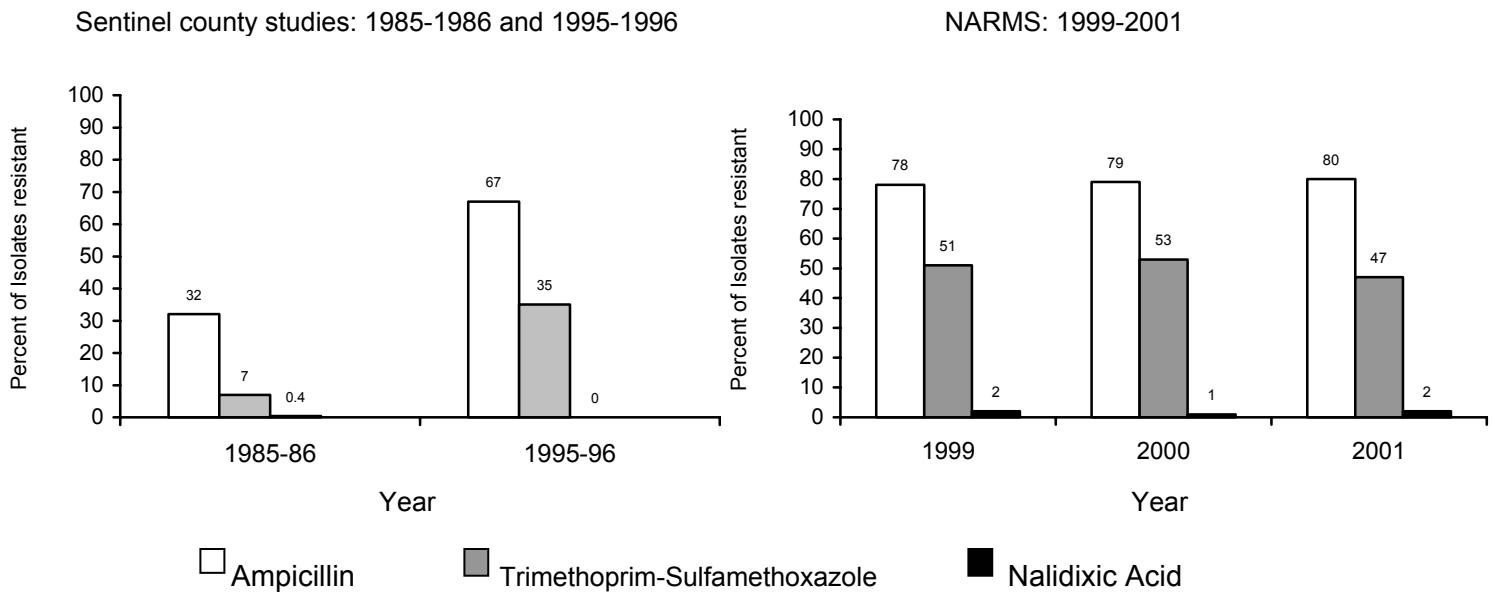
NARMS: 1997-2001



For *Campylobacter jejuni*, a sentinel county survey was conducted in 1989-90 (8). Isolates were received and tested at CDC. NARMS began testing *Campylobacter* in 1997. In NARMS, one *Campylobacter* isolate per week is forwarded to CDC from nine states and tested by E-test for susceptibility to eight antimicrobial agents.

Over the last quarter century, resistance among *Campylobacter jejuni* has increased to a number of clinically important antimicrobial agents. Resistance to tetracycline was already 42% in 1989-90. Resistance to ciprofloxacin increased more recently. No isolates resistant to ciprofloxacin were identified in 1989-90, 12% were resistant in 1997, and 18% in 2001. Resistance to erythromycin has remained low at 2% or less. Because the primary reservoir for *Campylobacter jejuni* is among poultry, it is likely that this increasing ciprofloxacin resistance is related to the use of fluoroquinolones, which were approved for use in poultry farming in 1995. A public health concern raised by this resistance is the threat to the efficacy of fluoroquinolones. The clinical implications of resistance to fluoroquinolones include an increased duration of illness and potential treatment failure (9). For more information on treatment of *Campylobacter* see [Diagnosis and Management of Foodborne Illness: A Primer for Physicians](#) (7).

Summary of Long Term Changes in Antimicrobial Resistance in *Shigella* Isolates, 1985-2001



For *Shigella*, sentinel county surveys were conducted in 1985-86 and 1995-96 (10). Isolates were received and tested at CDC. NARMS began testing *Shigella* in 1999. In NARMS, every 10th *Shigella* isolate received at participating state public health laboratories is forwarded to CDC and tested by broth microdilution to determine partial range MICs to 17 antimicrobial agents.

Over the last 17 years, resistance among *Shigella* has increased to a number of clinically important antimicrobial agents. Resistance to ampicillin was already 32% in 1985-86 and increased to 67% by 1995. Resistance to nalidixic acid emerged more recently. One *Shigella* isolate resistant to nalidixic acid was identified in 1985-86. The percentage of *Shigella* isolates resistant to nalidixic acid increased to 2% in 1999 and remains at 2% in 2001.

As *Shigella* have no environmental or animal reservoir except humans, it is likely that this resistance is related to the use of antimicrobials in human medicine. A public health concern raised by these resistances is the loss of efficacious agents to treat *Shigella* infections. The clinical implication of current resistance levels is potential treatment failure. This may be particularly important for infections related to international travel (10). For more information on treatment of *Shigella* see [Diagnosis and Management of Foodborne Illness: A Primer for Physicians](#) (7).

References

1. Riley L, Cohen M, Seals J, Blaser M, Birkness K, Hargrett N, Martin S, Feldman R. Importance of host factors in human salmonellosis caused by multiresistant strains of *Salmonella*. *Journal of Infectious Diseases* 1984; 149:878-83.
2. MacDonald K, Cohen M, Hargrett-Bean N, Wells J, Puhr N, Collin S, Blake P. Changes in antimicrobial resistance of *Salmonella* isolated from humans in the United States. *JAMA* 1987; 258:1496-9.
3. Lee L, Puhr N, Maloney E, Bean N, Tauxe R. Increase in antimicrobial-resistance *Salmonella* infectious in the United States, 1989-1990. *Journal of Infectious Diseases* 1994; 170:128-34.
4. Herikstad H, Hayes PS, Hogan J, Floyd P, Snyder L, Angulo FJ. Ceftriaxone-resistant *Salmonella* in the United States. *Pediatric Infectious Disease Journal* 1997; 16(9):904-5.
5. Molbak K, Baggesen D, Aarestrup F, Ebbesen J, Engberg J, Frydendahl K, Gerner-Smidt P, Petersen A, Wegener H. An outbreak of multidrug-resistant, quinolone-resistant *Salmonella enterica* serotype Typhimurium DT104. *New England Journal of Medicine* 1999; 341(19):1420-5.
6. Holmberg S, Solomon S, Blake P. Health and economic impacts of antimicrobial resistance. *Reviews of Infectious Diseases* 1987; 9:1065-78.
7. Available from URL: <http://www.cdc.gov/mmwr/preview/mmwrhtml/rr5002a1.htm>
8. Sobel J, Tauxe R, Ries A, Patton C, Maloney K. The Burden of *Campylobacter jejuni* Infections: A Target for Early Treatment? *In Program and Abstracts of the 45th Annual Conference of the Epidemic Intelligence Service, Atlanta, GA, April 1995.*
9. Marano N, Vugia D, Fiorentino T, Segler S, Carter M, Kassenborg H, Smith K, Zansky S, Hollinger K, Angulo F, and the EIP FoodNet Working Group. Fluoroquinolone-resistant *Campylobacter* causes longer duration of diarrhea than fluoroquinolone-susceptible *Campylobacter* strains in FoodNet sites. *In Program and Abstracts of the 2nd International Conference on Emerging Infectious Diseases, Atlanta, GA, July 2000.*
10. Cook K, Boyce T, Puhr N, Tauxe R, Mintz E. Increasing antimicrobial-resistant *Shigella* infections in the United States. *In Abstracts of the 36th Interscience Conference on Antimicrobial Agents and Chemotherapy, New Orleans, LA, September 1996.*
11. National Committee for Clinical Laboratory Standards, 2001. Performance Standards for antimicrobial susceptibility testing. NCCLS approved standard M100-S11. National Committee for Clinical Laboratory Standards, Wayne, PA.

Publications and Abstracts, 2001

Publications

1. McDonald C, Rossiter S, Mackinson C, Wany Y, Johnson S, Sullivan M, Sokolow R, DeBess E, Gilbert L, Benson J, Hill B, Angulo F. Quinupristin-dalfopristin-resistant *Enterococcus faecium* on chicken and in human stool specimens. New England Journal of Medicine 2001; 345(16):1155-60.
2. Olsen S, DeBess E, McGivern T, Marano N, Eby T, Mauvais S, Balan V, Zirnstein G, Cieslak P, Angulo F. A nosocomial outbreak of fluoroquinolone-resistant *Salmonella* infection. New England Journal of Medicine 2001; 344(21):1572-9.

Abstracts

1. Angulo F, Stamey K, Rossiter S, Barrett T, and the NARMS Working Group. Three prevalent multidrug-resistant strains among human *Salmonella* isolates in the United States, 1999-2000: *S. Typhimurium* R-type ACSSuT, *S. Typhimurium* R-type AKSSuT, and *S. Newport* R-type ACSSuT. 41st Interscience Conference on Antimicrobial Agents and Chemotherapy. Chicago, IL, December 2001.
2. Gupta A, Rossiter S, McClellan J, Stamey K, Barrett T, Angulo F, and the NARMS Working Group. Fluoroquinolone-resistant *Campylobacter* infections in the United States, 1997-2000: National Antimicrobial Resistance Monitoring System's data lead to regulatory action. 50th Annual Epidemic Intelligence Service Conference. Atlanta, GA, April 2001.
3. Gupta A, Rossiter S, McClellan J, Stamey K, Barrett T, Angulo F, and the NARMS Working Group. Fluoroquinolone-resistant *Campylobacter jejuni* infections in the United States, 1997-2000: National Antimicrobial Resistance Monitoring System's data lead to regulatory action. 39th Annual Meeting of the Infectious Diseases Society of America. San Francisco, CA, October 2001.
4. Gupta A, Crowe C, Bolstorff B, Fontana J, Stout A, Montgomery S, McGuill M, Matyas B, Johnson B, Schoenfeld S, and Angulo F. Multistate investigation of multidrug-resistant *Salmonella* serotype Newport infections in the Northeastern United States, 2000: human infections associated with dairy farms. 39th Annual Meeting of the Infectious Diseases Society of America. San Francisco, CA, October 2001.

5. McClellan J, Joyce K, Rossiter S, Barrett T, Angulo F, and the NARMS Enterococci Working Group. High-level gentamicin resistant enterococci and quinupristin/dalfopristin-resistant *E. faecium* from ground pork purchased from grocery stores. 41st Interscience Conference on Antimicrobial Agents and Chemotherapy. Chicago, IL, December 2001.
6. Rossiter S, Joyce K, Johnson S, Gregg C, Steiner C, Gilbert L, Franko E, DeBess E, Taylor B, Madden J, Angulo F. High prevalence of bacitracin-resistant *E. faecium* and *E. faecalis* in human stools and on chicken in the United States. 101st General Meeting of the American Society for Microbiology. Orlando, FL, May 2001.
7. Sivapalasingam S, McClellan J, Joyce K, Reddy V, Agasan A, Goldbaum R, Leano F, Barrett T, Angulo F, Mintz E, and the NARMS Working Group. Increasing antimicrobial resistance among *Shigella* isolates in the United States, 1999-2000. 39th Annual Meeting of the Infectious Diseases Society of America. San Francisco, CA, October 2001.
8. Stamey K, Rossiter S, Kubota K, Van Duyne S, Blackmore C, Farah R, Fiorella P, Baker R, Rinehardt C, Barrett T, Angulo F. PFGE patterns of multidrug-resistant *Salmonella* serotype Senftenberg isolated from patients in a Florida hospital. 101st General Meeting of the American Society for Microbiology. Orlando, FL, May 2001.
9. Wright J. Outbreak of multidrug-resistant *Salmonella* Typhimurium infections in humans associated with veterinary facilities. 138th American Veterinary Medical Association Convention. Boston, MA, July 2001.

Part II
Summary Tables and Graphs

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 1. Population size and number of isolates received and tested, by site, 2001

| Site | Population Size* | | Non-Typhi <i>Salmonella</i> | <i>Salmonella</i> Typhi | <i>Shigella</i> | <i>E. coli</i> O157 | <i>Campylobacter</i> ** | |
|-------------------------------|-----------------------------|-----|--------------------------------|----------------------------|------------------|------------------------|-------------------------|------|
| | No. | (%) | | | | | No. | (%) |
| California ⁽¹⁾ | 3,204,675 | (3) | 53 | (4) | 18 | (9) | 20 | (6) |
| Colorado | 4,417,714 | (4) | 63 | (4) | 0 | (0) | 23 | (7) |
| Connecticut | 3,425,074 | (3) | 50 | (3) | 5 | (2) | 6 | (2) |
| Florida | 16,396,515 (15) | | 55 | (4) | 4 | (2) | 1 | (0) |
| Georgia | 8,383,915 | (8) | 185 | (13) | 8 | (4) | 41 | (12) |
| Kansas | 2,694,641 | (2) | 28 | (2) | 0 | (0) | 5 | (1) |
| Los Angeles ⁽²⁾ | 9,637,494 | (9) | 102 | (7) | 19 | (10) | 17 | (5) |
| Maryland | 5,375,156 | (5) | 86 | (6) | 12 | (6) | 14 | (4) |
| Massachusetts | 6,379,304 | (6) | 131 | (9) | 10 | (5) | 19 | (5) |
| Minnesota | 4,972,294 | (5) | 63 | (4) | 6 | (3) | 45 | (13) |
| New Jersey | 8,484,431 | (8) | 118 | (8) | 40 | (20) | 35 | (10) |
| New York City ⁽³⁾ | 8,019,033 | (7) | 139 | (10) | 47 | (24) | 45 | (13) |
| New York State ⁽⁴⁾ | 10,992,345 (10) | | 135 | (9) | 11 | (6) | 23 | (7) |
| Oregon | 3,472,867 | (3) | 26 | (2) | 9 | (5) | 10 | (3) |
| Tennessee | 5,740,021 | (5) | 84 | (6) | 2 | (1) | 12 | (3) |
| Washington | 5,987,973 | (5) | 76 | (5) | 6 | (3) | 23 | (7) |
| West Virginia | 1,801,916 | (2) | 25 | (2) | 0 | (0) | 5 | (1) |
| Totals | 109,385,368 (100) | | 1419 (100) | 197 (100) | 344 (100) | 277 (100) | 384 (100) | |

* County population 2001, U.S. Census Bureau, post-census estimates

** *Campylobacter* isolates are submitted only from FoodNet sites, population size of FoodNet sites is 34.3 million persons (see <http://www.cdc.gov/foodnet/>)

(1) Alameda, Contra Costa, and San Francisco counties

(2) Los Angeles County

(3) Five boroughs of New York City (Bronx, Brooklyn, Manhattan, Queens, Staten Island)

(4) Excluding New York City

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 2. Antimicrobial agents used for susceptibility testing for *Salmonella*, *Shigella*, *E. coli* O157, and *Campylobacter* isolates, 2001

| Antimicrobial Agent | Antimicrobial Agent Concentration Range ($\mu\text{g}/\text{ml}$) | Breakpoints [R] | [I] | [S] |
|---------------------------------|--|--------------------|---------|-------------|
| Amikacin | 4 - 32 | ≥ 64 | 32 | ≤ 16 |
| Amoxicillin – Clavulanic Acid | 0.5/0.25 – 32/16 | $\geq 32/16$ | 16/8 | $\leq 8/4$ |
| Ampicillin | 2 – 32 | ≥ 32 | 16 | ≤ 8 |
| Apramycin** | 2 – 32 | ≥ 64 | 16 – 32 | ≤ 8 |
| Azithromycin | 0.016 – 256* | ≥ 2 | 0.5-1 | ≤ 0.25 |
| Cefoxitin | 4-32 | ≥ 32 | 16 | ≤ 8 |
| Ceftiofur** | 0.5 – 16 | ≥ 8 | 4 | ≤ 2 |
| Ceftriaxone | 0.25 – 64 | ≥ 64 | 16 - 32 | ≤ 8 |
| Cephalothin | 1 – 32 | ≥ 32 | 16 | ≤ 8 |
| Chloramphenicol | 4 – 32 0.016 – 256* | ≥ 32 | 16 | ≤ 8 |
| Ciprofloxacin | 0.015 – 4 0.002 – 32* | ≥ 4 | 2 | ≤ 1 |
| Clindamycin | 0.016 – 256* | ≥ 4 | 1-2 | ≤ 0.5 |
| Erythromycin | 0.016 – 256* | ≥ 8 | 1-4 | ≤ 0.5 |
| Gentamicin | 0.25 – 16 0.016 – 256* | ≥ 16 | 8 | ≤ 4 |
| Imipenem | 0.25 – 8 | ≥ 16 | 8 | ≤ 4 |
| Kanamycin | 16 – 64 | ≥ 64 | 32 | ≤ 16 |
| Nalidixic Acid | 4 –64 0.016 – 256* | ≥ 32 | | ≤ 16 |
| Streptomycin | 32 – 64 | ≥ 64 | | ≤ 32 |
| Sulfamethoxazole | 128 – 512 | ≥ 512 | | ≤ 256 |
| Tetracycline | 8 – 16 0.016 – 256* | ≥ 16 | 8 | ≤ 4 |
| Trimethoprim - Sulfamethoxazole | 0.12/2.4 – 4/76 | $\geq 4/76$ | | $\leq 2/38$ |

* *Campylobacter* antimicrobial agents and concentration ranges used

** No NCCLS interpretive standards for this antimicrobial agent

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 3. Results of ceftriaxone confirmatory testing for non-Typhi *Salmonella* isolates meeting screening criteria*, 2001

| Isolate | Sensititre® MIC | | | | By-hand broth microdilution MIC | |
|---------|-----------------|---------|-------------|----------|---------------------------------|----------|
| | Ceftiofur | | Ceftriaxone | | Ceftriaxone | |
| | µg/ml | S/I/R** | µg/ml | S/I/R*** | µg/ml | S/I/R*** |
| 1 | >16 | R | 32 | I | 64 | R |
| 2 | 16 | R | 4 | S | 64 | R |
| 3 | >16 | R | 16 | I | 64 | R |
| 4 | >16 | R | 8 | S | 64 | R |
| 5 | >16 | R | 32 | I | 64 | R |
| 6 | >16 | R | 32 | I | 64 | R |
| 7 | >16 | R | 32 | I | 64 | R |
| 8 | >16 | R | 16 | I | 64 | R |
| 9 | 16 | R | 8 | S | 64 | R |
| 10 | 16 | R | 8 | S | 64 | R |
| 11 | >16 | R | 16 | I | 64 | R |
| 12 | >16 | R | 16 | I | 64 | R |
| 13 | >16 | R | 16 | I | 64 | R |
| 14 | >16 | R | 8 | S | 64 | R |
| 15 | >16 | R | 16 | I | 64 | R |
| 16 | >16 | R | 32 | I | 64 | R |
| 17 | >16 | R | 32 | I | >64 | R |
| 18 | >16 | R | 16 | I | 64 | R |
| 19 | >16 | R | 16 | I | >64 | R |
| 20 | >16 | R | 16 | I | 64 | R |
| 21 | >16 | R | 16 | I | 64 | R |
| 22 | >16 | R | 16 | I | 64 | R |
| 23 | >16 | R | 16 | I | 64 | R |
| 24 | >16 | R | 32 | I | >64 | R |
| 25 | >16 | R | 32 | I | >64 | R |
| 26 | >16 | R | 16 | I | 64 | R |
| 27 | 16 | R | 16 | I | 64 | R |
| 28 | >16 | R | 16 | I | 64 | R |
| 29 | >16 | R | 32 | I | >64 | R |
| 30 | >16 | R | 8 | S | 64 | R |
| 31 | >16 | R | 16 | I | 64 | R |
| 32 | >16 | R | 16 | I | 64 | R |
| 33 | >16 | R | 16 | I | 64 | R |
| 34 | >16 | R | 16 | I | 64 | R |

*Decreased susceptibility to ceftiofur [MIC \geq 4 µg/ml] and/or ceftriaxone [MIC \geq 2 µg/ml], by Sensititre®

**Ceftiofur: S = Susceptible (MIC \leq 2 µg/ml); I = Intermediate (MIC = 4 µg/ml); R = Resistant (MIC \geq 8 µg/ml)

***Ceftriaxone: S = Susceptible (MIC \leq 8 µg/ml); I = Intermediate (MIC 16-32 µg/ml); R = Resistant (MIC \geq 64 µg/ml)

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 3. Results of ceftriaxone confirmatory testing for non-Typhi *Salmonella* isolates meeting screening criteria*, 2001

| Isolate | Sensititre® MIC | | | | By-hand broth microdilution MIC | |
|---------|-----------------|---------|-------------|----------|---------------------------------|----------|
| | Ceftiofur | | Ceftriaxone | | Ceftriaxone | |
| | µg/ml | S/I/R** | µg/ml | S/I/R*** | µg/ml | S/I/R*** |
| 35 | 16 | R | 8 | S | 16 | I |
| 36 | 16 | R | 16 | I | 32 | I |
| 37 | 8 | R | 4 | S | 16 | I |
| 38 | 16 | R | 16 | I | 32 | I |
| 39 | >16 | R | 16 | I | 32 | I |
| 40 | 16 | R | 8 | S | 32 | I |
| 41 | 16 | R | 8 | S | 32 | I |
| 42 | 16 | R | 8 | S | 32 | I |
| 43 | 16 | R | 8 | S | 32 | I |
| 44 | 8 | R | 8 | S | 16 | I |
| 45 | 16 | R | 16 | I | 32 | I |
| 46 | >16 | R | 16 | I | 32 | I |
| 47 | 16 | R | 8 | S | 32 | I |
| 48 | 16 | R | 8 | S | 32 | I |
| 49 | >16 | R | 2 | S | <=0.125 | S |
| 50 | 4 | S | <=0.250 | S | 0.5 | S |
| 51 | >16 | R | 8 | S | <=0.125 | S |
| 52 | >16 | R | 2 | S | <=0.125 | S |
| 53 | >16 | R | 1 | S | <=0.125 | S |
| 54 | >16 | R | 0.500 | S | <=0.125 | S |
| 55 | >16 | R | 4 | S | <=0.125 | S |
| 56 | >16 | R | 2 | S | <=0.125 | S |
| 57 | >16 | R | 2 | S | <=0.125 | S |
| 58 | >16 | R | 4 | S | <=0.125 | S |
| 59 | >16 | R | 4 | S | <=0.125 | S |
| 60 | 4 | I | <=0.250 | S | <=0.125 | S |

*Decreased susceptibility to ceftiofur [MIC \geq 4 µg/ml] and/or ceftriaxone [MIC \geq 2 µg/ml], by Sensititre®

**Ceftiofur: S = Susceptible (MIC \leq 2 µg/ml); I = Intermediate (MIC = 4 µg/ml); R = Resistant (MIC \geq 8 µg/ml)

***Ceftriaxone: S = Susceptible (MIC \leq 8 µg/ml); I = Intermediate (MIC 16-32 µg/ml); R = Resistant (MIC \geq 64 µg/ml)

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 4. Antimicrobial resistance of *Salmonella*, *Shigella*, and *E. coli* O157 isolates, 2001

| Antimicrobial Agent | Non-Typhi <i>Salmonella</i> (N=1419) | | <i>Salmonella</i> Typhi (N=197) | | <i>Shigella</i> (N=344) | | <i>E. coli</i> O157 (N=277) | |
|------------------------------------|--|-------|------------------------------------|-------|----------------------------|-------|--------------------------------|-------|
| | N | % | N | % | N | % | N | % |
| Amikacin | 0 | (0) | 0 | (0) | 0 | (0) | 0 | (0) |
| Amoxicillin – Clavulanic Acid | 66 | (5) | 0 | (0) | 15 | (4) | 2 | (1) |
| Ampicillin | 247 | (17) | 40 | (20) | 274 | (80) | 6 | (2) |
| Apramycin | 0 | (0) | 0 | (0) | 0 | (0) | 0 | (0) |
| Cefoxitin | 48 | (3) | 1 | (0.5) | 4 | (1) | 2 | (1) |
| Ceftiofur | 58 | (4) | 0 | (0) | 0 | (0) | 3 | (1) |
| Ceftriaxone | 34 | (2) | 0 | (0) | 0 | (0) | 0 | (0) |
| Cephalothin | 57 | (4) | 1 | (0.5) | 31 | (9) | 4 | (1) |
| Chloramphenicol | 164 | (12) | 41 | (21) | 74 | (21) | 4 | (1) |
| Ciprofloxacin | 3 | (0.2) | 0 | (0) | 1 | (0.3) | 0 | (0) |
| Gentamicin | 27 | (2) | 0 | (0) | 0 | (0) | 1 | (0.4) |
| Imipenem | 0 | (0) | 0 | (0) | 0 | (0) | 0 | (0) |
| Kanamycin | 68 | (5) | 1 | (0.5) | 2 | (1) | 0 | (0) |
| Nalidixic Acid | 37 | (3) | 59 | (30) | 6 | (2) | 3 | (1) |
| Streptomycin | 241 | (17) | 40 | (20) | 183 | (53) | 5 | (2) |
| Sulfamethoxazole | 251 | (18) | 41 | (21) | 194 | (56) | 14 | (5) |
| Tetracycline | 280 | (20) | 41 | (21) | 204 | (59) | 15 | (5) |
| Trimethoprim - Sulfamethoxazole | 28 | (2) | 41 | (21) | 161 | (47) | 2 | (1) |

National Antimicrobial Resistance Monitoring System For Enteric Bacteria
Table 5a. Additional resistance by antimicrobial agent for non-Typhi *Salmonella*, 2001

| Resistance to: N (%) | Amika | Amo-Cl | Ampic | Afram | Cefox | Cefti | Ceftr | Cepha | Chlor | Cipro | Genta | Imipe | Kanam | Naldx | Strep | Sulfa | Tetra | Tri-Su |
|--------------------------------------|-----------|------------|------------|-----------|------------|------------|-----------|------------|------------|-----------|-----------|-----------|-----------|-----------|------------|------------|------------|-----------|
| Amikacin [Amika] 0 (0) | | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | |
| Amox-Clav [Amo-Cl] 66 (5) | 0 0% | | 65 98% | 0 0% | 48 73% | 52 79% | 34 51% | 50 76% | 47 71% | 2 3% | 4 6% | 0 0% | 11 17% | 3 4% | 50 76% | 53 80% | 47 71% | 3 4% |
| Ampicillin [Ampic] 247 (17) | 0 0% | 65 26% | | 0 0% | 48 19% | 53 21% | 34 14 | 55 22% | 157 64% | 3 1% | 9 4% | 0 0% | 43 17% | 8 3% | 176 71% | 193 78% | 191 77% | 14 6% |
| Apramycin [Afram] 0 (0) | 0 100% | 0 100% | 0 100% | | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | |
| Cefoxitin [Cefox] 48 (3) | 0 0% | 48 100% | 48 100% | 0 0% | | 47 98% | 34 71% | 48 100% | 36 75% | 0 0% | 3 6% | 0 0% | 9 19% | 1 2% | 39 81% | 38 79% | 37 77% | 1 2% |
| Ceftiofur [Cefti] 58 (4) | 0 0% | 52 90% | 53 91% | 0 0% | 47 81% | | 34 59% | 48 83% | 36 62% | 1 2% | 4 7% | 0 0% | 10 17% | 2 3% | 38 65% | 41 71% | 37 64% | 1 2% |
| Ceftriaxone [Ceftr] 34 (2) | 0 0% | 34 100% | 34 100% | 0 0% | 34 100% | 34 100% | | 34 100% | 30 88% | 0 0% | 2 6% | 0 0% | 7 21% | 0 0% | 32 94% | 31 91% | 31 91% | 1 3% |
| Cephalothin [Cepha] 57 (4) | 0 0% | 50 88% | 55 96% | 0 0% | 48 84% | 48 84% | 34 60% | | 38 67% | 2 3% | 5 9% | 0 0% | 14 25% | 4 7% | 43 75% | 42 74% | 38 67% | 3 5% |
| Chloramphenicol [Chlor] 164 (12) | 0 0% | 47 29% | 157 96% | 0 0% | 36 22% | 36 22% | 30 18% | 38 23% | | 2 1% | 6 4% | 0 0% | 17 10% | 2 1% | 147 90% | 162 99% | 162 99% | 11 7% |
| Ciprofloxacin [Cipro] 3 (0.2) | 0 0% | 2 67% | 3 100% | 0 0% | 0 0% | 1 33% | 0 0% | 2 67% | 2 67% | | 2 67% | 0 0% | 2 67% | 3 100% | 1 33% | 2 67% | 1 33% | 1 33% |
| Gentamicin [Genta] 27 (2) | 0 0% | 4 15% | 9 33% | 0 0% | 3 11% | 4 15% | 2 7% | 5 18% | 6 22% | 2 7% | | 0 0% | 7 26% | 2 7% | 23 85% | 26 96% | 13 48% | 4 15% |
| Imipenem [Imipe] 0 (0) | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | |
| Kanamycin [Kanam] 68 (5) | 0 0% | 11 16% | 43 63% | 0 0% | 9 13% | 10 15% | 7 10% | 14 21% | 17 25% | 2 3% | 7 10% | 0 0% | | 4 6% | 56 82% | 43 63% | 58 85% | 5 7% |
| Nalidixic Acid [Naldx] 37 (3) | 0 0% | 3 8% | 8 22% | 0 0% | 1 3% | 2 5% | 0 0% | 4 11% | 2 5% | 3 8% | 2 5% | 0 0% | 4 11% | | 3 8% | 5 13% | 6 16% | 4 11% |
| Streptomycin [Strep] 241 (17) | 0 0% | 50 21% | 176 73% | 0 0% | 39 16% | 38 16% | 32 13% | 43 18% | 147 61% | 1 0.4% | 23 9% | 0 0% | 56 23% | 3 1% | | 202 84% | 212 88% | 13 5% |
| Sulfamethoxazole [Sulfa] 251 (18) | 0 0% | 53 21% | 193 77% | 0 0% | 38 15% | 41 16% | 31 12% | 42 17% | 162 64% | 2 1% | 26 10% | 0 0% | 43 17% | 5 2% | 202 80% | | 210 84% | 27 11% |
| Tetracycline [Tetra] 280 (20) | 0 0% | 47 17% | 191 68% | 0 0% | 37 13% | 37 13% | 31 11% | 38 14% | 162 58% | 1 0.4% | 13 5% | 0 0% | 58 21% | 6 2% | 212 76% | 210 75% | | 19 7% |
| Trimeth-Sulfa [Tri-Su] 28 (2) | 0 0% | 3 11% | 14 50% | 0 0% | 1 4% | 1 4% | 1 4% | 3 11% | 11 39% | 1 4% | 4 14% | 0 0% | 5 18% | 4 14% | 13 14% | 27 46% | 19 68% | |

National Antimicrobial Resistance Monitoring System For Enteric Bacteria
Table 5b. Additional resistance by antimicrobial agent for *Salmonella* Typhi, 2001

| Resistance to: N (%) | Amika | AmoCl | Ampic | Afram | Cefox | Cefti | Ceftr | Cepha | Chlor | Cipro | Genta | Imipe | Kanam | Naldx | Strep | Sulfa | Tetra | Tri-Su |
|-------------------------------------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|------------|-----------|-----------|
| Amikacin [Amika] 0 (0) | | 0 100% | 0 100% | |
| Amox-Clav [Amo-Cl] 0 (0) | 0 100% | | 0 100% | 0 100% | |
| Ampicillin [Ampic] 40 (20) | 0 0% | 0 0% | | 0 0% | 0 0% | 0 0% | 0 0% | 38 95% | 0 0% | 0 0% | 0 0% | 1 2% | 23 57% | 38 95% | 39 97% | 36 90% | 36 90% | |
| Apramycin [Afram] 0 (0) | 0 100% | 0 100% | 0 100% | | 0 100% | 0 100% | |
| Cefoxitin [Cefox] 1 (0.5) | 0 0% | 0 0% | 0 0% | 0 0% | | 0 0% | 1 100% | 0 0% | 0 0% | |
| Ceftiofur [Cefti] 0 (0) | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | | 0 100% | 0 100% | |
| Ceftriaxone [Ceftr] 0 (0) | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | | 0 100% | 0 100% | |
| Cephalothin [Cepha] 1 (0.5) | 0 0% | 0 0% | 0 0% | 0 0% | 1 100% | 0 0% | | 0 0% | 0 0% | |
| Chloramphenicol [Chlor] 41 (21) | 0 0% | 0 0% | 38 93% | 0 0% | 0 0% | 0 0% | 0 0% | | 0 0% | 0 0% | 0 0% | 0 0% | 26 63% | 36 88% | 37 90% | 38 93% | 38 93% | |
| Ciprofloxacin [Cipro] 0 (0) | 0 100% | | 0 100% | 0 100% | |
| Gentamicin [Genta] 0 (0) | 0 100% | | 0 100% | 0 100% | |
| Imipenem [Imipe] 0 (0) | 0 100% | | 0 100% | 0 100% | |
| Kanamycin [Kanam] 1 (0.5) | 0 0% | 0 0% | 1 100% | 0 0% | 0 0% | 0 0% | 0 0% | 0 0% | | 0 0% | 0 0% | 0 0% | | 0 0% | 1 100% | 1 100% | 1 100% | |
| Nalidixic Acid [Naldx] 59 (30) | 0 0% | 0 0% | 23 39% | 0 0% | 0 0% | 0 0% | 0 0% | 0 0% | 26 44% | 0 0% | 0 0% | 0 0% | | | 22 37% | 22 37% | 26 44% | 24 41% |
| Streptomycin [Strep] 40 (20) | 0 0% | 0 0% | 38 95% | 0 0% | 0 0% | 0 0% | 0 0% | 0 0% | 36 90% | 0 0% | 0 0% | 0 0% | 1 2% | 22 55% | | 40 100% | 34 85% | 37 92% |
| Sulfamethoxazole [Sulfa] 41 (21) | 0 0% | 0 0% | 39 95% | 0 0% | 0 0% | 0 0% | 0 0% | 0 0% | 37 90% | 0 0% | 0 0% | 0 0% | 1 2% | 22 54% | 40 98% | | 35 85% | 38 93% |
| Tetracycline [Tetra] 41 (21) | 0 0% | 0 0% | 36 88% | 0 0% | 0 0% | 0 0% | 0 0% | 0 0% | 38 93% | 0 0% | 0 0% | 0 0% | 1 2% | 26 63% | 34 83% | 35 85% | | 35 85% |
| Trimeth-Sulfa [Tri-Su] 41 (21) | 0 0% | 0 0% | 36 88% | 0 0% | 0 0% | 0 0% | 0 0% | 0 0% | 38 93% | 0 0% | 0 0% | 0 0% | 0 0% | 24 58% | 37 90% | 38 93% | 35 85% | |

National Antimicrobial Resistance Monitoring System For Enteric Bacteria
Table 5c. Additional resistance by antimicrobial agent for *Shigella*, 2001

| Resistance to: N (%) | Amika | AmoCl | Ampic | Apram | Cefox | Cefti | Ceftr | Cepha | Chlor | Cipro | Genta | Imipe | Kanam | Naldx | Strep | Sulfa | Tetra | Tri-Su |
|---|-----------|-----------|------------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|------------|------------|------------|------------|
| Amikacin [Amika] 0 (0) | | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | |
| Amox-Clav [Amo-Cl] 15 (4) | 0 0% | | 15 100% | 0 0% | 4 27% | 0 0% | 0 0% | 10 67% | 4 27% | 0 0% | 0 0% | 0 0% | 0 0% | 0 0% | 8 53% | 10 67% | 4 27% | 9 60% |
| Ampicillin [Ampic] 274 (80) | 0 0% | 15 5% | | 0 0% | 4 1% | 0 0% | 0 0% | 31 11% | 69 25% | 1 0.4% | 0 0% | 0 0% | 2 1% | 3 1% | 153 56% | 152 55% | 161 59% | 129 47% |
| Apramycin [Apram] 0 (0) | 0 100% | 0 100% | 0 100% | | 0 100% | 0 100% | 0 100% | |
| Cefoxitin [Cefox] 4 (1) | 0 0% | 4 100% | 4 100% | 0 0% | | 0 0% | 0 0% | 4 100% | 0 0% | 0 0% | 0 0% | 0 0% | 0 0% | 0 0% | 3 75% | 3 75% | 0 0% | 3 75% |
| Ceftiofur [Cefti] 0 (0) | 0 0% | 0 0% | 0 0% | 0 0% | 0 0% | | 0 0% | 0 0% | 0 0% | |
| Ceftriaxone [Ceftr] 0 (0) | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | | 0 100% | 0 100% | 0 100% | |
| Cephalothin [Cepha] 31 (9) | 0 0% | 10 32% | 31 100% | 0 0% | 4 13% | 0 0% | 0 0% | | 1 3% | 0 0% | 0 0% | 0 0% | 0 0% | 0 0% | 24 77% | 24 77% | 17 55% | 24 77% |
| Chloramphenicol [Chlor] 74 (21) | 0 0% | 4 5% | 69 93% | 0 0% | 0 0% | 0 0% | 0 0% | 1 1% | | 1 1% | 0 0% | 0 0% | 0 0% | 2 3% | 41 55% | 45 61% | 72 97% | 27 36% |
| Ciprofloxacin [Cipro] 1 (0.3) | 0 0% | 0 0% | 1 100% | 0 0% | 0 0% | 0 0% | 0 0% | 1 100% | | 0 0% | 0 0% | 0 0% | 1 100% | 1 100% | 1 100% | 1 100% | 1 100% | 1 100% |
| Gentamicin [Genta] 0 (0) | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | |
| Imipenem [Imipe] 0 (0) | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | |
| Kanamycin [Kanam] 2 (1) | 0 0% | 0 0% | 2 100% | 0 0% | | 0 0% | 2 100% | 2 100% | 2 100% | 1 50% | |
| Nalidixic Acid [Naldx] 6 (2) | 0 0% | 0 0% | 3 50% | 0 0% | 0 0% | 0 0% | 0 0% | 0 0% | 2 33% | 1 17% | 0 0% | 0 0% | 0 0% | | 5 83% | 5 83% | 6 100% | 5 83% |
| Streptomycin [Strep] 183 (53) | 0 0% | 8 4% | 153 84% | 0 0% | 3 2% | 0 0% | 0 0% | 24 13% | 41 22% | 1 0.5% | 0 0% | 0 0% | 2 1% | 5 3% | | 156 85% | 148 81% | 146 80% |
| Sulfamethoxazole [Sulfa] 194 (56) | 0 0% | 10 5% | 152 78% | 0 0% | 3 1% | 0 0% | 0 0% | 24 12% | 45 23% | 1 0.5% | 0 0% | 0 0% | 2 1% | 5 3% | 156 80% | | 166 86% | 158 81% |
| Tetracycline [Tetra] 204 (59) | 0 0% | 4 2% | 161 79% | 0 0% | 0 0% | 0 0% | 0 0% | 17 8% | 72 35% | 1 0.5% | 0 0% | 0 0% | 2 1% | 6 3% | 148 72% | 166 81% | | 134 66% |
| Trimeth-Sulfa [Tri-Su] 161 (47) | 0 0% | 9 6% | 129 80% | 0 0% | 3 2% | 0 0% | 0 0% | 24 15% | 27 17% | 1 1% | 0 0% | 0 0% | 1 1% | 5 3% | 146 91% | 158 98% | 134 83% | |

National Antimicrobial Resistance Monitoring System For Enteric Bacteria
Table 5d. Additional resistance by antimicrobial agent for *E. coli* O157, 2001

| Resistance to: N (%) | Amika | AmoCl | Ampic | Afram | Cefox | Cefti | Ceftr | Cepha | Chlor | Cipro | Genta | Imipe | Kanam | Nalidix | Strep | Sulfa | Tetra | Tri-Su |
|--|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|--------|
| Amikacin [Amika] 0 (0) | | 0 100% | |
| Amox-Clav [Amo-Cl] 2 (1) | 0 0% | | 2 100% | 0 0% | 2 100% | 0 0% | 2 100% | 1 50% | 0 0% | 0 0% | 0 0% | 0 0% | 0 0% | 1 50% | 1 50% | 1 50% | 0 0% | |
| Ampicillin [Ampic] 6 (2) | 0 0% | 2 33% | | 0 0% | 2 33% | 0 0% | 3 50% | 1 17% | 0 0% | 0 0% | 0 0% | 0 0% | 0 0% | 2 33% | 2 33% | 2 33% | 1 17% | |
| Aframycin [Afram] 0 (0) | 0 100% | 0 100% | 0 100% | | 0 100% | |
| Cefoxitin [Cefox] 2 (1) | 0 0% | 2 100% | 2 100% | 0 0% | | 2 100% | 0 0% | 2 100% | 1 50% | 0 0% | 0 0% | 0 0% | 0 0% | 1 50% | 1 50% | 1 50% | 0 0% | |
| Ceftiofur [Cefti] 3 (1) | 0 0% | 2 67% | 2 67% | 0 0% | 2 67% | | 0 0% | 2 67% | 1 33% | 0 0% | 0 0% | 0 0% | 0 0% | 1 33% | 1 33% | 1 33% | 0 0% | |
| Ceftriaxone [Ceftr] 0 (0) | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | | 0 100% | |
| Cephalothin [Cepha] 4 (1) | 0 0% | 2 50% | 3 75% | 0 0% | 2 50% | 2 50% | 0 0% | | 2 50% | 0 0% | 0 0% | 0 0% | 0 0% | 3 75% | 3 75% | 3 75% | 1 25% | |
| Chloramphenicol [Chlor] 4 (1) | 0 0% | 1 25% | 1 25% | 0 0% | 1 25% | 1 25% | 0 0% | 2 50% | | 0 0% | 0 0% | 0 0% | 0 0% | 3 75% | 4 100% | 4 100% | 1 25% | |
| Ciprofloxacin [Cipro] 0 (0) | 0 100% | | 0 100% | |
| Gentamicin [Genta] 1 (0.4) | 0 0% | | 0 0% | 0 0% | 0 0% | 0 0% | 1 100% | 1 100% | 0 0% | 0 0% | |
| Imipenem [Imipe] 0 (0) | 0 100% | | 0 100% | |
| Kanamycin [Kanam] 0 (0) | 0 100% | | 0 100% | |
| Nalidixic Acid [Nalidix] 3 (1) | 0 0% | | 0 0% | |
| Streptomycin [Strep] 5 (2) | 0 0% | 1 20% | 2 40% | 0 0% | 1 20% | 1 20% | 0 0% | 3 60% | 3 60% | 0 0% | 1 20% | 0 0% | 0 0% | | 5 100% | 4 80% | 1 20% | |
| Sulfamethoxazole [Sulfa] 14 (5) | 0 0% | 1 7% | 2 14% | 0 0% | 1 7% | 1 7% | 0 0% | 3 21% | 4 29% | 0 0% | 1 7% | 0 0% | 0 0% | 5 36% | | 13 93% | 2 14% | |
| Tetracycline [Tetra] 15 (5) | 0 0% | 1 7% | 2 13% | 0 0% | 1 7% | 1 7% | 0 0% | 3 20% | 4 27% | 0 0% | 0 0% | 0 0% | 0 0% | 4 27% | 13 87% | | 2 13% | |
| Trimeth-Sulfa [Tri-Su] 2 (1) | 0 0% | 0 0% | 1 50% | 0 0% | 0 0% | 0 0% | 0 0% | 1 50% | 1 50% | 0 0% | 0 0% | 0 0% | 0 0% | 1 50% | 2 100% | 2 100% | | |

National Antimicrobial Resistance Monitoring System For Enteric Bacteria
Table 5e. Additional resistance by antimicrobial agent for *Campylobacter*, 2001

| Resistance to: N (%) | Azith | Chlor | Cipro | Clind | Eryth | Genta | Naldx | Tetra |
|-----------------------------------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|
| Azithromycin [Azith] 8 (2) | | 0 0% | 3 37% | 8 100% | 8 100% | 0 0% | 3 37% | 3 37% |
| Chloramphenicol [Chlor] 0 (0) | 0 100% | | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% |
| Ciprofloxacin [Cipro] 75 (19) | 3 4% | 0 0% | | 3 4% | 3 4% | 0 0% | 73 97% | 42 56% |
| Clindamycin [Clind] 8 (2) | 8 100% | 0 0% | 3 37% | | 8 100% | 0 0% | 3 37% | 3 37% |
| Erythromycin [Eryth] 8 (2) | 8 100% | 0 0% | 3 37% | 8 100% | | 0 0% | 3 37% | 3 37% |
| Gentamicin [Genta] 0 (0) | 0 100% | 0 100% | 0 100% | 0 100% | 0 100% | | 0 100% | 0 100% |
| Nalidixic Acid [Naldx] 77 (20) | 3 4% | 0 0% | 73 95% | 3 4% | 3 4% | 0 0% | | 43 56% |
| Tetracycline [Tetra] 156 (41) | 3 2% | 0 0% | 42 27% | 3 2% | 3 2% | 0 0% | 43 28% | |

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 6. The 16 most common non-Typhi *Salmonella* serotypes, 2001

| Serotype | N | % |
|-----------------|-------------|------------|
| Typhimurium | 325 | 23 |
| Enteritidis | 282 | 20 |
| Newport | 124 | 9 |
| Heidelberg | 100 | 7 |
| Javiana | 51 | 4 |
| Oranienburg | 33 | 2 |
| Montevideo | 29 | 2 |
| Muenchen | 29 | 2 |
| Thompson | 27 | 2 |
| Saint Paul | 21 | 1 |
| Berta | 18 | 1 |
| Braenderup | 18 | 1 |
| Agona | 17 | 1 |
| Infantis | 17 | 1 |
| Java | 17 | 1 |
| Stanley | 17 | 1 |
| Not serotyped | 20 | 1 |
| Other serotypes | 274 | 19 |
| Total | 1419 | 100 |

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 7. Frequency of pansusceptibility*, and resistance to one or more antimicrobial agents, among the 16 most common non-Typhi *Salmonella* serotypes, 2001

| Serotype | Total Isolates | | Pansusceptible Isolates | | Resistant Isolates | |
|-------------|----------------|------------------------|-------------------------|---------------|--------------------|---------------|
| | Number | % of <i>Salmonella</i> | Number | % of Serotype | Number | % of Serotype |
| Javiana | 51 | 4 | 51 | 100 | 0 | 0 |
| Oranienburg | 33 | 2 | 32 | 97 | 1 | 3 |
| Thompson | 27 | 2 | 26 | 96 | 1 | 4 |
| Infantis | 17 | 1 | 16 | 94 | 1 | 6 |
| Montevideo | 29 | 2 | 26 | 90 | 3 | 10 |
| Muenchen | 29 | 2 | 25 | 86 | 4 | 14 |
| Enteritidis | 282 | 20 | 242 | 86 | 40 | 14 |
| Java | 17 | 1 | 14 | 82 | 3 | 18 |
| Saint Paul | 21 | 1 | 17 | 81 | 4 | 19 |
| Braenderup | 18 | 1 | 14 | 78 | 4 | 22 |
| Newport | 124 | 9 | 81 | 65 | 43 | 35 |
| Heidelberg | 100 | 7 | 64 | 64 | 36 | 36 |
| Berta | 18 | 1 | 10 | 56 | 8 | 44 |
| Stanley | 17 | 1 | 9 | 53 | 8 | 47 |
| Typhimurium | 325 | 23 | 160 | 49 | 165 | 51 |
| Agona | 17 | 1 | 8 | 47 | 9 | 53 |

*Pansusceptible to 18 antimicrobial agents tested in 2001: amikacin, amoxicillin-clavulanic acid, ampicillin, apramycin, cefoxitin, ceftiofur, ceftriaxone, cephalothin, chloramphenicol, ciprofloxacin, gentamicin, imipenem, kanamycin, nalidixic acid, streptomycin, sulfamethoxazole, tetracycline, trimethoprim-sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 8. Frequency of resistance and multidrug* resistance among the 16 most common non-Typhi *Salmonella* serotypes, 2001

| Serotype | Total | | Number resistant to ≥ 1 antimicrobial agents | | Number resistant to ≥ 2 antimicrobial agents | | Number resistant to ≥ 5 antimicrobial agents | | Number resistant to ≥ 8 antimicrobial agents | |
|-----------------|-------|-----|--|----|--|----|--|----|--|-----|
| | N | % | N | % | N | % | N | % | N | % |
| Typhimurium | 325 | 23 | 165 | 51 | 156 | 48 | 114 | 35 | 5 | 1 |
| Enteritidis | 282 | 20 | 40 | 14 | 16 | 6 | 3 | 1 | 1 | 0.4 |
| Newport | 124 | 9 | 43 | 35 | 39 | 31 | 33 | 27 | 31 | 25 |
| Heidelberg | 100 | 7 | 36 | 36 | 30 | 30 | 5 | 5 | 1 | 1 |
| Javiana | 51 | 4 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Oranienburg | 33 | 2 | 1 | 3 | 1 | 3 | 0 | 0 | 0 | 0 |
| Montevideo | 29 | 2 | 3 | 10 | 2 | 7 | 1 | 3 | 0 | 0 |
| Muenchen | 29 | 2 | 4 | 14 | 4 | 14 | 0 | 0 | 0 | 0 |
| Thompson | 27 | 2 | 1 | 4 | 0 | 0 | 0 | 0 | 0 | 0 |
| Saint Paul | 21 | 1 | 4 | 19 | 4 | 19 | 1 | 5 | 1 | 5 |
| Berta | 18 | 1 | 8 | 44 | 3 | 17 | 0 | 0 | 0 | 0 |
| Braenderup | 18 | 1 | 4 | 22 | 2 | 11 | 0 | 0 | 0 | 0 |
| Agona | 17 | 1 | 9 | 53 | 5 | 29 | 0 | 0 | 0 | 0 |
| Infantis | 17 | 1 | 1 | 6 | 1 | 6 | 0 | 0 | 0 | 0 |
| Java | 17 | 1 | 3 | 18 | 3 | 18 | 3 | 18 | 0 | 0 |
| Stanley | 17 | 1 | 8 | 47 | 8 | 47 | 1 | 6 | 0 | 0 |
| Other Serotypes | 274 | 19 | 64 | 23 | 41 | 15 | 15 | 5 | 1 | 0.4 |
| Total Serotyped | 1399 | 100 | 394 | 28 | 315 | 22 | 176 | 13 | 40 | 3 |
| Not Serotyped | 20 | NA | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |

*Multidrug resistant to 18 antimicrobial agents tested in 2001: amikacin, amoxicillin-clavulanic acid, ampicillin, apramycin, cefoxitin, ceftiofur, ceftriaxone, cephalothin, chloramphenicol, ciprofloxacin, gentamicin, imipenem, kanamycin, nalidixic acid, streptomycin, sulfamethoxazole, tetracycline, trimethoprim-sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 9. Frequency of multidrug* resistance among the most common non-Typhi *Salmonella* serotypes in each site, 2001

| Site | Most Common Serotypes | Total | | Number resistant to ≥ 1 antimicrobial agents | | Number resistant to ≥ 2 antimicrobial agents | | Number resistant to ≥ 5 antimicrobial agents | | Number resistant to ≥ 8 antimicrobial agents | |
|-------------------|-----------------------|-------|-----|---|----|---|----|---|----|---|----|
| | | N | % | N | % | N | % | N | % | N | % |
| California | Heidelberg | 9 | 17 | 2 | 22 | 2 | 22 | 0 | 0 | 0 | 0 |
| | Typhimurium | 9 | 17 | 5 | 55 | 4 | 44 | 2 | 22 | 0 | 0 |
| | Newport | 6 | 11 | 5 | 83 | 5 | 83 | 5 | 83 | 5 | 83 |
| | Enteritidis | 5 | 9 | 1 | 20 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Oranienburg | 3 | 6 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Other serotypes | 21 | 40 | 2 | 9 | 1 | 5 | 0 | 0 | 0 | 0 |
| | Total | 53 | 100 | 15 | 28 | 12 | 23 | 7 | 13 | 5 | 9 |
| Colorado | Typhimurium | 20 | 32 | 10 | 50 | 10 | 50 | 8 | 40 | 1 | 5 |
| | Enteritidis | 8 | 13 | 1 | 12 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Heidelberg | 8 | 13 | 2 | 25 | 2 | 25 | 0 | 0 | 0 | 0 |
| | Braenderup | 4 | 6 | 2 | 50 | 2 | 50 | 0 | 0 | 0 | 0 |
| | Newport | 4 | 6 | 1 | 25 | 1 | 25 | 1 | 25 | 1 | 25 |
| | Other | 19 | 30 | 4 | 21 | 2 | 10 | 0 | 0 | 0 | 0 |
| | Total | 63 | 100 | 20 | 32 | 17 | 27 | 9 | 14 | 2 | 3 |

*Multidrug resistant to 18 antimicrobial agents tested in 2001: amikacin, amoxicillin-clavulanic acid, ampicillin, apramycin, cefoxitin, ceftiofur, ceftriaxone, cephalothin, chloramphenicol, ciprofloxacin, gentamicin, imipenem, kanamycin, nalidixic acid, streptomycin, sulfamethoxazole, tetracycline, trimethoprim-sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 9. Frequency of multidrug* resistance among the most common non-Typhi *Salmonella* serotypes in each site, 2001

| Site | Most Common Serotypes | Total | | Number resistant to ≥ 1 antimicrobial agents | | Number resistant to ≥ 2 antimicrobial agents | | Number resistant to ≥ 5 antimicrobial agents | | Number resistant to ≥ 8 antimicrobial agents | |
|--------------------|-----------------------|-------|-----|--|----|--|----|--|----|--|----|
| | | N | % | N | % | N | % | N | % | N | % |
| Connecticut | Enteritidis | 15 | 30 | 4 | 27 | 1 | 7 | 1 | 7 | 1 | 7 |
| | Typhimurium | 11 | 22 | 8 | 73 | 8 | 73 | 6 | 54 | 1 | 9 |
| | Newport | 7 | 14 | 3 | 43 | 3 | 43 | 3 | 43 | 3 | 43 |
| | Montevideo | 3 | 6 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Oranienburg | 2 | 4 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Thompson | 2 | 4 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Other | 10 | 20 | 2 | 20 | 2 | 20 | 0 | 0 | 0 | 0 |
| | Total | 50 | 100 | 17 | 34 | 14 | 28 | 10 | 20 | 5 | 10 |
| Florida | Javiana | 15 | 27 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Newport | 10 | 18 | 1 | 10 | 1 | 10 | 1 | 10 | 0 | 0 |
| | Enteritidis | 7 | 13 | 2 | 29 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Typhimurium | 5 | 9 | 2 | 40 | 2 | 40 | 1 | 20 | 0 | 0 |
| | Lindenberg | 2 | 4 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Manhattan | 2 | 4 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Muenchen | 2 | 4 | 1 | 50 | 1 | 50 | 0 | 0 | 0 | 0 |
| | Rubislaw | 2 | 4 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Other | 10 | 18 | 1 | 10 | 1 | 10 | 1 | 10 | 0 | 0 |
| | Total | 55 | 100 | 7 | 13 | 5 | 9 | 3 | 5 | 0 | 0 |

*Multidrug resistant to 18 antimicrobial agents tested in 2001: amikacin, amoxicillin-clavulanic acid, ampicillin, apramycin, cefoxitin, ceftiofur, ceftriaxone, cephalothin, chloramphenicol, ciprofloxacin, gentamicin, imipenem, kanamycin, nalidixic acid, streptomycin, sulfamethoxazole, tetracycline, trimethoprim-sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 9. Frequency of multidrug* resistance among the most common non-Typhi *Salmonella* serotypes in each site, 2001

| Site | Most Common Serotypes | Total | | Number resistant to ≥ 1 antimicrobial agents | | Number resistant to ≥ 2 antimicrobial agents | | Number resistant to ≥ 5 antimicrobial agents | | Number resistant to ≥ 8 antimicrobial agents | |
|----------------|-----------------------|------------|------------|--|-----------|--|-----------|--|-----------|--|------------|
| | | N | % | N | % | N | % | N | % | N | % |
| Georgia | Typhimurium | 38 | 20 | 22 | 58 | 21 | 55 | 16 | 42 | 0 | 0 |
| | Newport | 35 | 19 | 3 | 9 | 2 | 6 | 1 | 3 | 1 | 3 |
| | Javiana | 18 | 10 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Heidelberg | 15 | 8 | 9 | 60 | 9 | 60 | 0 | 0 | 0 | 0 |
| | Enteritidis | 11 | 6 | 2 | 18 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Not serotyped | 6 | 3 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Other serotypes | 62 | 33 | 8 | 13 | 6 | 10 | 1 | 2 | 0 | 0 |
| | Total | 185 | 100 | 44 | 52 | 38 | 20 | 18 | 10 | 1 | 0.5 |
| Kansas | Typhimurium | 9 | 32 | 3 | 33 | 3 | 33 | 2 | 22 | 0 | 0 |
| | Enteritidis | 5 | 18 | 1 | 20 | 1 | 20 | 1 | 20 | 0 | 0 |
| | Newport | 5 | 18 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Heidelberg | 2 | 7 | 2 | 100 | 2 | 100 | 0 | 0 | 0 | 0 |
| | Oranienburg | 2 | 7 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Not serotyped | 1 | 4 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Other serotypes | 4 | 14 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Total | 28 | 100 | 6 | 21 | 6 | 21 | 3 | 11 | 0 | 0 |

*Multidrug resistant to 18 antimicrobial agents tested in 2001: amikacin, amoxicillin-clavulanic acid, ampicillin, apramycin, cefoxitin, ceftiofur, ceftriaxone, cephalothin, chloramphenicol, ciprofloxacin, gentamicin, imipenem, kanamycin, nalidixic acid, streptomycin, sulfamethoxazole, tetracycline, trimethoprim-sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 9. Frequency of multidrug* resistance among the most common non-Typhi *Salmonella* serotypes in each site, 2001

| Site | Most Common Serotypes | Total | | Number resistant to ≥ 1 antimicrobial agents | | Number resistant to ≥ 2 antimicrobial agents | | Number resistant to ≥ 5 antimicrobial agents | | Number resistant to ≥ 8 antimicrobial agents | |
|---------------------------|-----------------------|-------|-----|--|-----|--|----|--|----|--|----|
| | | N | % | N | % | N | % | N | % | N | % |
| Los Angeles County | Enteritidis | 21 | 21 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Typhimurium | 18 | 18 | 8 | 44 | 8 | 44 | 6 | 33 | 0 | 0 |
| | Newport | 8 | 8 | 4 | 50 | 3 | 37 | 3 | 37 | 3 | 37 |
| | Heidelberg | 7 | 7 | 3 | 43 | 3 | 43 | 0 | 0 | 0 | 0 |
| | Agona | 6 | 6 | 3 | 50 | 2 | 33 | 0 | 0 | 0 | 0 |
| | Other serotypes | 42 | 41 | 8 | 19 | 6 | 14 | 1 | 2 | 0 | 0 |
| | Total | 102 | 100 | 26 | 25 | 22 | 22 | 10 | 10 | 3 | 3 |
| Maryland | Enteritidis | 26 | 30 | 4 | 15 | 3 | 11 | 1 | 4 | 0 | 0 |
| | Typhimurium | 16 | 19 | 11 | 69 | 11 | 69 | 10 | 62 | 0 | 0 |
| | Oranienburg | 6 | 7 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Newport | 5 | 6 | 2 | 40 | 2 | 40 | 2 | 40 | 2 | 40 |
| | Hadar | 3 | 3 | 3 | 100 | 2 | 67 | 0 | 0 | 0 | 0 |
| | Javiana | 3 | 3 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Not serotyped | 2 | 2 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Other serotypes | 25 | 29 | 6 | 24 | 4 | 16 | 0 | 0 | 0 | 0 |
| | Total | 86 | 100 | 26 | 30 | 22 | 26 | 11 | 13 | 2 | 2 |

*Multidrug resistant to 18 antimicrobial agents tested in 2001: amikacin, amoxicillin-clavulanic acid, ampicillin, apramycin, cefoxitin, ceftiofur, ceftriaxone, cephalothin, chloramphenicol, ciprofloxacin, gentamicin, imipenem, kanamycin, nalidixic acid, streptomycin, sulfamethoxazole, tetracycline, trimethoprim-sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 9. Frequency of multidrug* resistance among the most common non-Typhi *Salmonella* serotypes in each site, 2001

| Site | Most Common Serotypes | Total | | Number resistant to ≥ 1 antimicrobial agents | | Number resistant to ≥ 2 antimicrobial agents | | Number resistant to ≥ 5 antimicrobial agents | | Number resistant to ≥ 8 antimicrobial agents | |
|----------------------|------------------------|------------|------------|--|-----------|--|-----------|--|-----------|--|----------|
| | | N | % | N | % | N | % | N | % | N | % |
| Massachusetts | Enteritidis | 46 | 35 | 7 | 15 | 4 | 9 | 0 | 0 | 0 | 0 |
| | Typhimurium | 26 | 20 | 12 | 46 | 12 | 46 | 9 | 35 | 0 | 0 |
| | Heidelberg | 7 | 5 | 2 | 29 | 1 | 14 | 1 | 14 | 0 | 0 |
| | Newport | 6 | 5 | 4 | 67 | 3 | 50 | 2 | 33 | 2 | 33 |
| | Monophasic Typhimurium | 5 | | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Not serotyped | 2 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Other serotypes | 39 | 30 | 6 | 15 | 4 | 10 | 4 | 10 | 1 | 3 |
| | Total | 131 | 100 | 31 | 24 | 24 | 18 | 14 | 11 | 3 | 2 |
| Minnesota | Typhimurium | 23 | 36 | 11 | 48 | 11 | 48 | 8 | 35 | 1 | 4 |
| | Enteritidis | 11 | 17 | 2 | 18 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Heidelberg | 6 | 9 | 2 | 33 | 2 | 33 | 1 | 17 | 0 | 0 |
| | Java | 3 | 5 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Newport | 3 | 5 | 1 | 33 | 1 | 33 | 1 | 33 | 1 | 33 |
| | Other serotypes | 15 | 24 | 2 | 13 | 1 | 7 | 0 | 0 | 0 | 0 |
| | Not serotyped | 2 | 3 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Total | 63 | 100 | 18 | 29 | 15 | 24 | 11 | 17 | 2 | 3 |

*Multidrug resistant to 18 antimicrobial agents tested in 2001: amikacin, amoxicillin-clavulanic acid, ampicillin, apramycin, cefoxitin, ceftiofur, ceftriaxone, cephalothin, chloramphenicol, ciprofloxacin, gentamicin, imipenem, kanamycin, nalidixic acid, streptomycin, sulfamethoxazole, tetracycline, trimethoprim-sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 9. Frequency of multidrug* resistance among the most common non-Typhi *Salmonella* serotypes in each site, 2001

| Site | Most Common Serotypes | Total | | Number resistant to ≥ 1 antimicrobial agents | | Number resistant to ≥ 2 antimicrobial agents | | Number resistant to ≥ 5 antimicrobial agents | | Number resistant to ≥ 8 antimicrobial agents | |
|--------------|------------------------|------------|------------|---|-----------|---|-----------|---|-----------|---|----------|
| | | N | % | N | % | N | % | N | % | N | % |
| New Jersey | Enteritidis | 32 | 27 | 3 | 9 | 2 | 6 | 0 | 0 | 0 | 0 |
| | Typhimurium | 28 | 24 | 20 | 74 | 20 | 74 | 13 | 48 | 0 | 0 |
| | Newport | 8 | 7 | 3 | 37 | 2 | 25 | 1 | 12 | 1 | 12 |
| | Heidelberg | 4 | 3 | 1 | 25 | 1 | 25 | 0 | 0 | 0 | 0 |
| | Braenderup | 3 | 2 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Infantis | 3 | 2 | 1 | 33 | 1 | 33 | 0 | 0 | 0 | 0 |
| | Javiana | 3 | 2 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Monophasic Typhimurium | 3 | 2 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Thompson | 3 | 2 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Not serotyped | 2 | 2 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Other serotypes | 29 | 24 | 6 | 21 | 4 | 14 | 1 | 3 | 0 | 0 |
| Total | | 118 | 100 | 34 | 29 | 30 | 25 | 15 | 13 | 1 | 1 |

*Multidrug resistant to 18 antimicrobial agents tested in 2001: amikacin, amoxicillin-clavulanic acid, ampicillin, apramycin, cefoxitin, ceftiofur, ceftriaxone, cephalothin, chloramphenicol, ciprofloxacin, gentamicin, imipenem, kanamycin, nalidixic acid, streptomycin, sulfamethoxazole, tetracycline, trimethoprim-sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 9. Frequency of multidrug* resistance among the most common non-Typhi *Salmonella* serotypes in each site, 2001

| Site | Most Common Serotypes | Total | | Number resistant to ≥ 1 antimicrobial agents | | Number resistant to ≥ 2 antimicrobial agents | | Number resistant to ≥ 5 antimicrobial agents | | Number resistant to ≥ 8 antimicrobial agents | |
|----------------|-----------------------|------------|------------|--|-----------|--|-----------|--|-----------|--|----------|
| | | N | % | N | % | N | % | N | % | N | % |
| New York City | Enteritidis | 43 | 31 | 9 | 21 | 3 | 7 | 0 | 0 | 0 | 0 |
| | Typhimurium | 32 | 23 | 17 | 8 | 14 | 44 | 7 | 22 | 0 | 0 |
| | Berta | 7 | 5 | 5 | 71 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Heidelberg | 6 | 4 | 2 | 33 | 1 | 17 | 0 | 0 | 0 | 0 |
| | Muenchen | 4 | 3 | 1 | 25 | 1 | 25 | 0 | 0 | 0 | 0 |
| | Uganda | 4 | 3 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Other serotypes | 43 | 31 | 19 | 44 | 12 | 28 | 5 | 12 | 0 | 0 |
| | Total | 139 | 100 | 53 | 38 | 31 | 22 | 12 | 9 | 0 | 0 |
| New York State | Typhimurium | 36 | 27 | 14 | 39 | 11 | 30 | 10 | 28 | 0 | 0 |
| | Enteritidis | 23 | 17 | 1 | 4 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Heidelberg | 16 | 12 | 4 | 25 | 1 | 6 | 1 | 6 | 0 | 0 |
| | Newport | 8 | 6 | 6 | 75 | 6 | 75 | 4 | 50 | 4 | 50 |
| | Thompson | 7 | 5 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Not serotyped | 1 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Other serotypes | 44 | 33 | 8 | 18 | 6 | 14 | 3 | 7 | 0 | 0 |
| | Total | 135 | 100 | 33 | 24 | 24 | 18 | 18 | 13 | 4 | 3 |

*Multidrug resistant to 18 antimicrobial agents tested in 2001: amikacin, amoxicillin-clavulanic acid, ampicillin, apramycin, cefoxitin, ceftiofur, ceftriaxone, cephalothin, chloramphenicol, ciprofloxacin, gentamicin, imipenem, kanamycin, nalidixic acid, streptomycin, sulfamethoxazole, tetracycline, trimethoprim-sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 9. Frequency of multidrug* resistance among the most common non-Typhi *Salmonella* serotypes in each site, 2001

| Site | Most Common Serotypes | Total | | Number resistant to ≥ 1 antimicrobial agents | | Number resistant to ≥ 2 antimicrobial agents | | Number resistant to ≥ 5 antimicrobial agents | | Number resistant to ≥ 8 antimicrobial agents | |
|------------------|-----------------------|-------|-----|---|----|---|----|---|----|---|----|
| | | N | % | N | % | N | % | N | % | N | % |
| Oregon | Typhimurium | 10 | 38 | 2 | 20 | 2 | 20 | 2 | 20 | 0 | 0 |
| | Enteritidis | 4 | 15 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Muenchen | 3 | 11 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Montevideo | 2 | 8 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Other serotypes | 7 | 27 | 2 | 29 | 1 | 14 | 0 | 0 | 0 | 0 |
| | Total | 26 | 100 | 4 | 15 | 3 | 11 | 2 | 8 | 0 | 0 |
| Tennessee | Typhimurium | 23 | 27 | 12 | 52 | 11 | 48 | 9 | 39 | 0 | 0 |
| | Newport | 9 | 11 | 2 | 22 | 2 | 22 | 2 | 22 | 2 | 22 |
| | Heidelberg | 7 | 8 | 2 | 29 | 2 | 29 | 0 | 0 | 0 | 0 |
| | Javiana | 5 | 6 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Mississippi | 5 | 6 | 1 | 20 | 1 | 20 | 0 | 0 | 0 | 0 |
| | Not serotyped | 3 | 4 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Other serotypes | 32 | 38 | 7 | 22 | 6 | 19 | 2 | 6 | 0 | 0 |
| | Total | 84 | 100 | 24 | 29 | 22 | 26 | 13 | 15 | 2 | 2 |

*Multidrug resistant to 18 antimicrobial agents tested in 2001: amikacin, amoxicillin-clavulanic acid, ampicillin, apramycin, cefoxitin, ceftiofur, ceftriaxone, cephalothin, chloramphenicol, ciprofloxacin, gentamicin, imipenem, kanamycin, nalidixic acid, streptomycin, sulfamethoxazole, tetracycline, trimethoprim-sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 9. Frequency of multidrug* resistance among the most common non-Typhi *Salmonella* serotypes in each site, 2001

| Site | Most Common Serotypes | Total | | Number resistant to ≥ 1 antimicrobial agents | | Number resistant to ≥ 2 antimicrobial agents | | Number resistant to ≥ 5 antimicrobial agents | | Number resistant to ≥ 8 antimicrobial agents | |
|---------------|-----------------------|-----------|------------|--|-----------|--|-----------|--|-----------|--|-----------|
| | | N | % | N | % | N | % | N | % | N | % |
| Washington | Enteritidis | 15 | 20 | 2 | 13 | 2 | 13 | 0 | 0 | 0 | 0 |
| | Typhimurium | 15 | 20 | 8 | 53 | 8 | 53 | 5 | 33 | 2 | 13 |
| | Saint Paul | 10 | 13 | 3 | 30 | 3 | 30 | 1 | 10 | 1 | 10 |
| | Heidelberg | 9 | 12 | 4 | 44 | 3 | 33 | 2 | 22 | 1 | 11 |
| | Newport | 7 | 9 | 5 | 71 | 5 | 71 | 4 | 57 | 4 | 57 |
| | Not serotyped | 1 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Other serotypes | 19 | 25 | 7 | 37 | 4 | 21 | 1 | 5 | 0 | 0 |
| | Total | 76 | 100 | 29 | 38 | 25 | 33 | 13 | 17 | 8 | 10 |
| West Virginia | Enteritidis | 6 | 24 | 1 | 17 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Typhimurium | 6 | 24 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Montevideo | 3 | 12 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Newport | 2 | 8 | 2 | 100 | 2 | 100 | 2 | 100 | 2 | 100 |
| | Paratyphi | 2 | 8 | 1 | 50 | 1 | 50 | 1 | 50 | 0 | 0 |
| | Other serotypes | 6 | 24 | 2 | 33 | 2 | 33 | 0 | 0 | 0 | 0 |
| | Total | 25 | 100 | 6 | 24 | 5 | 20 | 3 | 12 | 2 | 8 |

*Multidrug resistant to 18 antimicrobial agents tested in 2001: amikacin, amoxicillin-clavulanic acid, ampicillin, apramycin, cefoxitin, ceftiofur, ceftriaxone, cephalothin, chloramphenicol, ciprofloxacin, gentamicin, imipenem, kanamycin, nalidixic acid, streptomycin, sulfamethoxazole, tetracycline, trimethoprim-sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 10. *Salmonella* Typhimurium isolates with at least ACSSuT*, ACKSSuT, or AKSSuT*** resistance patterns, by site, 2001**

| Site | # non-Typhi <i>Salmonella</i> isolates tested | <i>Salmonella</i> Typhimurium | | ACSSuT Typhimurium | | ACKSSuT Typhimurium | | AKSSuT Typhimurium | |
|----------------|---|----------------------------------|-----------|-----------------------|-----------|------------------------|----------|-----------------------|----------|
| | | N | % | N | % | N | % | N | % |
| California | 53 | 9 | 17 | 1 | 11 | 0 | 0 | 1 | 11 |
| Colorado | 63 | 20 | 32 | 5 | 26 | 0 | 0 | 3 | 15 |
| Connecticut | 50 | 11 | 22 | 5 | 45 | 0 | 0 | 0 | 0 |
| Florida | 55 | 5 | 9 | 1 | 20 | 0 | 0 | 0 | 0 |
| Georgia | 185 | 38 | 20 | 15 | 39 | 2 | 5 | 3 | 8 |
| Kansas | 28 | 9 | 32 | 1 | 12 | 0 | 0 | 1 | 12 |
| Los Angeles | 102 | 18 | 18 | 4 | 22 | 0 | 0 | 2 | 11 |
| Maryland | 86 | 16 | 19 | 8 | 50 | 0 | 0 | 0 | 0 |
| Massachusetts | 131 | 26 | 20 | 7 | 27 | 0 | 0 | 1 | 4 |
| Minnesota | 63 | 23 | 36 | 6 | 26 | 0 | 0 | 1 | 4 |
| New Jersey | 118 | 28 | 24 | 13 | 48 | 0 | 0 | 0 | 0 |
| New York City | 139 | 32 | 23 | 6 | 19 | 1 | 3 | 1 | 3 |
| New York State | 135 | 36 | 27 | 8 | 22 | 0 | 0 | 1 | 3 |
| Oregon | 26 | 10 | 38 | 2 | 20 | 0 | 0 | 0 | 0 |
| Tennessee | 84 | 23 | 27 | 9 | 39 | 0 | 0 | 0 | 0 |
| Washington | 76 | 15 | 20 | 5 | 33 | 1 | 7 | 1 | 7 |
| West Virginia | 25 | 6 | 24 | 0 | 0 | 0 | 0 | 0 | 0 |
| Totals | 1419 | 325 | 23 | 96 | 30 | 4 | 1 | 15 | 5 |

*ACSSuT= Ampicillin, Chloramphenicol, Streptomycin, Sulfamethoxazole, Tetracycline

**ACKSSuT= Ampicillin, Chloramphenicol, Kanamycin, Streptomycin, Sulfamethoxazole, Tetracycline

***AKSSuT= Ampicillin, Kanamycin, Streptomycin, Sulfamethoxazole, Tetracycline

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 11. Additional antimicrobial resistance for *Salmonella* Typhimurium isolates with at least ACSSuT*, ACKSSuT, or AKSSuT*** resistance patterns, 2001**

| Antimicrobial Agent | ACSSuT (N=96) | | ACKSSuT (N=4) | | AKSSuT (N=15) | |
|-------------------------------|--------------------------|----------|--------------------------|----------|--------------------------|----------|
| | N | % | N | % | N | % |
| Amikacin | 0 | 0 | 0 | 0 | 0 | 0 |
| Amoxicillin-Clavulanic Acid | 13 | 13 | 1 | 25 | 1 | 7 |
| Apramycin | 0 | 0 | 0 | 0 | 0 | 0 |
| Cefoxitin | 4 | 4 | 1 | 25 | 1 | 7 |
| Ceftiofur | 4 | 4 | 1 | 25 | 1 | 7 |
| Ceftriaxone | 1 | 1 | 1 | 25 | 1 | 7 |
| Cephalothin | 4 | 4 | 1 | 25 | 1 | 7 |
| Chloramphenicol | NA | | NA | | 4 | 27 |
| Ciprofloxacin | 1 | 1 | 0 | 0 | 0 | 0 |
| Gentamicin | 2 | 2 | 0 | 0 | 1 | 7 |
| Kanamycin | 4 | 4 | NA | | NA | |
| Nalidixic Acid | 1 | 1 | 0 | 0 | 0 | 0 |
| Trimethoprim-Sulfamethoxazole | 3 | 3 | 0 | 0 | 1 | 7 |

**ACSSuT=Ampicillin, Chloramphenicol, Streptomycin, Sulfamethoxazole, Tetracycline

***ACKSSuT=Ampicillin, Chloramphenicol, Kanamycin, Streptomycin, Sulfamethoxazole, Tetracycline

****AKSSuT=Ampicillin, Kanamycin, Streptomycin, Sulfamethoxazole, Tetracycline

NA: Not Applicable

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 12. *Salmonella* Newport isolates with at least MDR-AmpC* resistance pattern, by site, 2001

| Site | # non-Typhi <i>Salmonella</i> isolates tested | <i>Salmonella</i> Newport | | MDR-AmpC Newport | |
|----------------|---|---------------------------|----------|---------------------|-----------|
| | | N | % | N | % |
| California | 53 | 6 | 11 | 5 | 83 |
| Colorado | 63 | 4 | 6 | 1 | 25 |
| Connecticut | 50 | 7 | 14 | 3 | 43 |
| Florida | 55 | 10 | 18 | 0 | 0 |
| Georgia | 185 | 35 | 19 | 1 | 3 |
| Kansas | 28 | 5 | 18 | 0 | 0 |
| Los Angeles | 102 | 8 | 8 | 3 | 37 |
| Maryland | 86 | 5 | 6 | 2 | 40 |
| Massachusetts | 131 | 6 | 5 | 2 | 33 |
| Minnesota | 63 | 3 | 5 | 1 | 33 |
| New Jersey | 118 | 8 | 7 | 1 | 12 |
| New York City | 139 | 1 | 1 | 0 | 0 |
| New York State | 135 | 8 | 6 | 4 | 50 |
| Oregon | 26 | 0 | 0 | 0 | 0 |
| Tennessee | 84 | 9 | 11 | 2 | 22 |
| Washington | 76 | 7 | 9 | 4 | 57 |
| West Virginia | 25 | 2 | 8 | 2 | 100 |
| Totals | 1419 | 124 | 9 | 31 | 25 |

*MDR-AmpC=Resistance to Amoxicillin/Clavulanic Acid, Ampicillin, Cephalothin, Cefoxitin, Ceftiofur, Chloramphenicol, Sulfamethoxazole, Streptomycin and Tetracycline, and decreased susceptibility to Ceftriaxone (MIC \geq 16 μ g/ml)

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 13. Additional antimicrobial resistance for *Salmonella* Newport isolates with at least MDR-AmpC* resistance pattern, 2001

| Antimicrobial Agent | MDR-AmpC (N=31) | |
|-------------------------------|----------------------------|----------|
| | N | % |
| Amikacin | 0 | 0 |
| Apramycin | 0 | 0 |
| Ceftriaxone | 29 | 93 |
| Ciprofloxacin | 0 | 0 |
| Gentamicin | 2 | 6 |
| Kanamycin | 6 | 19 |
| Nalidixic Acid | 0 | 0 |
| Trimethoprim-Sulfamethoxazole | 1 | 3 |

*MDR-AmpC=Resistance to Amoxicillin/Clavulanic Acid, Ampicillin, Cephalothin, Cefoxitin, Ceftiofur, Chloramphenicol, Sulfamethoxazole, Streptomycin and Tetracycline, and decreased susceptibility to Ceftriaxone (MIC \geq 16 $\mu\text{g/ml}$)

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 14. Most common multidrug* resistant patterns among non-Typhi *Salmonella*, 2001

| Resistance pattern | Number | % of <i>Salmonella</i> | % of multidrug-resistant <i>Salmonella</i> |
|--------------------------|--------|------------------------|--|
| S. Typhimurium ACSSuT** | 96 | 7 | 30 |
| S. Typhimurium AKSSuT*** | 15 | 1 | 5 |
| S. Newport MDR-AmpC**** | 31 | 2 | 10 |

*Multidrug resistant to 18 antimicrobial agents tested in 2001: amikacin, amoxicillin-clavulanic acid, ampicillin, apramycin, cefoxitin, ceftiofur, ceftriaxone, cephalothin, chloramphenicol, ciprofloxacin, gentamicin, imipenem, kanamycin, nalidixic acid, streptomycin, sulfamethoxazole, tetracycline, trimethoprim-sulfamethoxazole

**ACSSuT = Ampicillin, Chloramphenicol, Streptomycin, Sulfamethoxazole, and Tetracycline

***AKSSuT = Ampicillin, Kanamycin, Streptomycin, Sulfamethoxazole, and Tetracycline

****MDR-AmpC = Resistance to Amoxicillin/Clavulanic Acid, Ampicillin, Cephalothin, Cefoxitin, Ceftiofur, Chloramphenicol, Sulfamethoxazole, Streptomycin and Tetracycline, and decreased susceptibility to Ceftriaxone (MIC \geq 16 $\mu\text{g/ml}$)

National Antimicrobial Resistance Monitoring System for Enteric Bacteria

Table 15. Clinical source of non-Typhi *Salmonella* isolates, 2001

| Isolate | Blood | | Stool | | Other | | Unknown | | Total | |
|--------------------------|--------------|----------|--------------|-----------|--------------|----------|----------------|----------|--------------|------------|
| | N | % | N | % | N | % | N | % | N | % |
| S. Typhimurium | 11 | 3 | 283 | 87 | 23 | 7 | 8 | 2 | 325 | 23 |
| S. Typhimurium -- ACSSuT | 3 | 3 | 85 | 88 | 7 | 7 | 1 | 1 | 96 | 7 |
| S. Typhimurium -- AKSSuT | 1 | 7 | 14 | 93 | 0 | 0 | 0 | 0 | 15 | 1 |
| All other S. Typhimurium | 7 | 3 | 184 | 86 | 16 | 7 | 7 | 3 | 214 | 15 |
| S. Enteritidis | 16 | 6 | 251 | 89 | 8 | 3 | 7 | 2 | 282 | 20 |
| S. Newport | 0 | 0 | 116 | 93 | 7 | 6 | 1 | 1 | 124 | 9 |
| S. Newport -- MDR-AmpC | 0 | 0 | 28 | 90 | 3 | 10 | 0 | 0 | 31 | 2 |
| All other S. Newport | 0 | 0 | 88 | 95 | 4 | 4 | 1 | 1 | 93 | 6 |
| S. Heidelberg | 11 | 11 | 75 | 75 | 9 | 9 | 5 | 5 | 100 | 7 |
| S. Javiana | 0 | 0 | 47 | 92 | 3 | 6 | 1 | 2 | 51 | 4 |
| S. Oranienburg | 5 | 15 | 25 | 76 | 1 | 3 | 2 | 6 | 33 | 2 |
| S. Montevideo | 0 | 0 | 27 | 93 | 1 | 3 | 1 | 3 | 29 | 2 |
| S. Muenchen | 0 | 0 | 25 | 86 | 1 | 3 | 3 | 10 | 29 | 2 |
| S. Thompson | 1 | 4 | 25 | 93 | 1 | 4 | 0 | 0 | 27 | 2 |
| S. St. Paul | 0 | 0 | 18 | 86 | 3 | 14 | 0 | 0 | 21 | 1 |
| S. Berta | 1 | 5 | 14 | 78 | 3 | 17 | 0 | 0 | 18 | 1 |
| S. Braenderup | 0 | 0 | 17 | 94 | 1 | 6 | 0 | 0 | 18 | 1 |
| S. Agona | 0 | 0 | 16 | 94 | 0 | 0 | 1 | 6 | 17 | 1 |
| S. Infantis | 1 | 6 | 13 | 76 | 2 | 12 | 1 | 6 | 17 | 1 |
| S. Java | 1 | 6 | 12 | 71 | 3 | 18 | 1 | 6 | 17 | 1 |
| S. Stanley | 1 | 6 | 16 | 94 | 0 | 0 | 0 | 0 | 17 | 1 |
| Other <i>Salmonella</i> | 24 | 8 | 254 | 82 | 21 | 7 | 12 | 4 | 311 | 22 |
| Total | 71 | 5 | 1218 | 86 | 87 | 6 | 43 | 3 | 1419 | 100 |

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 16. Proportion of non-Typhi *Salmonella* isolates (N=1419) with clinically important resistance: decreased susceptibility or resistance to ciprofloxacin or ceftriaxone, 2001

| Antimicrobial Agent | # isolates with decreased susceptibility* | % of <i>Salmonella</i> | # of resistant isolates** | % of <i>Salmonella</i> |
|---------------------|---|------------------------|---------------------------|------------------------|
| Ciprofloxacin | 15 | 1 | 3 | 0.2 |
| Ceftriaxone | 48 | 3 | 34 | 2 |

*Ciprofloxacin = MIC \geq 0.25 $\mu\text{g}/\text{ml}$; Ceftriaxone = MIC \geq 16 $\mu\text{g}/\text{ml}$

**Ciprofloxacin = MIC \geq 4 $\mu\text{g}/\text{ml}$; Ceftriaxone = MIC \geq 64 $\mu\text{g}/\text{ml}$

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 17. Proportion of non-Typhi *Salmonella* isolates submitted, by site, with decreased susceptibility to ciprofloxacin (MIC \geq 0.25 $\mu\text{g/ml}$), 2001

| Site | # isolates with decreased susceptibility | # non-Typhi <i>Salmonella</i> isolates tested | % with decreased susceptibility | Serotype |
|----------------|--|---|---------------------------------|---|
| California | 1 | 53 | 2 | Paratyphi (1) |
| Colorado | 1 | 63 | 2 | Paratyphi (1) |
| Connecticut | 1 | 50 | 2 | Haardt (1) |
| Florida | 1 | 55 | 2 | Senftenberg* (1) |
| Georgia | 0 | 185 | 0 | |
| Kansas | 0 | 28 | 0 | |
| Los Angeles | 1 | 102 | 1 | Blockley (1) |
| Maryland | 0 | 86 | 0 | |
| Massachusetts | 2 | 131 | 1 | Agona (1), Senftenberg*(1) |
| Minnesota | 0 | 63 | 0 | |
| New Jersey | 3 | 118 | 2 | Paratyphi (2), Haifa (1) |
| New York City | 2 | 139 | 1 | Typhimurium (1), Virchow (1) |
| New York State | 0 | 135 | 0 | |
| Oregon | 0 | 26 | 0 | |
| Tennessee | 1 | 84 | 1 | Uganda (1) |
| Washington | 2 | 76 | 3 | Paratyphi (1), Typhimurium* (1) |
| West Virginia | 0 | 25 | 0 | |
| Total | 15 | 1419 | 1 | Paratyphi (5), Senftenberg* (2), Typhimurium* (2), Agona (1), Blockley (1), Haardt (1), Haifa (1), Uganda (1), Virchow (1) |

*These isolates were also resistant to ciprofloxacin

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 18. Proportion of non-Typhi *Salmonella* isolates submitted, by site, with decreased susceptibility to ceftriaxone (MIC \geq 16 $\mu\text{g/ml}$), 2001

| Site | # isolates with decreased susceptibility | # non-Typhi <i>Salmonella</i> isolates tested | % with decreased susceptibility | Serotype |
|----------------|--|---|---------------------------------|---|
| California | 5 | 53 | 9 | Newport* (5) |
| Colorado | 2 | 63 | 2 | Newport* (1), Typhimurium (1) |
| Connecticut | 6 | 50 | 12 | Newport* (3), Typhimurium* (2), Enteritidis* (1) |
| Florida | 1 | 55 | 2 | Senftenberg (1) |
| Georgia | 1 | 185 | 0.5 | Newport* (1) |
| Kansas | 0 | 28 | 0 | |
| Los Angeles | 3 | 102 | 3 | Newport* (3) |
| Maryland | 4 | 86 | 5 | Newport* (2), Typhimurium* (2) |
| Massachusetts | 4 | 131 | 3 | Newport* (2), Heidelberg (1), Typhimurium (1) |
| Minnesota | 3 | 63 | 5 | Montevideo (1), Newport* (1), Typhimurium (1) |
| New Jersey | 1 | 118 | 1 | Newport* (1) |
| New York City | 1 | 139 | 1 | Typhimurium (1) |
| New York State | 7 | 135 | 5 | Newport* (4), Heidelberg (1), Stanley (1), Typhimurium* (1) |
| Oregon | 0 | 26 | 0 | |
| Tennessee | 2 | 84 | 2 | Newport* (2) |
| Washington | 6 | 76 | 8 | Newport* (4), Heidelberg (1), Typhimurium* (1) |
| West Virginia | 2 | 25 | 8 | Newport* (2) |
| Total | 48 | 1419 | 3 | Newport* (31), Typhimurium* (10), Heidelberg (3), Enteritidis* (1), Montevideo (1), Senftenberg (1), Stanley (1) |

*These isolates were also resistant to ceftriaxone

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

**Table 19. Antimicrobial resistance of non-Typhi *Salmonella* isolates,
1996-2001**

| <i>Salmonella</i> , Non-Typhi | 1996 | 1997 | 1998 | 1999 | 2000 | 2001 |
|---|---------------|---------------|---------------|---------------|---------------|---------------|
| <i>Salmonella</i> isolates | 1326 | 1301 | 1465 | 1498 | 1378 | 1419 |
| Isolates resistant to ≥ 1 antimicrobial agents* | 37% (493) | 34% (443) | 27% (397) | 26% (390) | 26% (353) | 28% (394) |
| Isolates resistant to ≥ 2 antimicrobial agents* | 31% (404) | 25% (328) | 23% (334) | 21% (317) | 21% (284) | 22% (315) |
| Isolates resistant to ≥ 5 antimicrobial agents* | 12% (163) | 14% (180) | 13% (189) | 12% (174) | 12% (161) | 12% (170) |
| Isolates resistant to ≥ 8 antimicrobial agents* | 0.3% (4) | 1% (10) | 1% (14) | 2% (31) | 3% (41) | 3% (40) |
| Serotyped <i>Salmonella</i> isolates | 93% (1231) | 93% (1215) | 96% (1410) | 97% (1459) | 97% (1332) | 99% (1399) |
| Serotyped <i>Salmonella</i> which are Enteritidis | 29% (357) | 25% (301) | 17% (244) | 18% (270) | 24% (319) | 20% (282) |
| S. Enteritidis isolates resistant to ≥ 1 antimicrobial agents* | 31% (110) | 26% (78) | 12% (30) | 17% (45) | 11% (35) | 14% (40) |
| Serotyped <i>Salmonella</i> which are Typhimurium** | 23% (306) | 25% (326) | 26% (377) | 24% (362) | 22% (303) | 24% (325) |
| S. Typhimurium isolates resistant to ≥ 1 antimicrobial agents* | 64% (196) | 62% (202) | 53% (200) | 49% (179) | 50% (153) | 51% (165) |
| S. Typhimurium with at least ACSSuT resistance pattern | 34% (103) | 35% (115) | 32% (120) | 28% (102) | 28% (84) | 29% (96) |
| <i>Salmonella</i> isolates that were at least Typhimurium ACSSuT | 8% (103) | 9% (115) | 8% (120) | 7% (102) | 6% (84) | 7% (96) |
| S. Typhimurium with at least AKSSuT resistance pattern | 9% (27) | 13% (41) | 12% (47) | 11% (39) | 9% (28) | 5% (15) |
| <i>Salmonella</i> isolates that were at least Typhimurium AKSSuT | 2% (27) | 3% (41) | 3% (47) | 3% (39) | 2% (28) | 1% (15) |
| S. Typhimurium with at least ACKSSuT resistance pattern | 4% (13) | 3% (9) | 4% (17) | 3% (12) | 3% (8) | 1% (4) |

*Using only antimicrobial agents (n=15) tested in all six years

**Includes S. Typhimurium and S. Typhimurium variant Copenhagen

***Amox-Clav=Amoxicillin-Clavulanic Acid

****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

TE-test criteria include isolates with a ceftriaxone MIC of $\geq 16\mu\text{g}/\text{ml}$ (1996-1998), by Sensititre®; By-hand broth microdilution criteria include isolates with a ceftiofur MIC of $\geq 4\mu\text{g}/\text{ml}$ and/or ceftriaxone MIC of $\geq 2\mu\text{g}/\text{ml}$ (1999-2001), by Sensititre®

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 19. Summary: Antimicrobial resistance of non-Typhi *Salmonella* isolates, 1996-2001

| <i>Salmonella</i> , Non-Typhi | 1996 | 1997 | 1998 | 1999 | 2000 | 2001 |
|---|--------------|--------------|--------------|--------------|--------------|--------------|
| <i>Salmonella</i> isolates that were at least Typhimurium ACKSSuT | 1% (13) | 1% (9) | 1% (17) | 1% (12) | 1% (8) | 0.3% (4) |
| S. Typhimurium isolates at least ACSSuT, AKSSuT, or ACKSSuT | 42% (130) | 48% (156) | 44% (167) | 39% (141) | 37% (112) | 34% (111) |
| Serotyped <i>Salmonella</i> which are Newport | 4% (51) | 4% (48) | 5% (77) | 7% (98) | 9% (124) | 9% (124) |
| S. Newport isolates resistant to \geq 1 antimicrobial agents* | 18% (9) | 12% (6) | 5% (4) | 23% (23) | 24% (30) | 35% (43) |
| S. Newport with at least MDR-AmpC resistance pattern | 0% (0) | 0% (0) | 1% (1) | 17% (17) | 22% (27) | 25% (31) |
| <i>Salmonella</i> isolates that were at least Newport MDR-AmpC | 0% (0) | 0% (0) | 0.1% (1) | 1% (17) | 2% (27) | 2% (31) |
| Ciprofloxacin (MIC \geq 0.25) | 0.4% (5) | 0.5% (7) | 1% (10) | 1% (15) | 1% (20) | 1% (15) |
| Ciprofloxacin (MIC \geq 4) | 0% (0) | 0% (0) | 0.1% (1) | 0.1% (1) | 0.4% (5) | 0.2% (3) |
| Ceftriaxone (MIC \geq 64)† | 0.1% (1) | 0.4% (5) | 1% (10) | 2% (24) | 2% (25) | 2% (34) |
| Nalidixic Acid (MIC \geq 32) | 0.4% (5) | 1% (11) | 1% (20) | 1% (16) | 2% (34) | 3% (37) |

*Using only antimicrobial agents (n=15) tested in all six years

**Includes S. Typhimurium and S. Typhimurium variant Copenhagen

***Amox-Clav=Amoxicillin-Clavulanic Acid

****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

†E-test criteria include isolates with a ceftriaxone MIC of \geq 16 μ g/ml (1996-1998), by Sensititre®; By-hand broth microdilution criteria include isolates with a ceftiofur MIC of \geq 4 μ g/ml and/or ceftriaxone MIC of \geq 2 μ g/ml (1999-2001), by Sensititre®

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

**Table 19. Summary: Antimicrobial resistance of non-Typhi *Salmonella* isolates,
1996-2001**

| <i>Salmonella</i> , Non-Typhi | 1996 | 1997 | 1998 | 1999 | 2000 | 2001 |
|-------------------------------------|------------|------------|------------|------------|------------|-----------|
| Amikacin (MIC \geq 64) | Not Tested | 0% (0) | 0% (0) | 0.1% (2) | 0% (0) | 0% (0) |
| Amox-Clav*** (MIC \geq 32) | 1% (20) | 1% (19) | 2% (24) | 2% (36) | 4% (54) | 5% (66) |
| Ampicillin (MIC \geq 32) | 21% (279) | 18% (241) | 16% (241) | 16% (234) | 16% (219) | 17% (247) |
| Apramycin (MIC \geq 64) | 0% (0) | 0% (0) | 0% (0) | 0.3% (5) | 0.1% (2) | 0% (0) |
| Cefoxitin (MIC \geq 32) | Not Tested | Not Tested | Not Tested | Not Tested | 3% (43) | 3% (48) |
| Ceftiofur (MIC \geq 8) | 4% (53) | 3% (44) | 1% (14) | 2% (31) | 3% (44) | 4% (58) |
| Cephalothin (MIC \geq 32) | 3% (47) | 3% (43) | 2% (33) | 4% (55) | 4% (54) | 4% (57) |
| Chloramphenicol (MIC \geq 32) | 11% (141) | 10% (131) | 10% (145) | 9% (138) | 10% (138) | 12% (164) |
| Gentamicin (MIC \geq 16) | 5% (64) | 3% (38) | 3% (42) | 2% (34) | 3% (37) | 2% (27) |
| Imipenem (MIC \geq 16) | Not Tested | 0% (0) |
| Kanamycin (MIC \geq 64) | 5% (65) | 5% (66) | 6% (84) | 4% (66) | 6% (77) | 5% (68) |
| Streptomycin (MIC \geq 64) | 21% (275) | 22% (282) | 19% (273) | 17% (253) | 16% (223) | 17% (241) |
| Sulfamethoxazole (MIC \geq 512) | 23% (305) | 25% (328) | 19% (283) | 18% (271) | 17% (235) | 18% (251) |
| Tetracycline (MIC \geq 16) | 24% (321) | 22% (283) | 20% (295) | 19% (291) | 19% (256) | 20% (280) |
| Trimeth-Sulfa**** (MIC \geq 4/76) | 4% (51) | 2% (24) | 2% (34) | 2% (31) | 2% (29) | 2% (28) |

*Using only antimicrobial agents (n=15) tested in all six years

**Includes *S. Typhimurium* and *S. Typhimurium* variant Copenhagen

***Amox-Clav=Amoxicillin-Clavulanic Acid

****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

†E-test criteria include isolates with a ceftriaxone MIC of \geq 16 μ g/ml (1996-1998), by Sensititre®; By-hand broth microdilution criteria include isolates with a ceftiofur MIC of \geq 4 μ g/ml and/or ceftriaxone MIC of \geq 2 μ g/ml (1999-2001), by Sensititre®

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 20. Multidrug* resistance of non-Typhi *Salmonella* isolates, 1996-2001

Non-Typhi *Salmonella*

| Year | # isolates tested | Number resistant to ≥ 1 antimicrobial agents | | Number resistant to ≥ 2 antimicrobial agents | | Number resistant to ≥ 5 antimicrobial agents | | Resistant to at least ACSSuT** | | Resistant to at least AKSSuT** | | Resistant to ceftriaxone (MIC $\geq 64 \mu\text{g/ml}$) [†] | | Decreased susceptibility to ciprofloxacin (MIC $\geq .25 \mu\text{g/ml}$) | |
|------|-------------------|---|----|---|----|---|----|--------------------------------|----|--------------------------------|---|---|-----|--|-----|
| | | N | % | N | % | N | % | N | % | N | % | N | % | N | % |
| 1996 | 1326 | 493 | 37 | 404 | 31 | 163 | 12 | 117 | 9 | 31 | 2 | 1 | 0.1 | 5 | 0.4 |
| 1997 | 1301 | 443 | 34 | 328 | 25 | 180 | 14 | 125 | 10 | 46 | 3 | 5 | 0.4 | 7 | 0.5 |
| 1998 | 1465 | 397 | 27 | 334 | 23 | 189 | 13 | 130 | 9 | 50 | 3 | 10 | 1 | 10 | 1 |
| 1999 | 1498 | 390 | 26 | 317 | 21 | 174 | 12 | 127 | 8 | 44 | 3 | 24 | 2 | 15 | 1 |
| 2000 | 1378 | 353 | 26 | 284 | 21 | 161 | 12 | 122 | 9 | 44 | 3 | 25 | 2 | 20 | 1 |
| 2001 | 1419 | 394 | 28 | 315 | 22 | 170 | 12 | 142 | 10 | 28 | 2 | 34 | 2 | 15 | 1 |

***Salmonella* Enteritidis**

| Year | # isolates tested | Number resistant to ≥ 1 antimicrobial agents | | Number resistant to ≥ 2 antimicrobial agents | | Number resistant to ≥ 5 antimicrobial agents | | Resistant to at least ACSSuT** | | Resistant to at least AKSSuT*** | | Resistant to ceftriaxone (MIC $\geq 64 \mu\text{g/ml}$) ^{†‡} | | Decreased susceptibility to ciprofloxacin (MIC $\geq .25 \mu\text{g/ml}$) | |
|------|-------------------|---|----|---|----|---|---|--------------------------------|-----|---------------------------------|-----|--|-----|--|-----|
| | | N | % | N | % | N | % | N | % | N | % | N | % | N | % |
| 1996 | 357 | 110 | 31 | 84 | 23 | 13 | 4 | 1 | 0.3 | 0 | 0 | 0 | 0 | 3 | 1 |
| 1997 | 301 | 78 | 26 | 37 | 12 | 6 | 2 | 1 | 0.3 | 1 | 0.3 | 0 | 0 | 2 | 1 |
| 1998 | 244 | 30 | 12 | 16 | 7 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 2 | 1 |
| 1999 | 270 | 45 | 17 | 28 | 10 | 2 | 1 | 2 | 1 | 0 | 0 | 1 | 0.4 | 6 | 2 |
| 2000 | 319 | 35 | 11 | 9 | 3 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0.3 |
| 2001 | 282 | 40 | 14 | 16 | 6 | 3 | 1 | 0 | 0 | 1 | 0.3 | 1 | 0.3 | 0 | 0 |

* Using only antimicrobial agents (n=15) tested in all six years

**ACSSuT = Ampicillin, Chloramphenicol, Streptomycin, Sulfamethoxazole, and Tetracycline

***AKSSuT= Ampicillin, Kanamycin, Streptomycin, Sulfamethoxazole, and Tetracycline

****MDR-AmpC=ACSSuT, amoxicillin-clavulanic acid, cephalothin, cefotaxime, ceftiofur, decreased susceptibility to ceftriaxone (MIC $\geq 16 \mu\text{g/ml}$)

†E-test criteria include isolates with a ceftriaxone MIC of $\geq 16 \mu\text{g/ml}$ (1996-1998), by Sensititre®; By-hand broth microdilution criteria include isolates with a ceftiofur MIC of $\geq 4 \mu\text{g/ml}$ and/or ceftriaxone MIC of $\geq 2 \mu\text{g/ml}$ (1999-2001), by Sensititre®

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 20. Multidrug* resistance of non-Typhi *Salmonella* isolates, 1996-2001

***Salmonella* Typhimurium**

| Year | # isolates tested | Number resistant to ≥ 1 antimicrobial agents | | Number resistant to ≥ 2 antimicrobial agents | | Number resistant to ≥ 5 antimicrobial agents | | Resistant to at least ACSSuT** N | Resistant to at least AKSSuT** N | Resistant to ceftriaxone (MIC $\geq 64 \mu\text{g/ml}$) [†] N | Decreased susceptibility to ciprofloxacin (MIC $\geq 25 \mu\text{g/ml}$) N |
|------|-------------------|---|----|---|----|---|----|----------------------------------|----------------------------------|---|---|
| | | N | % | N | % | N | % | | | | |
| 1996 | 306 | 196 | 64 | 177 | 58 | 125 | 41 | 103 | 34 | 27 | 9 |
| 1997 | 326 | 204 | 63 | 188 | 58 | 154 | 47 | 115 | 35 | 41 | 13 |
| 1998 | 377 | 200 | 53 | 193 | 51 | 160 | 42 | 120 | 32 | 47 | 12 |
| 1999 | 362 | 179 | 49 | 167 | 46 | 132 | 36 | 102 | 28 | 39 | 11 |
| 2000 | 303 | 153 | 50 | 143 | 47 | 110 | 36 | 84 | 28 | 28 | 9 |
| 2001 | 325 | 164 | 51 | 155 | 48 | 111 | 34 | 96 | 30 | 14 | 4 |

***Salmonella* Newport**

| Year | # isolates tested | Number resistant to ≥ 1 antimicrobial agents | | Number resistant to ≥ 2 antimicrobial agents | | Number resistant to ≥ 5 antimicrobial agents | | Number resistant to ≥ 8 antimicrobial agents | | Resistant to at least MDR-AmpC**** N | Decreased susceptibility to Ciprofloxacin (MIC $\geq 25 \mu\text{g/ml}$) N |
|------|-------------------|---|----|---|----|---|----|---|----|--------------------------------------|---|
| | | N | % | N | % | N | % | N | % | | |
| 1996 | 51 | 9 | 18 | 4 | 8 | 3 | 6 | 2 | 4 | 0 | 0 |
| 1997 | 48 | 6 | 12 | 3 | 6 | 2 | 4 | 2 | 4 | 0 | 0 |
| 1998 | 77 | 4 | 5 | 2 | 3 | 2 | 3 | 2 | 3 | 1 | 1 |
| 1999 | 98 | 23 | 23 | 17 | 17 | 17 | 17 | 17 | 17 | 17 | 0 |
| 2000 | 124 | 30 | 24 | 28 | 23 | 28 | 23 | 27 | 22 | 27 | 22 |
| 2001 | 124 | 43 | 35 | 39 | 31 | 33 | 27 | 31 | 25 | 31 | 25 |

* Using only antimicrobial agents (n=15) tested in all six years

**ACSSuT = Ampicillin, Chloramphenicol, Streptomycin, Sulfamethoxazole, and Tetracycline

***AKSSuT= Ampicillin, Kanamycin, Streptomycin, Sulfamethoxazole, and Tetracycline

****MDR-AmpC=ACSSuT, amoxicillin-clavulanic acid, cephalothin, cefoxitin, ceftiofur, decreased susceptibility to ceftriaxone (MIC $\geq 16 \mu\text{g/ml}$)

†E-test criteria include isolates with a ceftriaxone MIC of $>=16 \mu\text{g/ml}$ (1996-1998), by Sensititre®; By-hand broth microdilution criteria include isolates with a ceftiofur MIC of $>=4 \mu\text{g/ml}$ and/or ceftriaxone MIC of $>=2 \mu\text{g/ml}$ (1999-2001), by Sensititre®

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

**Table 21. Antimicrobial resistance of *Salmonella* Typhi isolates,
1999-2001**

| <i>Salmonella</i> Typhi | 1999 | 2000 | 2001 |
|---|---------------|-------------|-------------|
| <i>Salmonella</i> Typhi isolates | 166 | 177 | 197 |
| Isolates resistant to \geq 1 antimicrobial agents* | 29% (49) | 28% (50) | 41% (81) |
| Isolates resistant to \geq 2 antimicrobial agents* | 15% (25) | 12% (21) | 23% (45) |
| <i>Salmonella</i> Typhi with at least ACSSuT resistance pattern | 9% (15) | 8% (14) | 17% (33) |
| <i>Salmonella</i> Typhi with at least ACSuTm** resistance pattern | 12% (20) | 9% (16) | 18% (35) |
| Amikacin (MIC \geq 64) | 0% (0) | 1% (2) | 0% (0) |
| Amox-Clav*** (MIC \geq 32) | 1% (1) | 0% (0) | 0% (0) |
| Ampicillin (MIC $>$ 32) | 13% (21) | 9% (16) | 20% (40) |
| Apramycin (MIC $>$ 64) | 0% (0) | 1% (2) | 0% (0) |
| Cefoxitin (MIC \geq 32) | Not Tested | 2% (3) | 0.5% (1) |
| Ceftiofur (MIC \geq 8) | 1% (2) | 1% (1) | 0% (0) |
| Ceftriaxone (MIC $>$ 64) | 0% (0) | 0% (0) | 0% (0) |

*Using only antimicrobial agents (n=15) tested in all three years

**ACSuTm=Ampicillin, Chloramphenicol, Sulfamethoxazole-Trimethoprim

***Amox-Clav=Amoxicillin-Clavulanic Acid

****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 21. Summary: Antimicrobial resistance of *Salmonella* Typhi isolates, 1999-2001

| <i>Salmonella</i> Typhi | 1999 | 2000 | 2001 |
|-----------------------------------|---------------|---------------|-------------|
| Cephalothin (MIC \geq 32) | 2% (4) | 1% (2) | 0.5% (1) |
| Chloramphenicol (MIC \geq 32) | 12% (20) | 11% (19) | 21% (41) |
| Ciprofloxacin (MIC \geq 0.25) | 15% (25) | 21% (38) | 20% (39) |
| Ciprofloxacin (MIC $>$ 4) | 0% (0) | 0% (0) | 0% (0) |
| Gentamicin (MIC $>$ 16) | 0% (0) | 1% (1) | 0% (0) |
| Imipenem (MIC \geq 16) | Not Tested | Not Tested | 0% (0) |
| Kanamycin (MIC \geq 64) | 0% (0) | 1% (1) | 0.5% (1) |
| Nalidixic Acid (MIC $>$ 32) | 19% (31) | 23% (41) | 30% (59) |
| Streptomycin (MIC \geq 64) | 14% (23) | 10% (18) | 20% (40) |
| Sulfamethoxazole (MIC \geq 512) | 17% (28) | 12% (21) | 21% (41) |
| Tetracycline (MIC $>$ 16) | 9% (15) | 11% (19) | 21% (41) |
| Trimeth-Sulfa**** (MIC $>$ 4/76) | 13% (21) | 9% (16) | 21% (41) |

*Using only antimicrobial agents (n=15) tested in all three years

**ACSuTm=Ampicillin, Chloramphenicol, Sulfamethoxazole-Trimethoprim

***Amox-Clav=Amoxicillin-Clavulanic Acid

****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 22. Antimicrobial resistance of *E. coli* O157 isolates, 1996-2001

| | 1996 | 1997 | 1998 | 1999 | 2000 | 2001 |
|--|---------------|---------------|---------------|---------------|---------------|-------------|
| <i>E. coli</i> O157 isolates | 201 | 161 | 313 | 292 | 407 | 277 |
| Isolates resistant to \geq 1 antimicrobial agents* | 21% (42) | 12% (20) | 7% (23) | 10% (30) | 10% (40) | 9% (24) |
| Isolates resistant to \geq 2 antimicrobial agents* | 8% (15) | 7% (11) | 5% (17) | 4% (12) | 7% (27) | 5% (15) |
| Amikacin (MIC \geq 64) | Not Tested | 0% (0) | 0% (0) | 0% (0) | 0% (0) | 0% (0) |
| Amox-Clav** (MIC \geq 32) | | 0% (0) | 0% (0) | 0% (0) | 0.3% (1) | 1% (4) |
| Ampicillin (MIC \geq 32) | | 1% (3) | 0% (0) | 3% (8) | 1% (4) | 3% (11) |
| Apramycin (MIC \geq 64) | | 0% (0) | 0% (0) | 0% (0) | 0% (0) | 0% (0) |
| Cefoxitin (MIC \geq 32) | Not Tested | Not Tested | Not Tested | Not Tested | 1% (4) | 1% (2) |
| Ceftiofur (MIC \geq 8) | | 5% (10) | 0% (0) | 0% (0) | 1% (4) | 1% (3) |
| Ceftriaxone (MIC \geq 64) | | 0% (0) | 0% (0) | 0% (0) | 0% (0) | 0% (0) |
| Cephalothin (MIC \geq 32) | | 3% (6) | 4% (6) | 0% (0) | 1% (2) | 1% (5) |
| Chloramphenicol (MIC \geq 32) | | 0.5% (1) | 0% (0) | 0.3% (1) | 0% (0) | 4% (15) |
| Ciprofloxacin (MIC \geq 4) | | 0% (0) | 0% (0) | 0% (0) | 0% (0) | 0% (0) |
| Gentamicin (MIC \geq 16) | | 0% (0) | 0% (0) | 0% (0) | 0.3% (1) | 0.5% (2) |
| Imipenem (MIC \geq 16) | Not Tested | Not Tested | Not Tested | Not Tested | Not Tested | 0% (0) |
| Kanamycin (MIC \geq 64) | | 0% (0) | 0% (0) | 0.3% (1) | 1% (2) | 1% (4) |
| Nalidixic Acid (MIC \geq 32) | | 0% (0) | 0% (0) | 0% (0) | 1% (2) | 0.5% (2) |
| Streptomycin (MIC \geq 64) | | 2% (4) | 2% (4) | 2% (6) | 3% (8) | 5% (21) |
| Sulfamethoxazole (MIC \geq 512) | | 14% (28) | 11% (17) | 6% (18) | 8% (24) | 6% (24) |
| Tetracycline (MIC \geq 16) | | 5% (10) | 3% (5) | 4% (14) | 3% (10) | 7% (29) |
| Trimeth-Sulfa*** (MIC \geq 4/76) | | 0% (0) | 0% (0) | 1% (2) | 1% (4) | 1% (3) |
| | | | | | | 1% (2) |

*Using only antimicrobial agents (n=15) tested in all six years

**Amox-Clav=Amoxicillin-Clavulanic Acid

***Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 23. Frequency of *Shigella* species, 2001

| Species | N | % |
|-----------------------------|------------|------------|
| <i>sonnei</i> | 239 | 70 |
| <i>flexneri</i> | 91 | 26 |
| <i>boydii</i> | 5 | 1 |
| <i>dysenteriae</i> | 3 | 1 |
| provisional <i>Shigella</i> | 6 | 2 |
| Total | 344 | 100 |

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 24. Antimicrobial resistance of *Shigella* isolates, by *Shigella* species, 2001

| Antimicrobial Agent | All <i>Shigella</i> (N=344) | | <i>Shigella sonnei</i> (N=239) | | <i>Shigella flexneri</i> (N=91) | | <i>Shigella boydii</i> (N=5) | |
|----------------------|--------------------------------|-------------|-----------------------------------|-------------|------------------------------------|-------------|---------------------------------|-------------|
| | # Resistant | % Resistant | # Resistant | % Resistant | # Resistant | % Resistant | # Resistant | % Resistant |
| Amikacin | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Amox-Clav* | 15 | 4 | 11 | 5 | 4 | 4 | 0 | 0 |
| Ampicillin | 274 | 80 | 198 | 83 | 66 | 72 | 1 | 20 |
| Apramycin | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Cefoxitin | 4 | 1 | 4 | 2 | 0 | 0 | 0 | 0 |
| Ceftiofur | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Ceftriaxone | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Cephalothin | 31 | 9 | 30 | 12 | 1 | 1 | 0 | 0 |
| Chloramphenicol | 74 | 21 | 3 | 1 | 68 | 75 | 0 | 0 |
| Ciprofloxacin | 1 | 0.3 | 0 | 0 | 1 | 1 | 0 | 0 |
| Gentamicin | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Imipenem | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Kanamycin | 2 | 1 | 1 | 0.4 | 1 | 1 | 0 | 0 |
| Nalidixic Acid | 6 | 2 | 2 | 1 | 3 | 3 | 1 | 20 |
| Streptomycin | 183 | 53 | 129 | 54 | 43 | 47 | 3 | 60 |
| Sulfamethoxazole | 194 | 56 | 130 | 54 | 52 | 57 | 4 | 80 |
| Tetracycline | 204 | 59 | 107 | 45 | 86 | 94 | 2 | 40 |
| Trimethoprim-Sulfa** | 161 | 47 | 121 | 51 | 31 | 34 | 2 | 40 |

*Amox-Clav=Amoxicillin-Clavulanic Acid

**Trimethoprim-Sulfa=Trimethoprim-Sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

**Table 25. Antimicrobial resistance of *Shigella* isolates,
1999-2001**

| | 1999 | 2000 | 2001 |
|---|---------------|--------------|--------------|
| <i>Shigella</i> isolates | 375 | 451 | 344 |
| Isolates resistant to \geq 1 antimicrobial agents* | 91% (341) | 93% (418) | 95% (327) |
| Isolates resistant to \geq 2 antimicrobial agents* | 65% (245) | 67% (302) | 71% (244) |
| <i>Shigella</i> with at least ACSSuT resistance pattern | 8% (32) | 5% (25) | 6% (22) |
| <i>Shigella</i> with at least ACSuTm** resistance pattern | 10% (37) | 7% (31) | 7% (24) |
| Amikacin (MIC \geq 64) | 0% (0) | 0.2% (1) | 0% (0) |
| Amox-Clav*** (MIC \geq 32) | 1% (4) | 2% (10) | 4% (15) |
| Ampicillin (MIC \geq 32) | 78% (291) | 79% (356) | 80% (274) |
| Apramycin (MIC \geq 64) | 0% (0) | 0.2% (1) | 0% (0) |
| Cefoxitin (MIC \geq 32) | Not Tested | 0.4% (2) | 1% (4) |
| Ceftiofur (MIC \geq 8) | 0% (0) | 0% (0) | 0% (0) |
| Ceftriaxone (MIC \geq 64) | 0% (0) | 0% (0) | 0% (0) |

*Using only antimicrobial agents (n=15) tested in all three years

**ACSuTm=Ampicillin, Chloramphenicol, Sulfamethoxazole-Trimethoprim

***Amox-Clav=Amoxicillin-Clavulanic Acid

****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

**Table 25. Summary: Antimicrobial resistance of *Shigella* isolates,
1999-2001**

| | 1999 | 2000 | 2001 |
|-------------------------------------|---------------|---------------|--------------|
| Cephalothin (MIC \geq 32) | 3% (12) | 8% (36) | 9% (31) |
| Chloramphenicol (MIC \geq 32) | 17% (65) | 14% (63) | 21% (74) |
| Ciprofloxacin (MIC \geq 0.25) | 1% (3) | 0.2% (1) | 0.3% (1) |
| Ciprofloxacin (MIC \geq 4) | 0% (0) | 0% (0) | 0.3% (1) |
| Gentamicin (MIC \geq 16) | 0.3% (1) | 0.2% (1) | 0% (0) |
| Imipenem (MIC \geq 16) | Not Tested | Not Tested | 0% (0) |
| Kanamycin (MIC \geq 64) | 0.5% (2) | 1% (6) | 1% (2) |
| Nalidixic Acid (MIC \geq 32) | 2% (6) | 1% (5) | 2% (6) |
| Streptomycin (MIC \geq 64) | 56% (209) | 57% (258) | 53% (183) |
| Sulfamethoxazole (MIC \geq 512) | 56% (210) | 56% (252) | 56% (194) |
| Tetracycline (MIC \geq 16) | 57% (215) | 45% (202) | 59% (204) |
| Trimeth-Sulfa**** (MIC \geq 4/76) | 51% (193) | 53% (239) | 47% (161) |

*Using only antimicrobial agents (n=15) tested in all three years

**ACSuTm=Ampicillin, Chloramphenicol, Sulfamethoxazole-Trimethoprim

***Amox-Clav=Amoxicillin-Clavulanic Acid

****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

**Table 26. Antimicrobial resistance of *Shigella sonnei* isolates,
1999-2001**

| | 1999 | 2000 | 2001 |
|--|---------------|--------------|--------------|
| <i>Shigella sonnei</i> isolates | 275 | 367 | 239 |
| Isolates resistant to ≥ 1 antimicrobial agents* | 89% (246) | 92% (339) | 95% (226) |
| Isolates resistant to ≥ 2 antimicrobial agents* | 58% (160) | 64% (233) | 62% (149) |
| <i>Shigella sonnei</i> with at least ACSSuT resistance pattern | 0.4% (1) | 1% (3) | 0% (0) |
| <i>Shigella sonnei</i> with at least ACSuTm** resistance pattern | 2% (5) | 2% (7) | 1% (2) |
| Amikacin (MIC ≥ 64) | 0% (0) | 0.3% (1) | 0% (0) |
| Amox-Clav*** (MIC ≥ 32) | 0.4% (1) | 2% (7) | 5% (11) |
| Ampicillin (MIC ≥ 32) | 80% (219) | 80% (295) | 83% (198) |
| Apramycin (MIC > 64) | 0% (0) | 0.3% (1) | 0% (0) |
| Cefoxitin (MIC ≥ 32) | Not Tested | 0.5% (2) | 2% (4) |
| Ceftiofur (MIC ≥ 8) | 0% (0) | 0% (0) | 0% (0) |
| Ceftriaxone (MIC ≥ 64) | 0% (0) | 0% (0) | 0% (0) |

*Using only antimicrobial agents (n=15) tested in all three years

**ACSuTm=Ampicillin, Chloramphenicol, Sulfamethoxazole-Trimethoprim

***Amox-Clav=Amoxicillin-Clavulanic Acid

****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 26. Summary: Antimicrobial resistance of *Shigella sonnei* isolates, 1999-2001

| | 1999 | 2000 | 2001 |
|-------------------------------------|---------------|---------------|--------------|
| Cephalothin (MIC \geq 32) | 3% (8) | 9% (32) | 12% (30) |
| Chloramphenicol (MIC \geq 32) | 2% (5) | 3% (10) | 1% (3) |
| Ciprofloxacin (MIC \geq 0.25) | 1% (2) | 0.3% (1) | 0% (0) |
| Ciprofloxacin (MIC \geq 4) | 0% (0) | 0% (0) | 0% (0) |
| Gentamicin (MIC \geq 16) | 0.4% (1) | 0.3% (1) | 0% (0) |
| Imipenem (MIC \geq 16) | Not Tested | Not Tested | 0% (0) |
| Kanamycin (MIC \geq 64) | 1% (2) | 2% (6) | 0.4% (1) |
| Nalidixic Acid (MIC \geq 32) | 1% (4) | 1% (5) | 1% (2) |
| Streptomycin (MIC \geq 64) | 52% (143) | 56% (206) | 54% (129) |
| Sulfamethoxazole (MIC \geq 512) | 54% (150) | 56% (206) | 54% (130) |
| Tetracycline (MIC \geq 16) | 46% (127) | 34% (126) | 45% (107) |
| Trimeth-Sulfa**** (MIC \geq 4/76) | 53% (146) | 55% (202) | 51% (121) |

*Using only antimicrobial agents (n=15) tested in all three years

**ACSuTm=Ampicillin, Chloramphenicol, Sulfamethoxazole-Trimethoprim

***Amox-Clav=Amoxicillin-Clavulanic Acid

****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

**Table 27. Antimicrobial resistance of *Shigella flexneri* isolates,
1999-2001**

| | 1999 | 2000 | 2001 |
|--|---------------|-------------|-------------|
| <i>Shigella flexneri</i> isolates | 87 | 75 | 91 |
| Isolates resistant to ≥ 1 antimicrobial agents* | 95% (83) | 96% (72) | 97% (88) |
| Isolates resistant to ≥ 2 antimicrobial agents* | 84% (73) | 83% (62) | 90% (82) |
| <i>Shigella flexneri</i> with at least ACSSuT resistance pattern | 33% (29) | 29% (22) | 22% (20) |
| <i>Shigella flexneri</i> with at least ACSuTm** resistance pattern | 34% (30) | 32% (24) | 23% (21) |
| Amikacin (MIC ≥ 64) | 0% (0) | 0% (0) | 0% (0) |
| Amox-Clav*** (MIC ≥ 32) | 3% (3) | 4% (3) | 4% (4) |
| Ampicillin (MIC ≥ 32) | 77% (67) | 77% (58) | 72% (66) |
| Apramycin (MIC > 64) | 0% (0) | 0% (0) | 0% (0) |
| Cefoxitin (MIC ≥ 32) | Not Tested | 0% (0) | 0% (0) |
| Ceftiofur (MIC ≥ 8) | 0% (0) | 0% (0) | 0% (0) |
| Ceftriaxone (MIC ≥ 64) | 0% (0) | 0% (0) | 0% (0) |

*Using only antimicrobial agents (n=15) tested in all three years

**ACSuTm=Ampicillin, Chloramphenicol, Sulfamethoxazole-Trimethoprim

***Amox-Clav=Amoxicillin-Clavulanic Acid

****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 27. Summary: Antimicrobial resistance of *Shigella flexneri* isolates, 1999-2001

| | 1999 | 2000 | 2001 |
|-------------------------------------|---------------|---------------|-------------|
| Cephalothin (MIC \geq 32) | 5% (4) | 3% (2) | 1% (1) |
| Chloramphenicol (MIC \geq 32) | 64% (56) | 69% (52) | 75% (68) |
| Ciprofloxacin (MIC \geq 0.25) | 1% (1) | 0% (0) | 1% (1) |
| Ciprofloxacin (MIC \geq 4) | 0% (0) | 0% (0) | 1% (1) |
| Gentamicin (MIC \geq 16) | 0% (0) | 0% (0) | 0% (0) |
| Imipenem (MIC \geq 16) | Not Tested | Not Tested | 0% (0) |
| Kanamycin (MIC \geq 64) | 0% (0) | 0% (0) | 1% (1) |
| Nalidixic Acid (MIC \geq 32) | 1% (1) | 0% (0) | 3% (3) |
| Streptomycin (MIC \geq 64) | 63% (55) | 61% (46) | 47% (43) |
| Sulfamethoxazole (MIC \geq 512) | 59% (51) | 53% (40) | 57% (52) |
| Tetracycline (MIC \geq 16) | 92% (80) | 92% (69) | 94% (86) |
| Trimeth-Sulfa**** (MIC \geq 4/76) | 48% (42) | 43% (32) | 34% (31) |

*Using only antimicrobial agents (n=15) tested in all three years

**ACSuTm=Ampicillin, Chloramphenicol, Sulfamethoxazole-Trimethoprim

***Amox-Clav=Amoxicillin-Clavulanic Acid

****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 28. Frequency of *Campylobacter* species, 2001

| Species | N | % |
|----------------|------------|------------|
| <i>jejuni</i> | 365 | 95 |
| <i>coli</i> | 17 | 4 |
| <i>fetus</i> | 1 | 0.3 |
| Other species | 1 | 0.3 |
| Total | 384 | 100 |

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

**Table 29. Antimicrobial resistance of *Campylobacter* isolates,
by *Campylobacter* species, 2001**

| Antimicrobial Agent | All <i>Campylobacter</i> (N=384) | | <i>Campylobacter jejuni</i> (N=365) | | <i>Campylobacter coli</i> (N=17) | |
|----------------------------|---|-------------|--|-------------|---|-------------|
| | # Resistant | % Resistant | # Resistant | % Resistant | # Resistant | % Resistant |
| Azithromycin | 8 | 2 | 7 | 2 | 1 | 6 |
| Chloramphenicol | 0 | 0 | 0 | 0 | 0 | 0 |
| Ciprofloxacin | 75 | 19 | 67 | 18 | 8 | 47 |
| Clindamycin | 8 | 2 | 7 | 2 | 1 | 6 |
| Erythromycin | 8 | 2 | 7 | 2 | 1 | 6 |
| Gentamicin | 0 | 0 | 0 | 0 | 0 | 0 |
| Nalidixic Acid | 77 | 20 | 69 | 19 | 7 | 41 |
| Tetracycline | 156 | 41 | 146 | 40 | 10 | 59 |

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 30. Antimicrobial resistance of *Campylobacter jejuni* isolates, by site, 2001

| Site | Antimicrobial Agent | <i>Campylobacter jejuni</i> (N=365) | |
|-------------------------------|---------------------|-------------------------------------|--------------------------|
| California (N=46) | | #Resistant | % Resistant within state |
| | Azithromycin | 0 | 0 |
| | Chloramphenicol | 0 | 0 |
| | Ciprofloxacin | 11 | 24 |
| | Clindamycin | 0 | 0 |
| | Erythromycin | 0 | 0 |
| | Gentamicin | 0 | 0 |
| | Nalidixic Acid | 11 | 24 |
| Colorado (N=40) | Tetracycline | 17 | 37 |
| | Azithromycin | 0 | 0 |
| | Chloramphenicol | 0 | 0 |
| | Ciprofloxacin | 4 | 10 |
| | Clindamycin | 0 | 0 |
| | Erythromycin | 0 | 0 |
| | Gentamicin | 0 | 0 |
| | Nalidixic Acid | 4 | 10 |
| Connecticut (N=43) | Tetracycline | 13 | 32 |
| | Azithromycin | 0 | 0 |
| | Chloramphenicol | 0 | 0 |
| | Ciprofloxacin | 12 | 28 |
| | Clindamycin | 0 | 0 |
| | Erythromycin | 0 | 0 |
| | Gentamicin | 0 | 0 |
| | Nalidixic Acid | 12 | 28 |
| | Tetracycline | 27 | 63 |

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 30. Antimicrobial resistance of *Campylobacter jejuni* isolates, by site, 2001

| Site | Antimicrobial Agent | <i>Campylobacter jejuni</i> (N=365) | |
|---------------------|---------------------|-------------------------------------|--------------------------|
| | | #Resistant | % Resistant within state |
| Georgia (N=52) | Azithromycin | 2 | 4 |
| | Chloramphenicol | 0 | 0 |
| | Ciprofloxacin | 14 | 27 |
| | Clindamycin | 2 | 4 |
| | Erythromycin | 2 | 4 |
| | Gentamicin | 0 | 0 |
| | Nalidixic Acid | 14 | 27 |
| | Tetracycline | 18 | 35 |
| Maryland (N=36) | Azithromycin | 0 | 0 |
| | Chloramphenicol | 0 | 0 |
| | Ciprofloxacin | 7 | 19 |
| | Clindamycin | 0 | 0 |
| | Erythromycin | 0 | 0 |
| | Gentamicin | 0 | 0 |
| | Nalidixic Acid | 7 | 19 |
| | Tetracycline | 9 | 25 |
| Minnesota (N=47) | Azithromycin | 2 | 4 |
| | Chloramphenicol | 0 | 0 |
| | Ciprofloxacin | 7 | 15 |
| | Clindamycin | 2 | 4 |
| | Erythromycin | 2 | 4 |
| | Gentamicin | 0 | 0 |
| | Nalidixic Acid | 8 | 17 |
| | Tetracycline | 27 | 57 |

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 30. Antimicrobial resistance of *Campylobacter jejuni* isolates, by site, 2001

| Site | Antimicrobial Agent | <i>Campylobacter jejuni</i> (N=365) | |
|----------------------------------|----------------------------|--|--------------------------|
| New York State (N=44) | | #Resistant | % Resistant within state |
| | Azithromycin | 1 | 2 |
| | Chloramphenicol | 0 | 0 |
| | Ciprofloxacin | 5 | 11 |
| | Clindamycin | 1 | 2 |
| | Erythromycin | 1 | 2 |
| | Gentamicin | 0 | 0 |
| | Nalidixic Acid | 5 | 11 |
| Oregon (N=26) | Tetracycline | 18 | 41 |
| | Azithromycin | 0 | 0 |
| | Chloramphenicol | 0 | 0 |
| | Ciprofloxacin | 2 | 8 |
| | Clindamycin | 0 | 0 |
| | Erythromycin | 0 | 0 |
| | Gentamicin | 0 | 0 |
| | Nalidixic Acid | 3 | 11 |
| Tennessee (N=31) | Tetracycline | 6 | 23 |
| | Azithromycin | 2 | 6 |
| | Chloramphenicol | 0 | 0 |
| | Ciprofloxacin | 5 | 16 |
| | Clindamycin | 2 | 6 |
| | Erythromycin | 2 | 6 |
| | Gentamicin | 0 | 0 |
| | Nalidixic Acid | 5 | 16 |
| | Tetracycline | 11 | 35 |

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

**Table 31. Antimicrobial resistance of *Campylobacter* isolates,
1997-2001**

| <i>Campylobacter</i> | 1997 | 1998 | 1999 | 2000 | 2001 |
|--|---------------|--------------|--------------|--------------|--------------|
| <i>Campylobacter</i> isolates | 217 | 310 | 318 | 324 | 384 |
| Isolates resistant to ≥ 1 antimicrobial agents* | 52% (113) | 55% (170) | 53% (169) | 48% (155) | 50% (193) |
| Isolates resistant to ≥ 2 antimicrobial agents* | 16% (34) | 18% (57) | 21% (67) | 16% (51) | 21% (81) |
| Ciprofloxacin (MIC ≥ 4) | 13% (28) | 13% (42) | 18% (58) | 14% (46) | 19% (75) |
| Nalidixic Acid (MIC ≥ 32) | 14% (31) | 17% (54) | 21% (67) | 17% (54) | 20% (77) |
| Erythromycin (MIC ≥ 8) | 2% (4) | 3% (8) | 2% (8) | 1% (5) | 2% (8) |
| Azithromycin (MIC ≥ 2) | Not Tested | 2% (5) | 3% (10) | 2% (7) | 2% (8) |
| Chloramphenicol (MIC ≥ 32) | 1% (3) | 2% (6) | 0.3% (1) | 0% (0) | 0% (0) |
| Clindamycin (MIC ≥ 4) | 2% (4) | 1% (4) | 2% (5) | 1% (4) | 2% (8) |
| Gentamicin (MIC ≥ 16) | Not Tested | 0% (0) | 0% (0) | 0.3% (1) | 0% (0) |
| Tetracycline (MIC ≥ 16) | 47% (102) | 45% (141) | 44% (140) | 38% (122) | 41% (156) |

*Using only *Campylobacter* antimicrobial agents (n=6) tested in all five years

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

**Table 32. Antimicrobial resistance of *Campylobacter jejuni* isolates,
1997-2001**

| <i>Campylobacter jejuni</i> | 1997 | 1998 | 1999 | 2000 | 2001 |
|--|---------------|--------------|--------------|--------------|--------------|
| <i>Campylobacter jejuni</i> isolates | 209 | 297 | 294 | 306 | 365 |
| Isolates resistant to ≥ 1 antimicrobial agents* | 51% (107) | 54% (162) | 53% (157) | 49% (149) | 50% (181) |
| Isolates resistant to ≥ 2 antimicrobial agents* | 14% (30) | 17% (50) | 19% (57) | 15% (45) | 20% (73) |
| Ciprofloxacin (MIC > 4) | 12% (26) | 14% (41) | 18% (52) | 14% (43) | 18% (67) |
| Nalidixic Acid (MIC ≥ 32) | 13% (28) | 16% (47) | 20% (59) | 16% (49) | 19% (69) |
| Erythromycin (MIC > 8) | 1% (3) | 2% (7) | 2% (6) | 1% (4) | 2% (7) |
| Azithromycin (MIC ≥ 2) | Not Tested | 1% (4) | 3% (8) | 2% (6) | 2% (7) |
| Chloramphenicol (MIC ≥ 32) | 1% (2) | 1% (2) | 0.3% (1) | 0% (0) | 0% (0) |
| Clindamycin (MIC ≥ 4) | 1% (2) | 1% (3) | 1% (3) | 1% (3) | 2% (7) |
| Gentamicin (MIC ≥ 16) | Not Tested | 0% (0) | 0% (0) | 0% (0) | 0% (0) |
| Tetracycline (MIC ≥ 16) | 47% (98) | 46% (137) | 46% (134) | 39% (118) | 40% (146) |

*Using only *Campylobacter* antimicrobial agents (n=6) tested in all five years

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

**Table 33. Antimicrobial resistance of *Campylobacter coli* isolates,
1997-2001**

| <i>Campylobacter coli</i> | 1997 | 1998 | 1999 | 2000 | 2001 |
|--|---------------|------------|-------------|------------|-------------|
| <i>Campylobacter coli</i> isolates | 6 | 8 | 20 | 12 | 17 |
| Isolates resistant to ≥ 1 antimicrobial agents* | 83% (5) | 50% (4) | 50% (10) | 33% (4) | 65% (11) |
| Isolates resistant to ≥ 2 antimicrobial agents* | 50% (3) | 50% (4) | 35% (7) | 25% (3) | 47% (8) |
| Ciprofloxacin (MIC ≥ 4) | 33% (2) | 0% (0) | 30% (6) | 25% (3) | 47% (8) |
| Nalidixic Acid (MIC ≥ 32) | 50% (3) | 50% (4) | 30% (6) | 25% (3) | 41% (7) |
| Erythromycin (MIC ≥ 8) | 0% (0) | 12% (1) | 10% (2) | 8% (1) | 6% (1) |
| Azithromycin (MIC > 2) | Not Tested | 12% (1) | 10% (2) | 8% (1) | 6% (1) |
| Chloramphenicol (MIC > 32) | 17% (1) | 25% (2) | 0% (0) | 0% (0) | 0% (0) |
| Clindamycin (MIC ≥ 4) | 17% (1) | 12% (1) | 10% (2) | 8% (1) | 6% (1) |
| Gentamicin (MIC ≥ 16) | Not Tested | 0% (0) | 0% (0) | 8% (1) | 0% (0) |
| Tetracycline (MIC ≥ 16) | 67% (4) | 50% (4) | 30% (6) | 25% (3) | 59% (10) |

*Using only *Campylobacter* antimicrobial agents (n=6) tested in all five years

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Table 34. Frequency of resistance and multidrug* resistance among *Salmonella* Typhi, *Shigella*, *E. coli* O157, and *Campylobacter* isolates, 2001

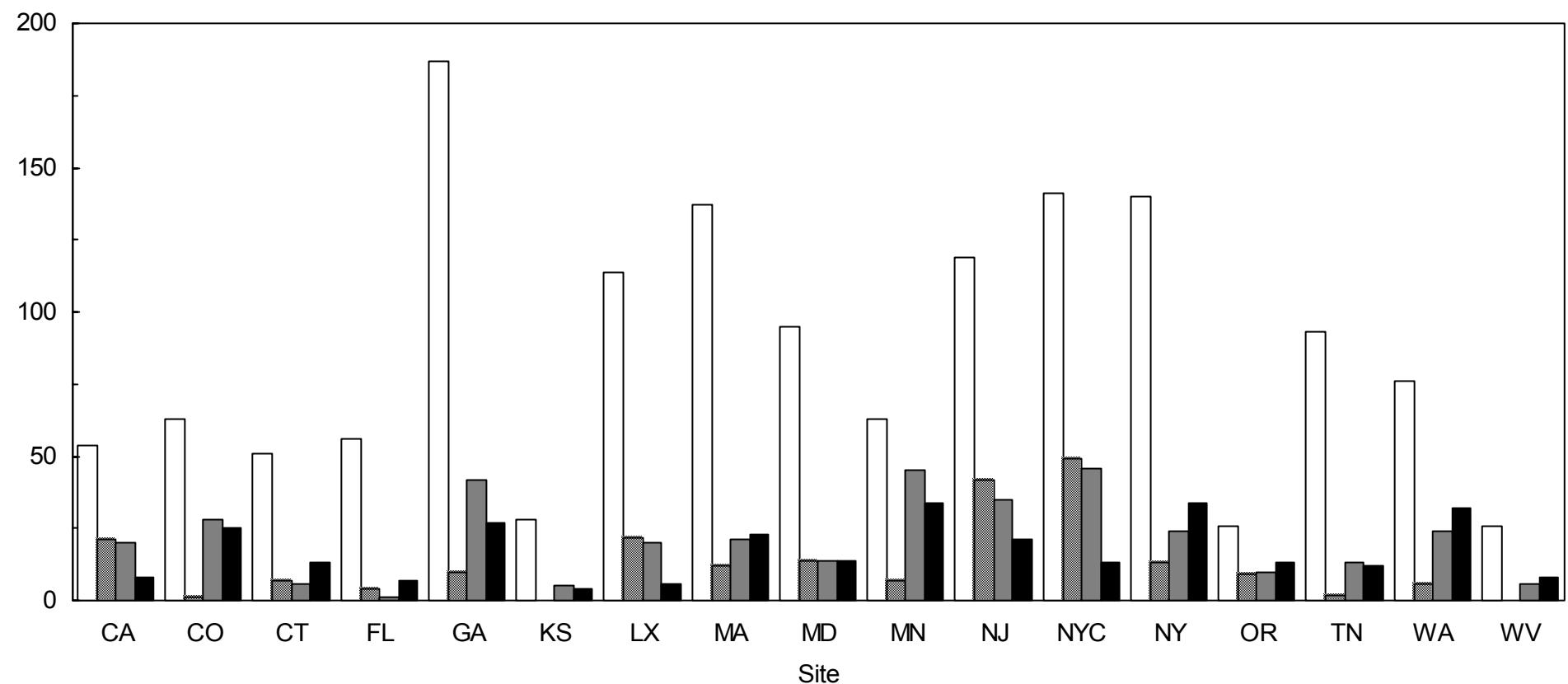
| Isolate | Total | | Number resistant to ≥ 1 antimicrobial agents | | Number resistant to ≥ 2 antimicrobial agents | | Number resistant to ≥ 5 antimicrobial agents | | Number resistant to ≥ 8 antimicrobial agents | |
|-------------------------|-------|-----|--|----|--|----|--|----|--|-----|
| | N | % | N | % | N | % | N | % | N | % |
| <i>Salmonella</i> Typhi | 197 | 100 | 81 | 41 | 46 | 23 | 38 | 19 | 0 | 0 |
| <i>Shigella</i> | 344 | 100 | 327 | 95 | 244 | 71 | 125 | 36 | 1 | 0.3 |
| <i>sonnei</i> | 239 | 69 | 226 | 95 | 149 | 62 | 86 | 36 | 0 | 0 |
| <i>flexneri</i> | 91 | 26 | 88 | 97 | 82 | 90 | 30 | 33 | 1 | 1 |
| <i>E. coli</i> O157 | 277 | 100 | 24 | 9 | 15 | 5 | 4 | 1 | 1 | 0.4 |
| <i>Campylobacter</i> | 384 | 100 | 193 | 50 | 81 | 21 | 3 | 1 | 0 | 0 |
| <i>jejuni</i> | 365 | 95 | 181 | 50 | 73 | 20 | 3 | 1 | 0 | 0 |
| <i>coli</i> | 17 | 4 | 11 | 65 | 8 | 47 | 0 | 0 | 0 | 0 |

*Multidrug resistance for *Salmonella*, *Shigella*, *E. coli* O157 to 18 antimicrobial agents tested in 2001: amikacin, amoxicillin-clavulanic acid, ampicillin, apramycin, cefoxitin, ceftiofur, ceftriaxone, cephalothin, chloramphenicol, ciprofloxacin, gentamicin, imipenem, kanamycin, nalidixic acid, streptomycin, sulfamethoxazole, tetracycline, trimethoprim-sulfamethoxazole; Multidrug resistance for *Campylobacter* to 8 antimicrobial agents tested in 2001: azithromycin, chloramphenicol, ciprofloxacin, clindamycin, erythromycin, gentamicin, naladixic acid, tetracycline

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

Figure 1. Number of isolates submitted, by site, 2001

Number of Isolates



□ Non-Typhi *Salmonella* (N=1469) ■ *Salmonella Typhi* (N=219) ■ *Shigella* (N=360) ■ *E. coli* O157 (N=294)

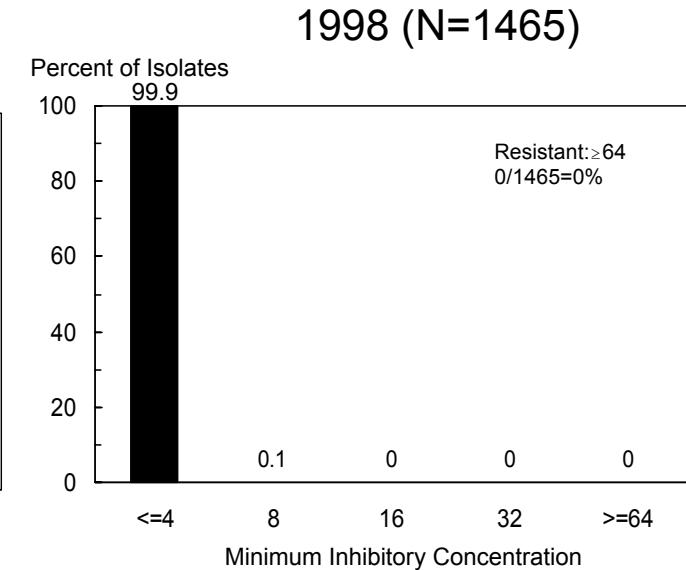
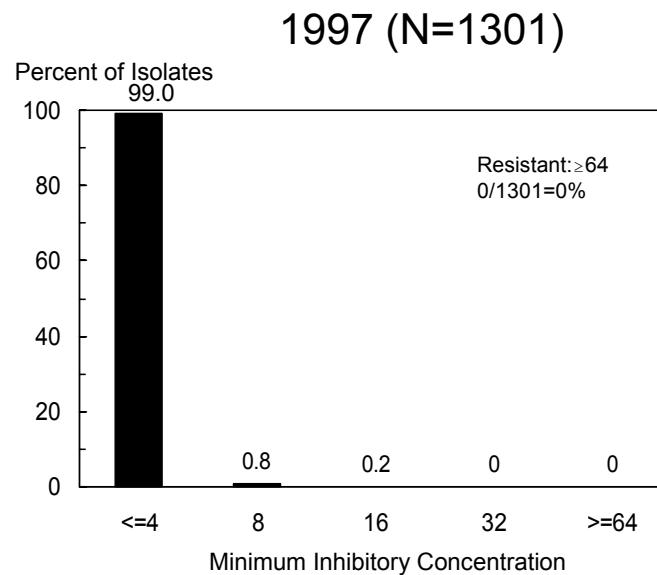
CA=Alameda, Contra Costa, and San Francisco counties
NY=excluding New York City

LX=Los Angeles County
NYC=New York City

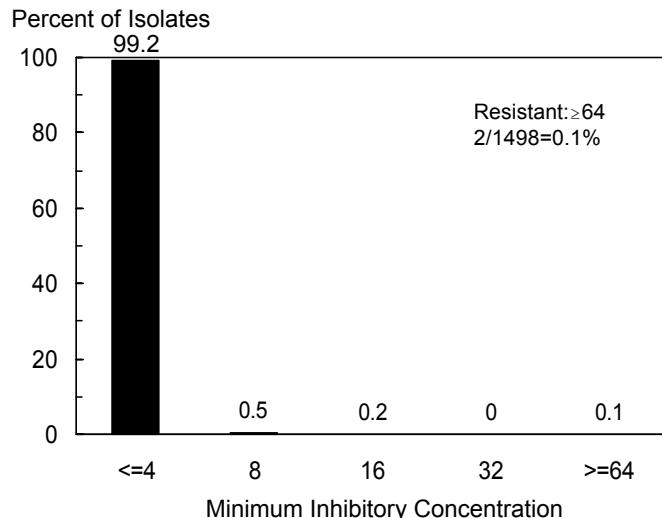
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Figure 2a. MICs for amikacin among non-Typhi *Salmonella* isolates, 1996-2001

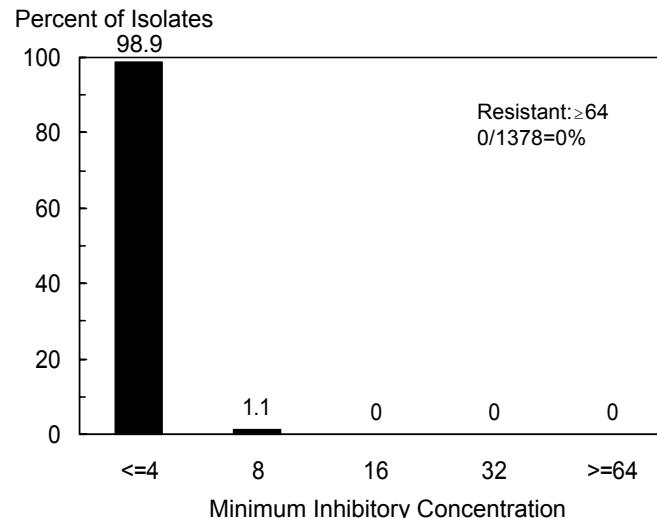
Not Tested in 1996



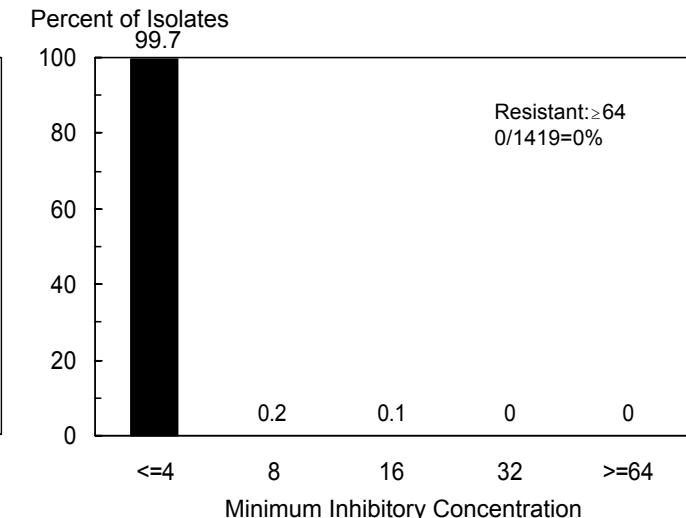
1999 (N=1498)



2000 (N=1378)



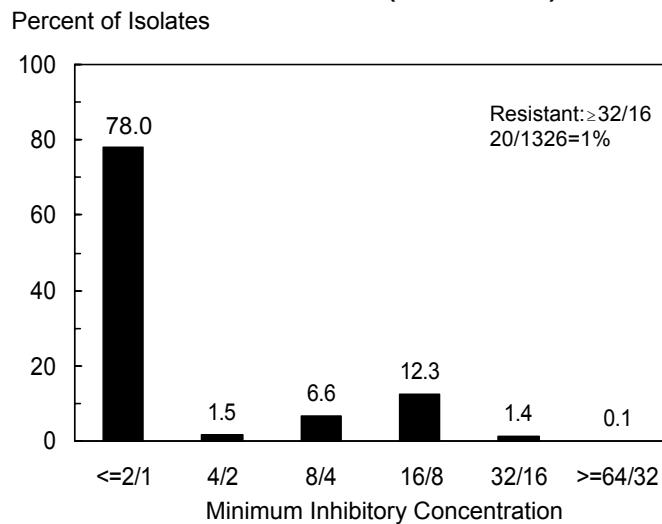
2001 (N=1419)



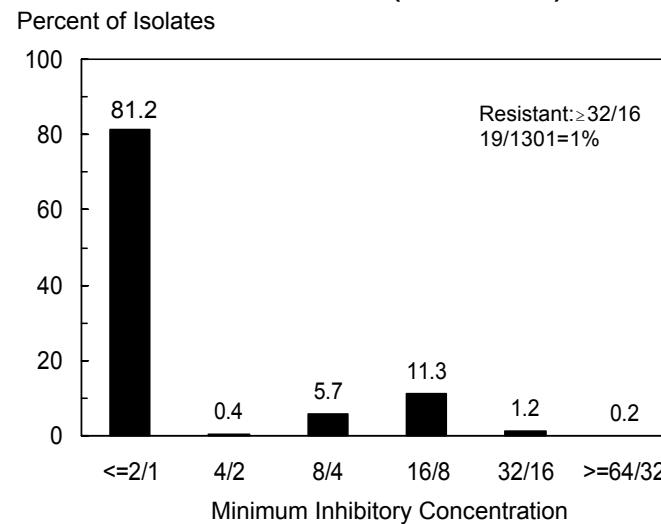
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Figure 2b. MICs for amoxicillin-clavulanic acid among non-Typhi *Salmonella* isolates, 1996-2001

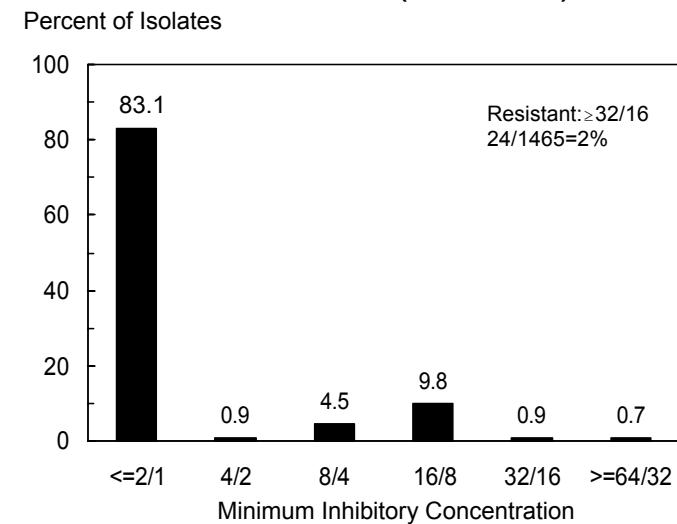
1996 (N=1326)



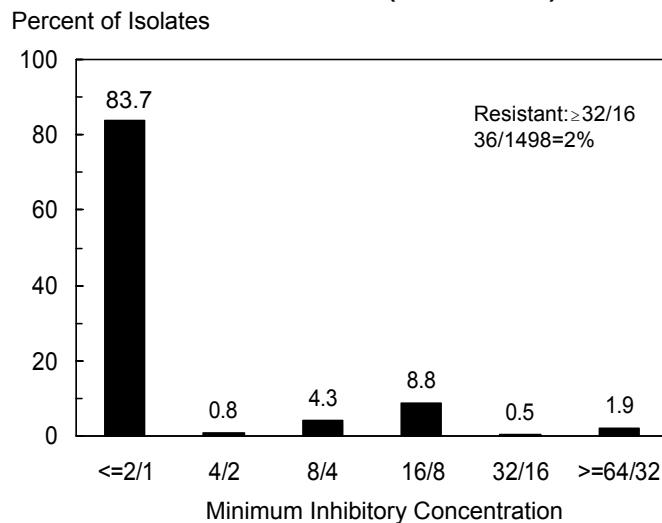
1997 (N=1301)



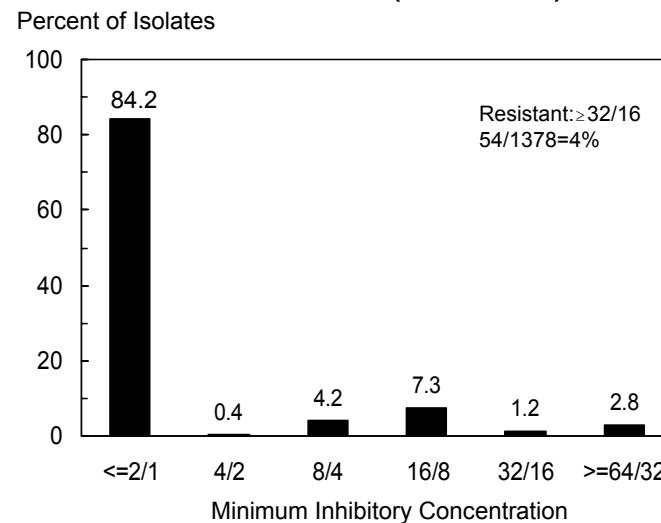
1998 (N=1465)



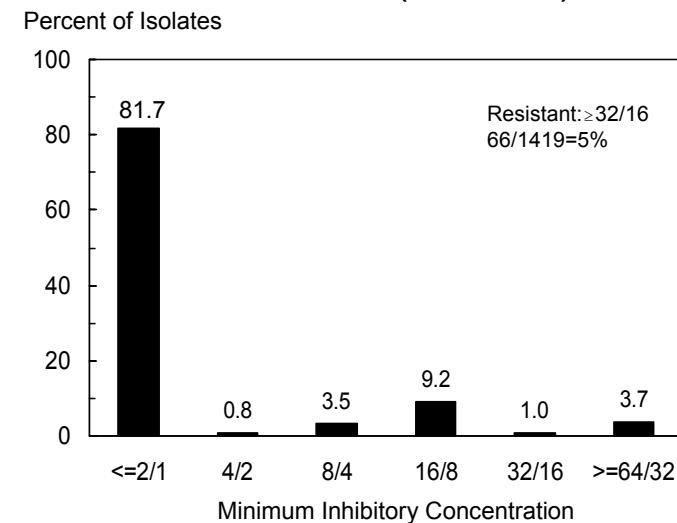
1999 (N=1498)



2000 (N=1378)

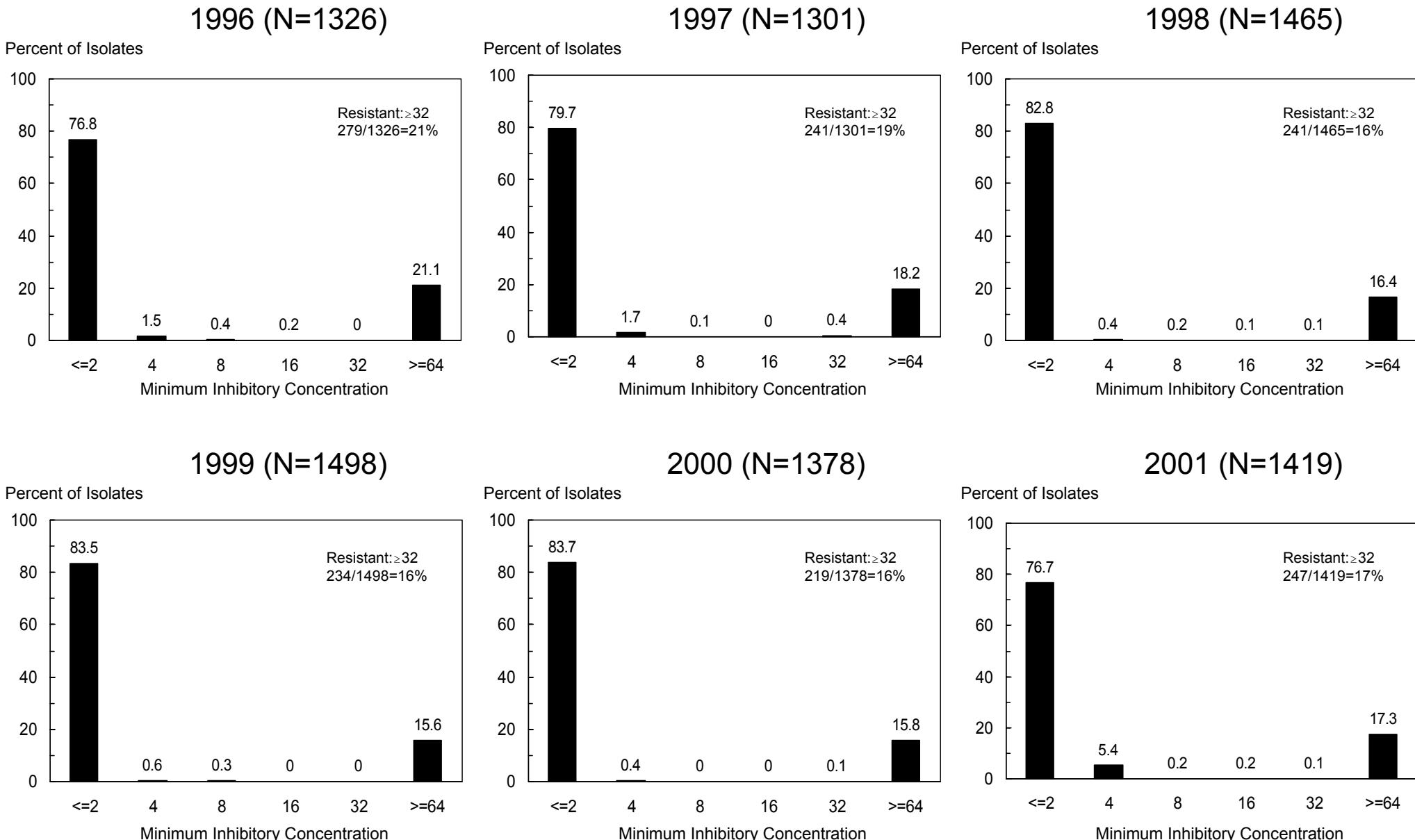


2001 (N=1419)



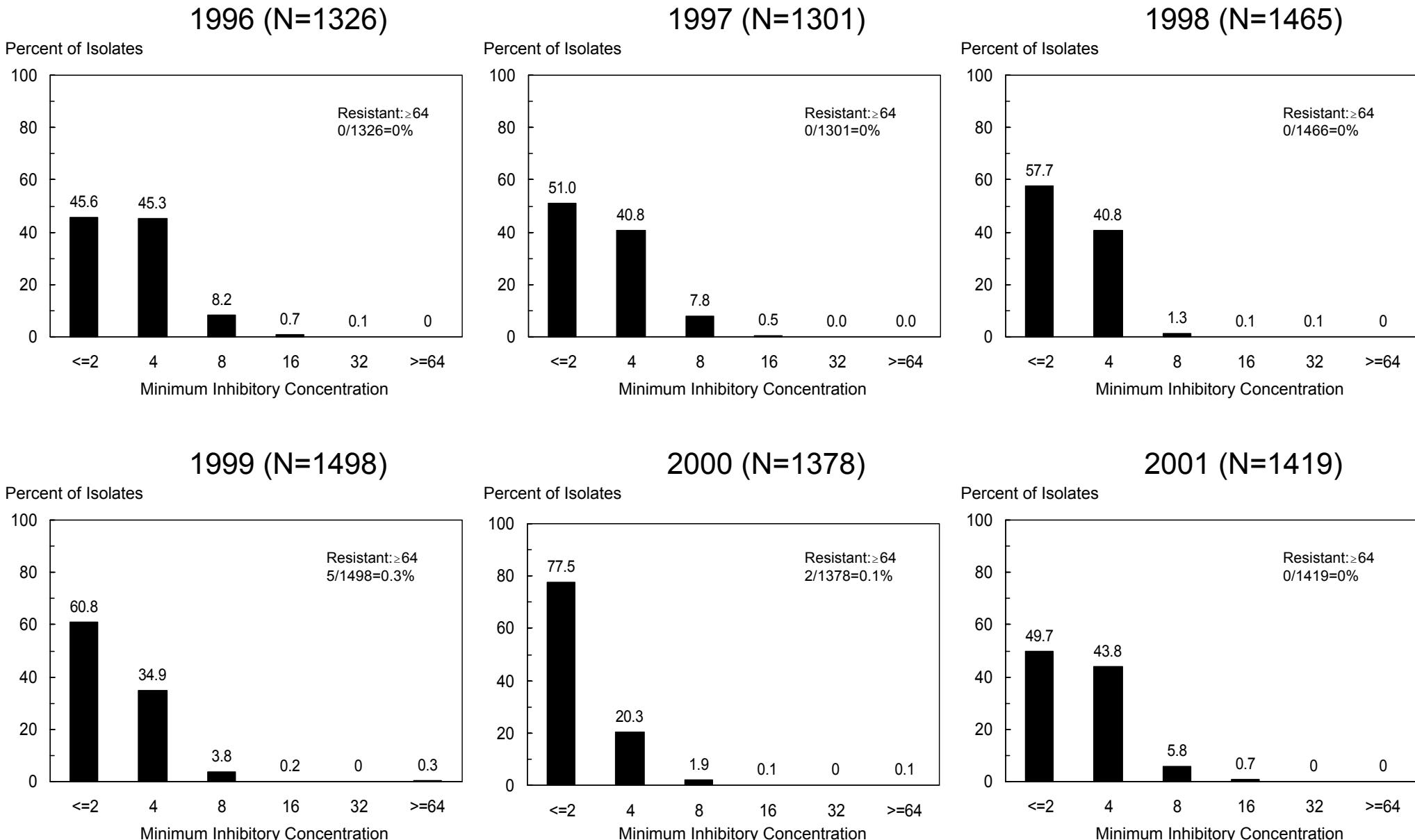
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Figure 2c. MICs for ampicillin among non-Typhi *Salmonella* isolates, 1996-2001



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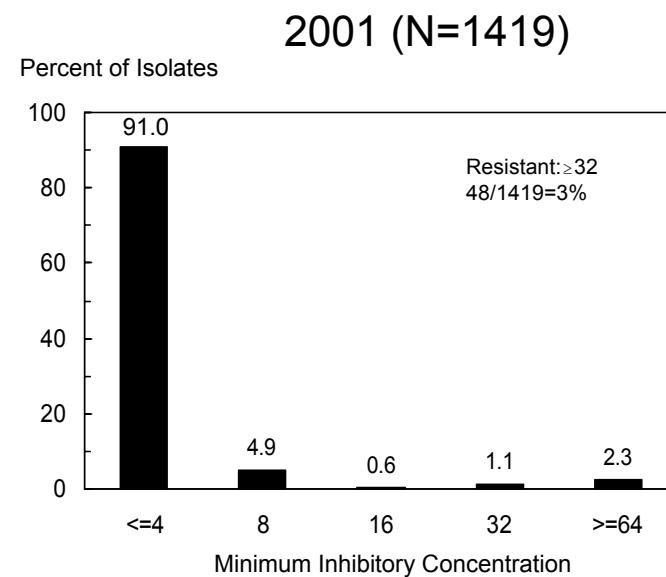
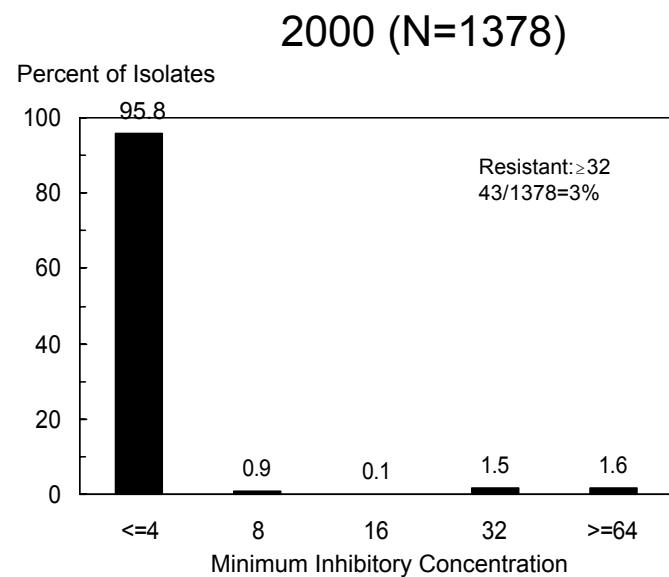
Figure 2d. MICs for apramycin among non-Typhi *Salmonella* isolates, 1996-2001



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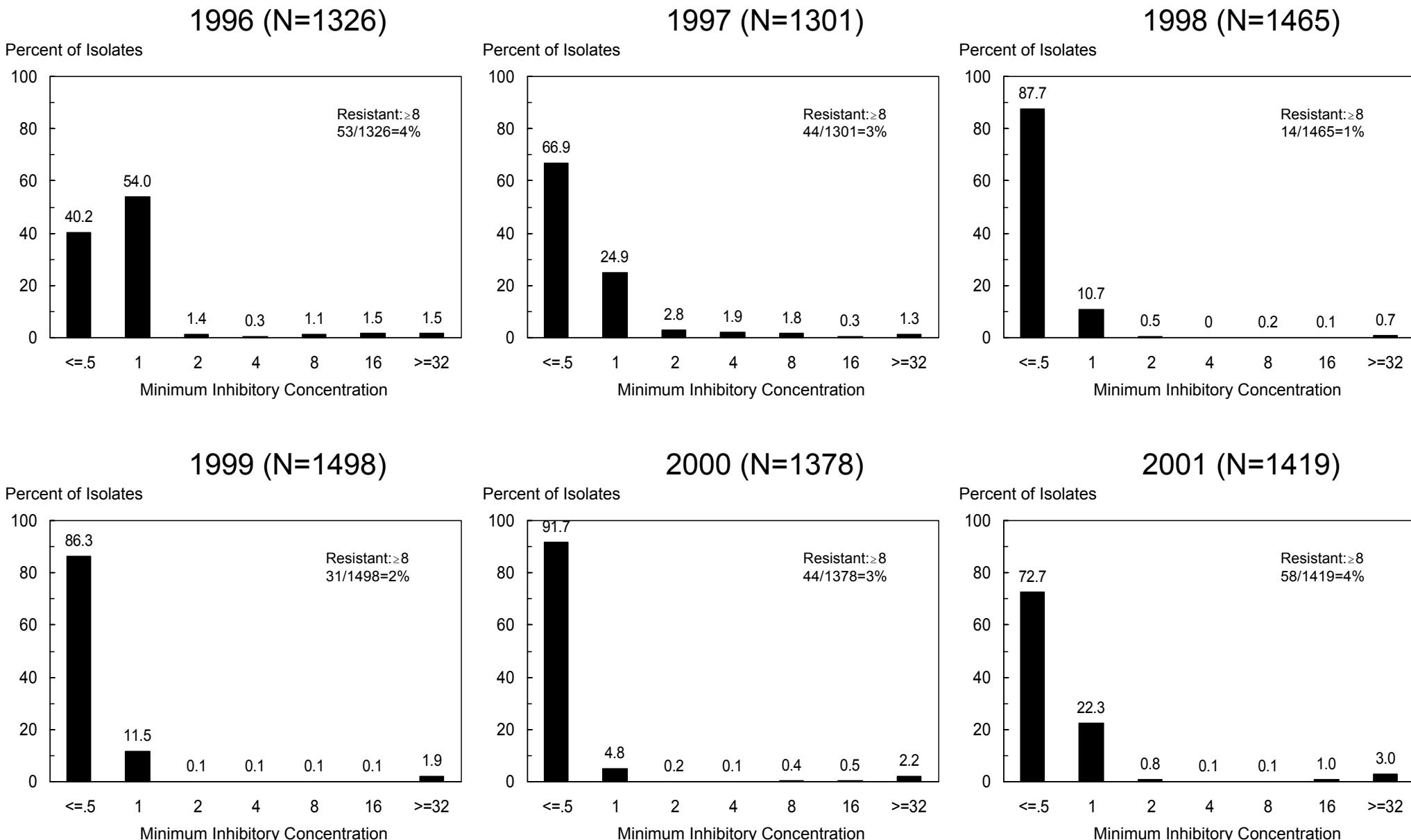
Figure 2e. MICs for cefoxitin among non-Typhi *Salmonella* isolates, 1996-2001

Not Tested in 1996 - 1999



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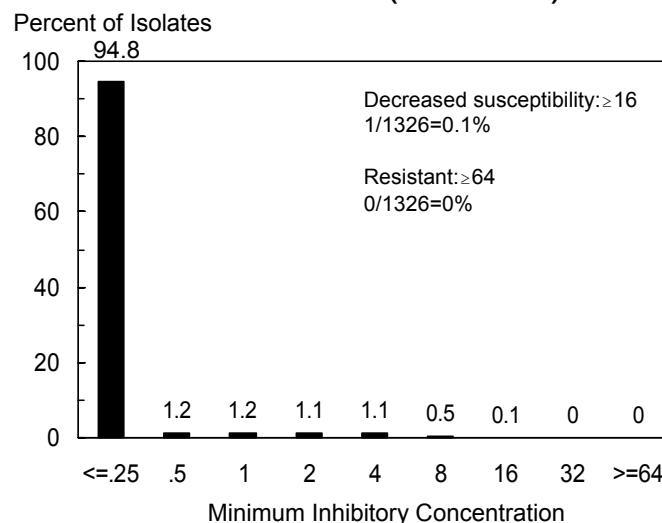
Figure 2f. MICs for ceftiofur among non-Typhi *Salmonella* isolates, 1996-2001



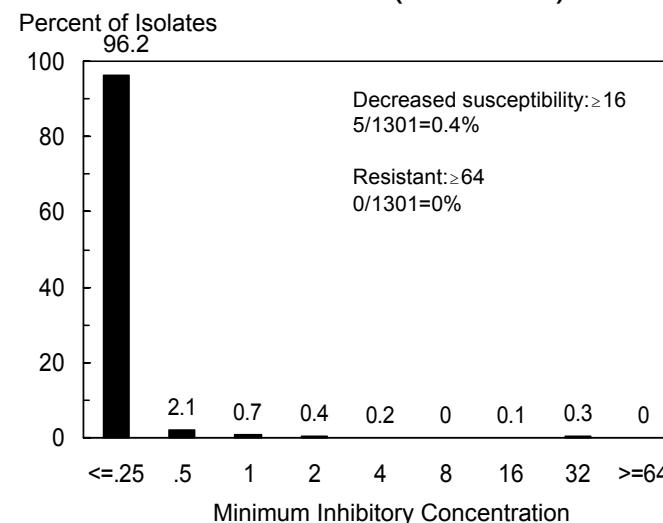
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Figure 2g. MICs for ceftriaxone* among non-Typhi *Salmonella* isolates, 1996-2001

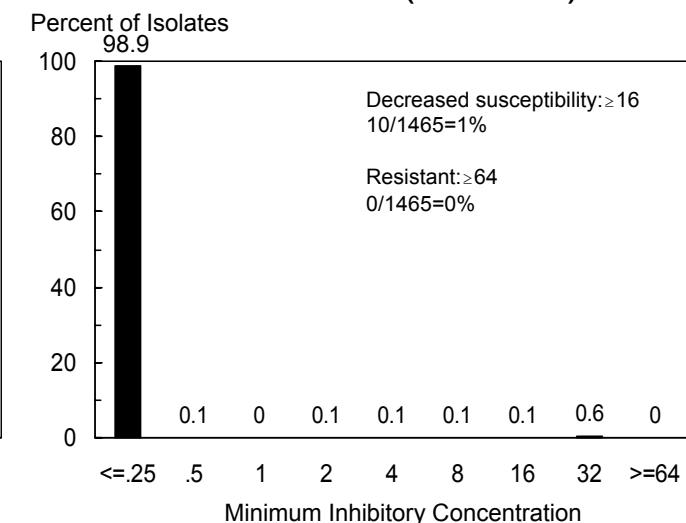
1996 (N=1326)



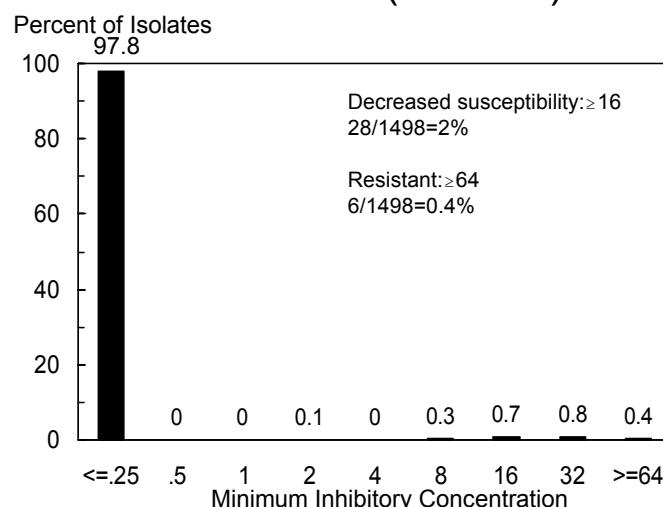
1997 (N=1301)



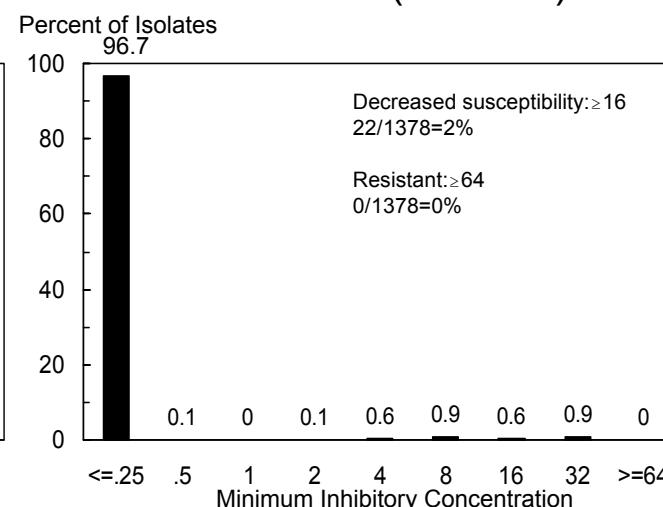
1998 (N=1465)



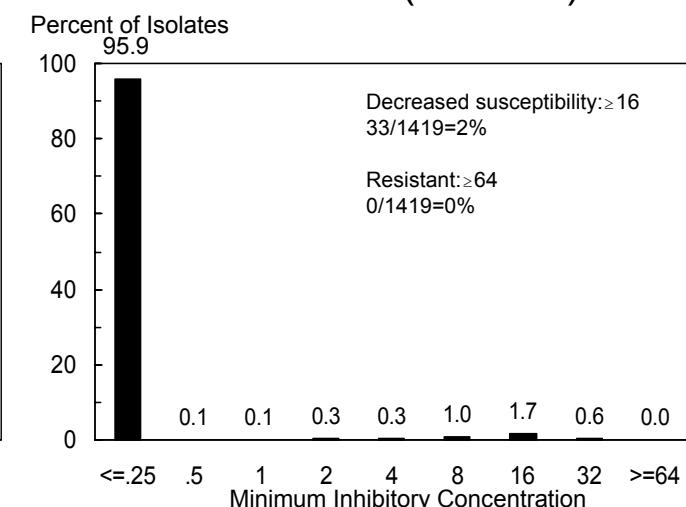
1999 (N=1498)



2000 (N=1378)



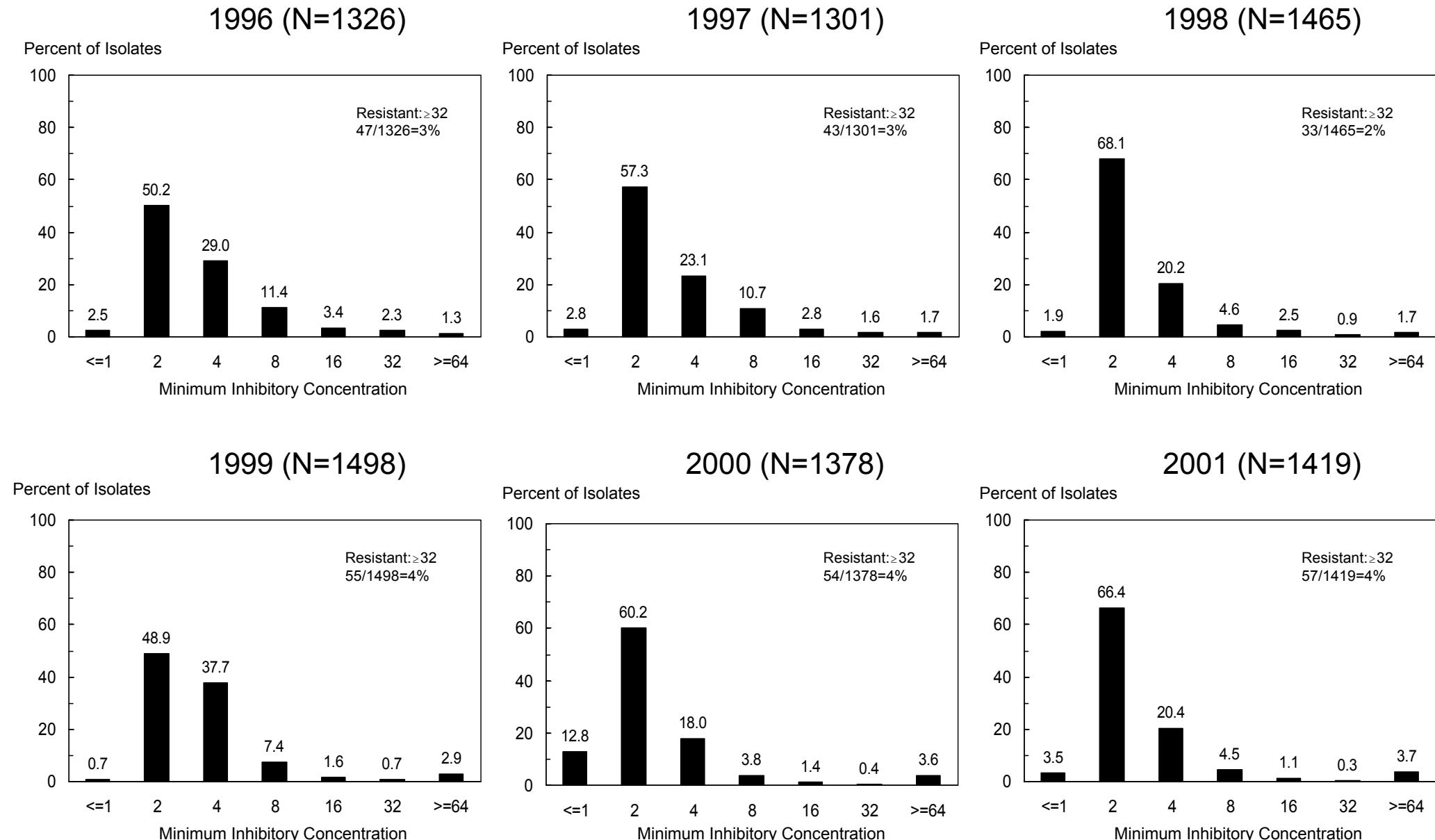
2001 (N=1419)



* Sensititre® results only

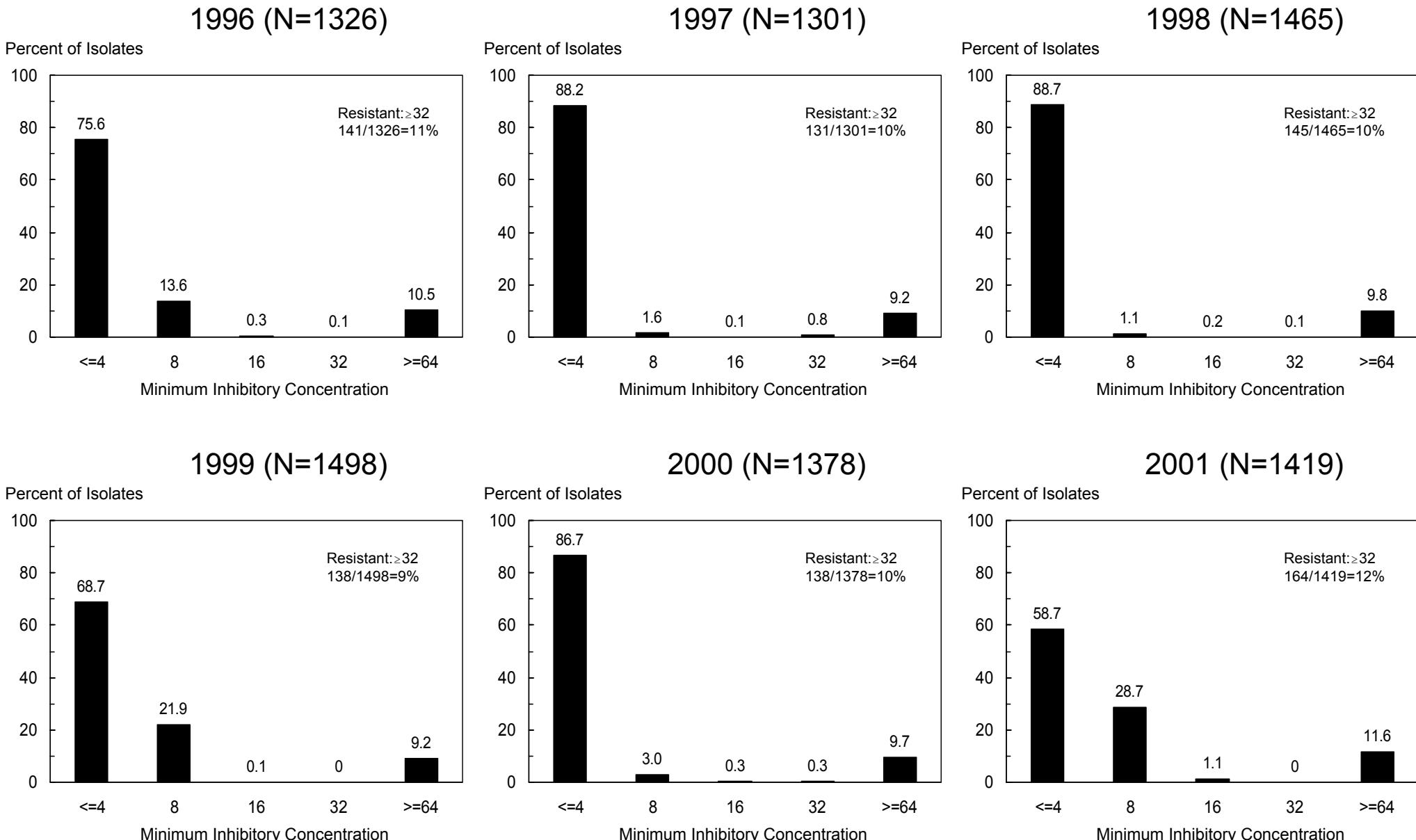
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Figure 2h. MICs for cephalothin among non-Typhi *Salmonella* isolates, 1996-2001



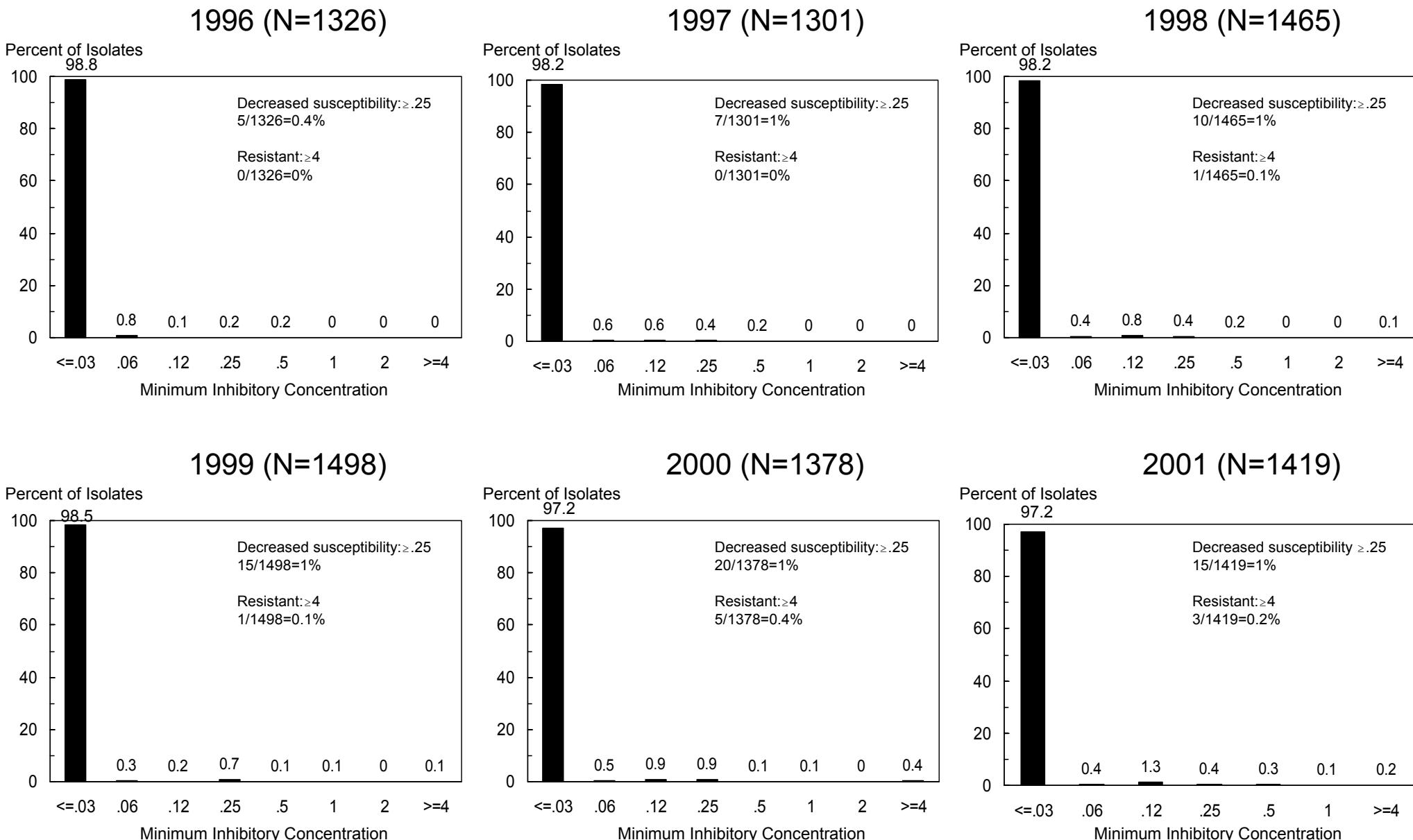
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Figure 2i. MICs for chloramphenicol among non-Typhi *Salmonella* isolates, 1996-2001



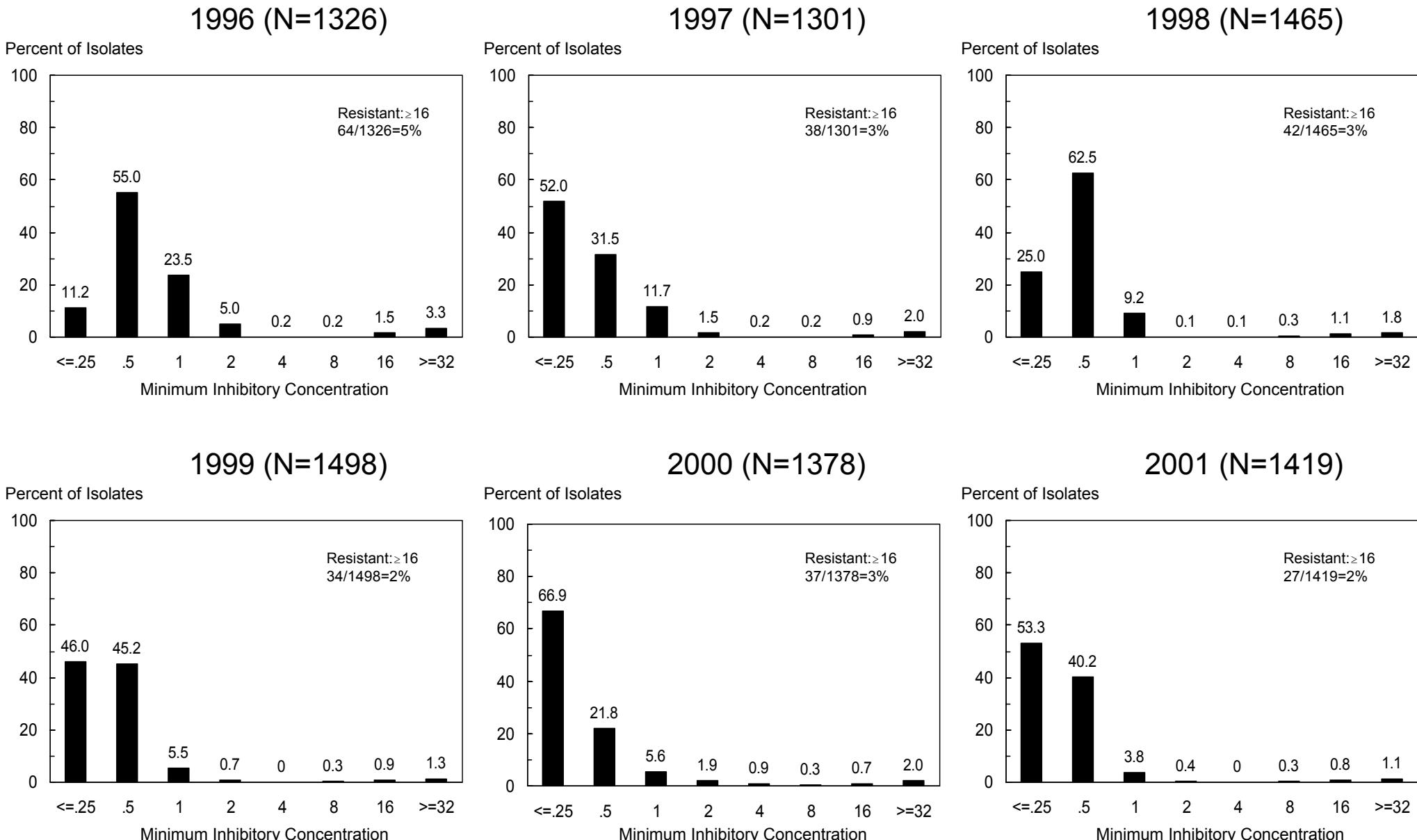
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Figure 2j. MICs for ciprofloxacin among non-Typhi *Salmonella* isolates, 1996-2001



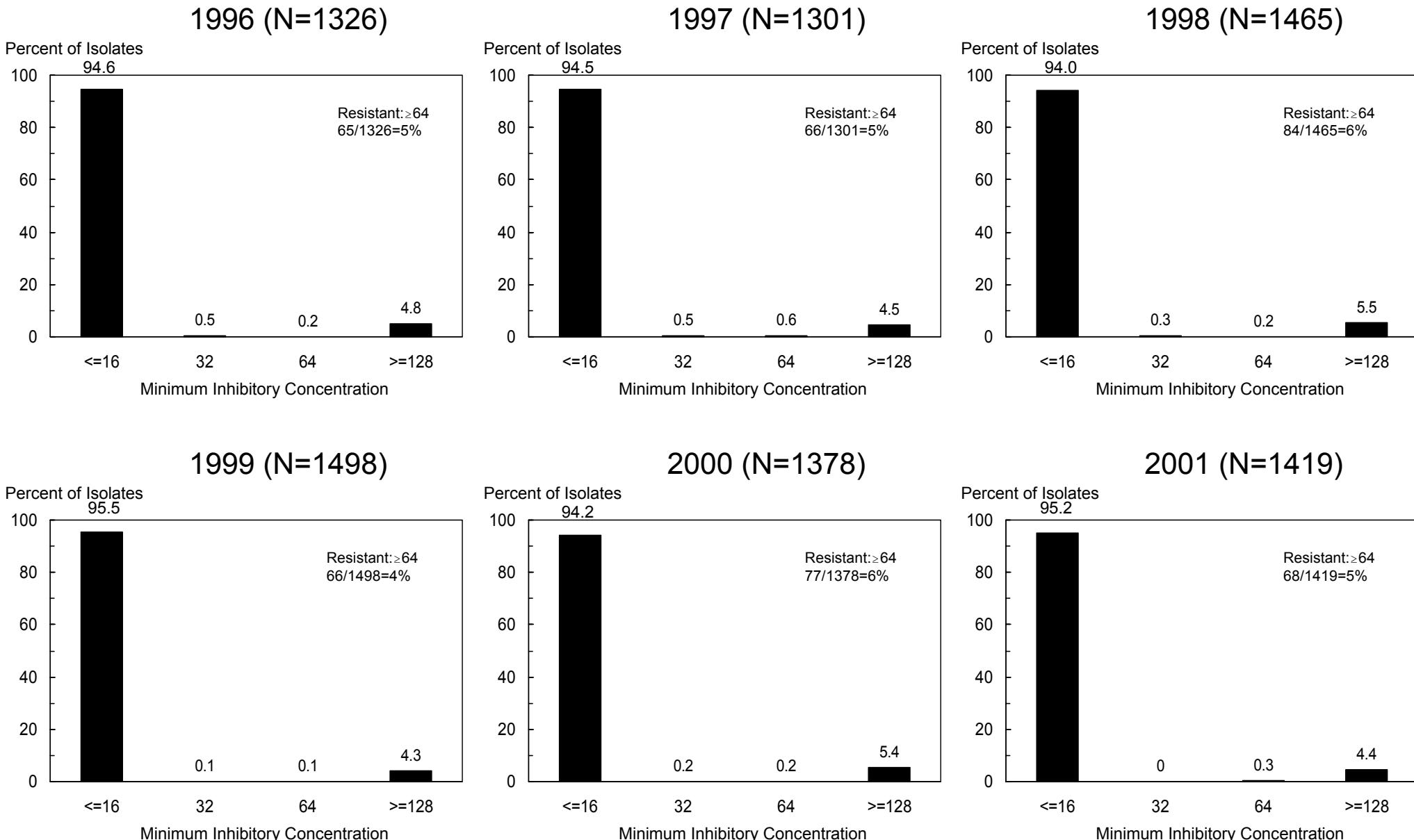
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Figure 2k. MICs for gentamicin among non-Typhi *Salmonella* isolates, 1996-2001



National Antimicrobial Resistance Monitoring System For Enteric Bacteria

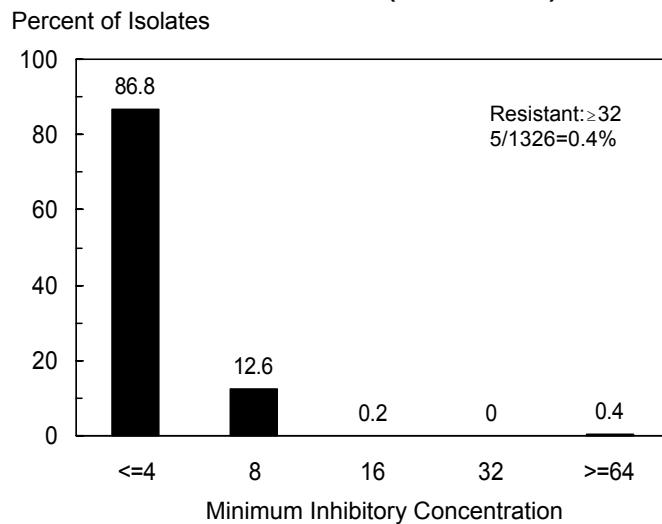
Figure 2I. MICs for kanamycin among non-Typhi *Salmonella* isolates, 1996-2001



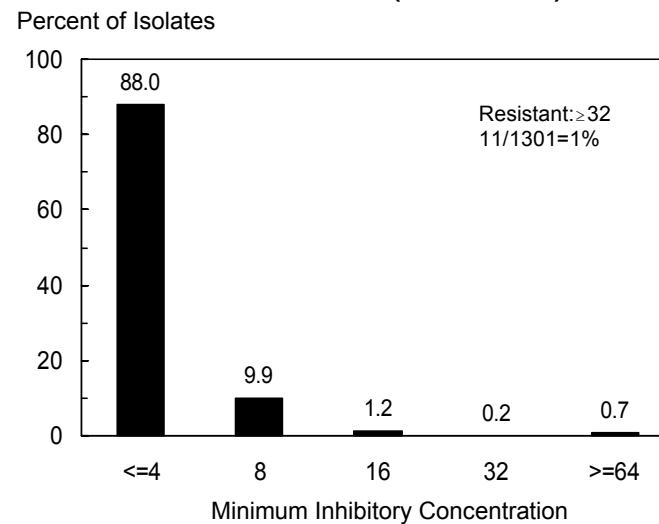
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Figure 2m. MICs for nalidixic acid among non-Typhi *Salmonella* isolates, 1996-2001

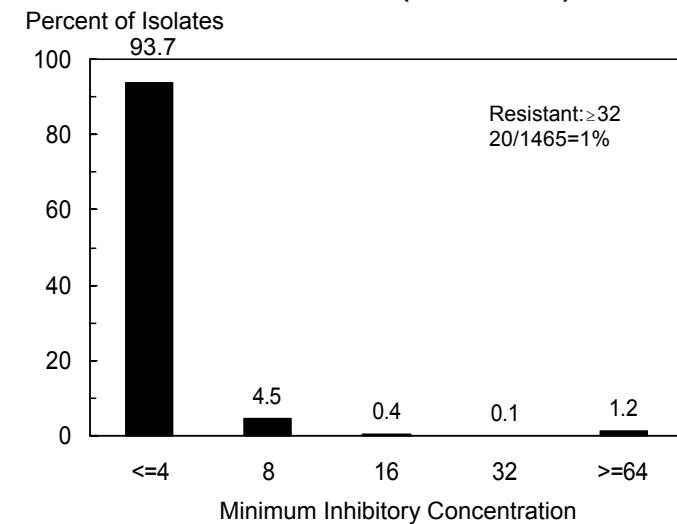
1996 (N=1326)



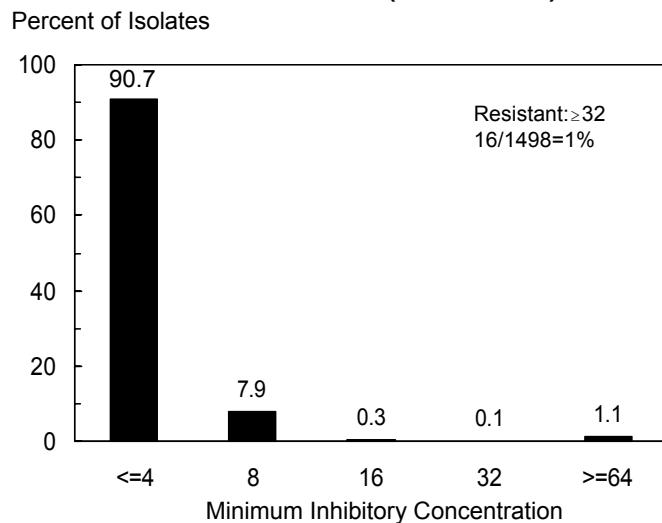
1997 (N=1301)



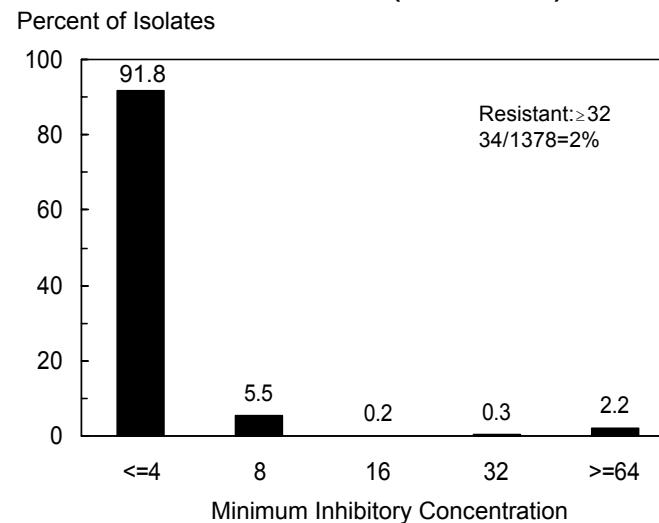
1998 (N=1465)



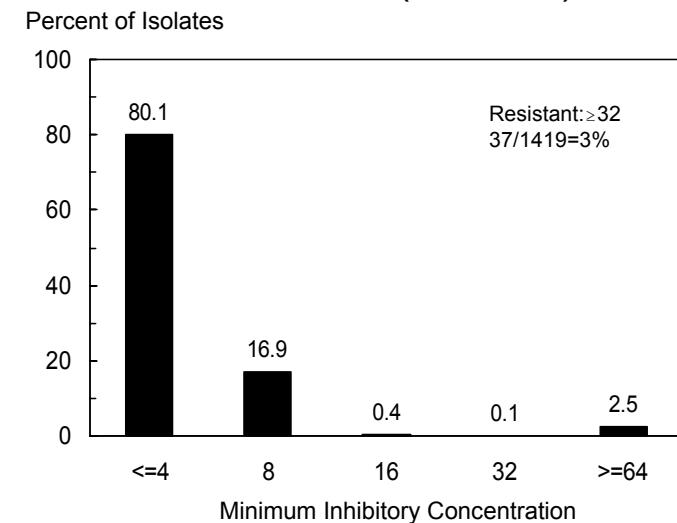
1999 (N=1498)



2000 (N=1378)



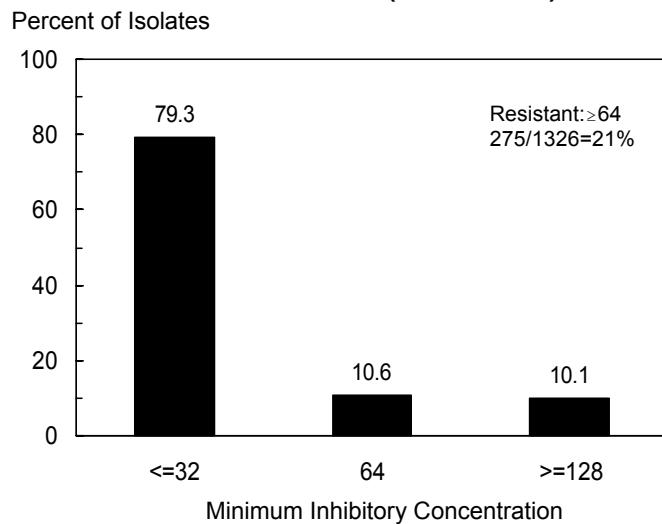
2001 (N=1419)



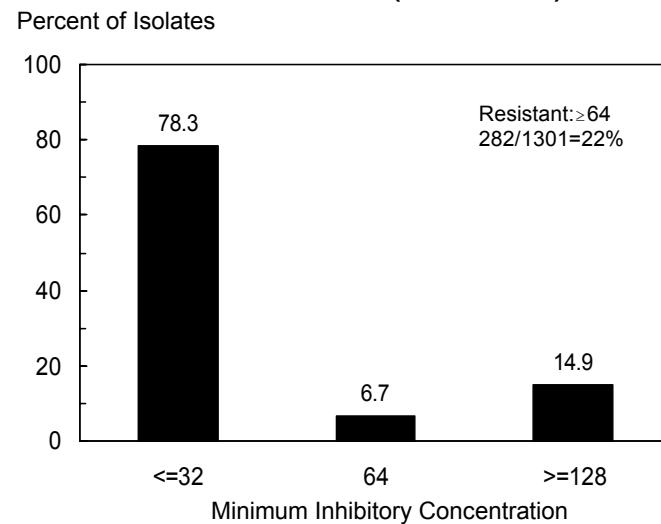
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Figure 2n. MICs for streptomycin among non-Typhi *Salmonella* isolates, 1996-2001

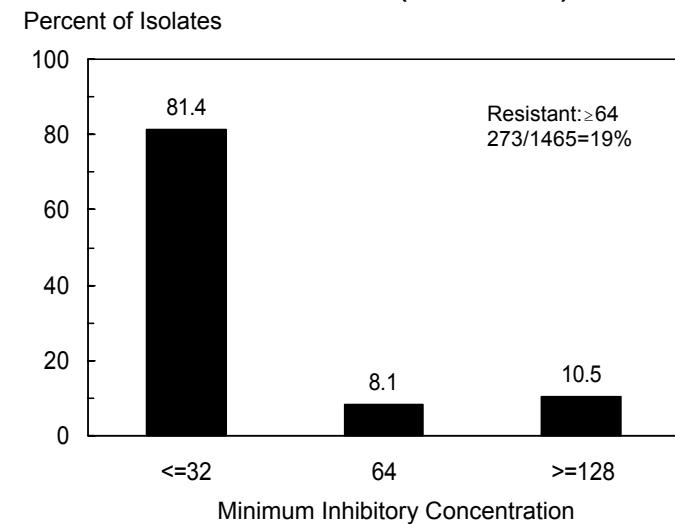
1996 (N=1326)



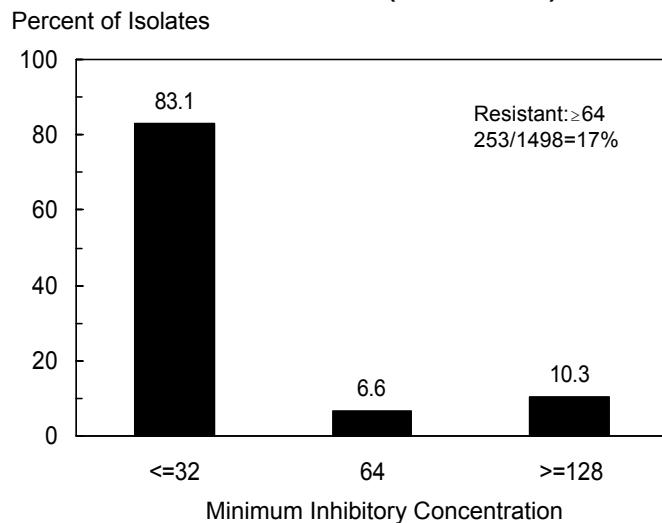
1997 (N=1301)



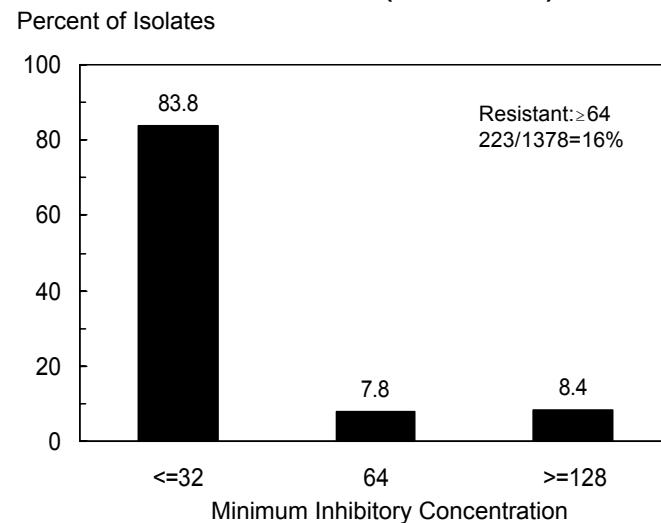
1998 (N=1465)



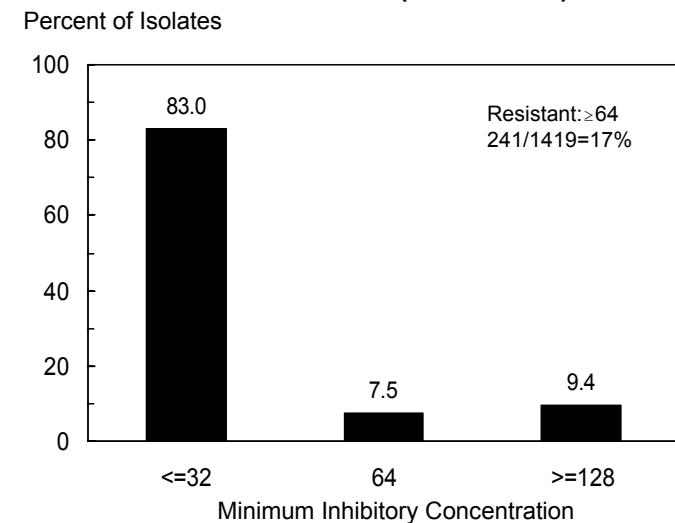
1999 (N=1498)



2000 (N=1378)

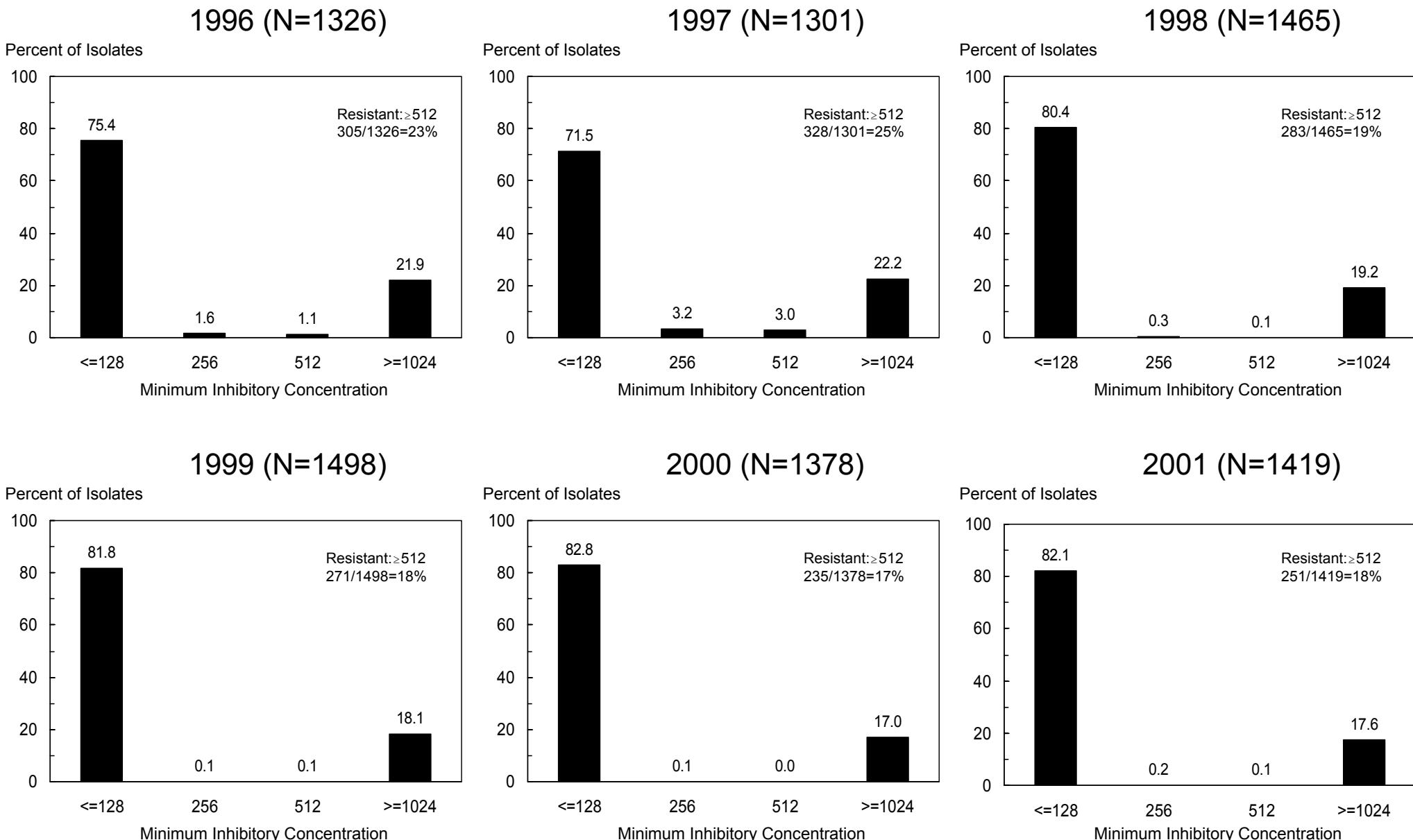


2001 (N=1419)



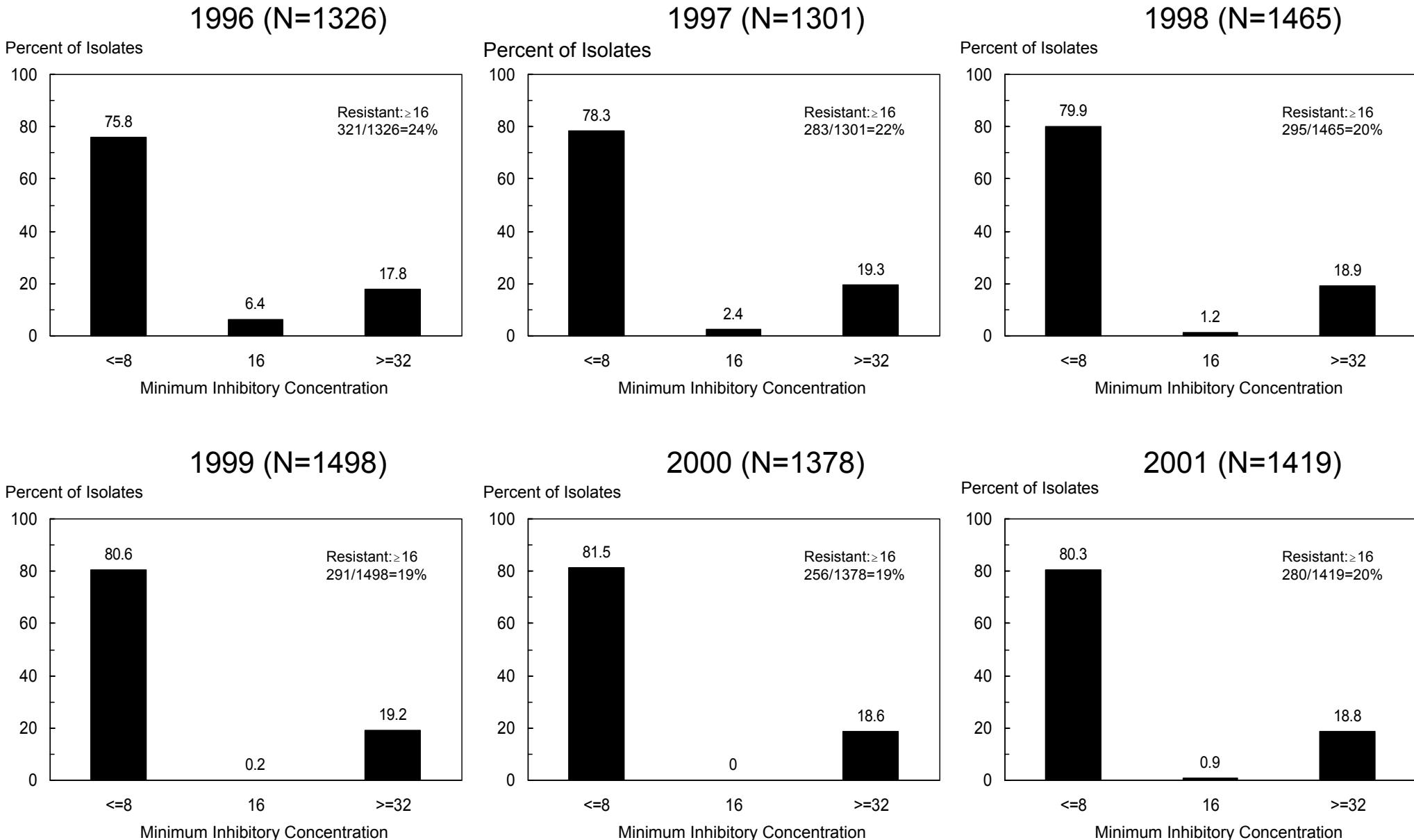
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Figure 2o. MICs for sulfamethoxazole among non-Typhi *Salmonella* isolates, 1996-2001



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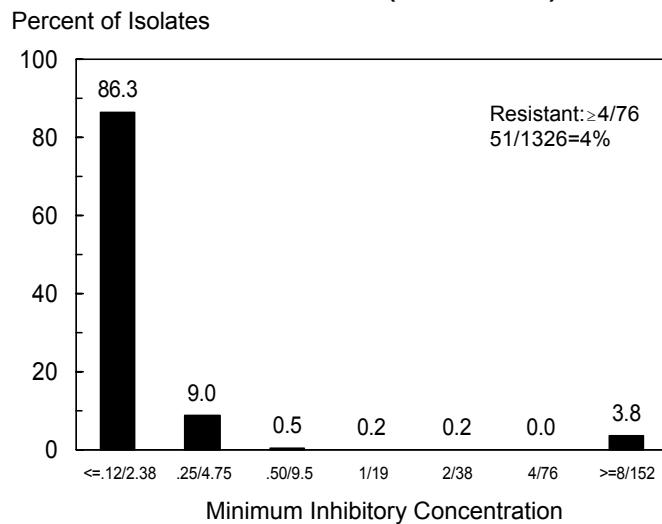
Figure 2p. MICs for tetracycline among non-Typhi *Salmonella* isolates, 1996-2001



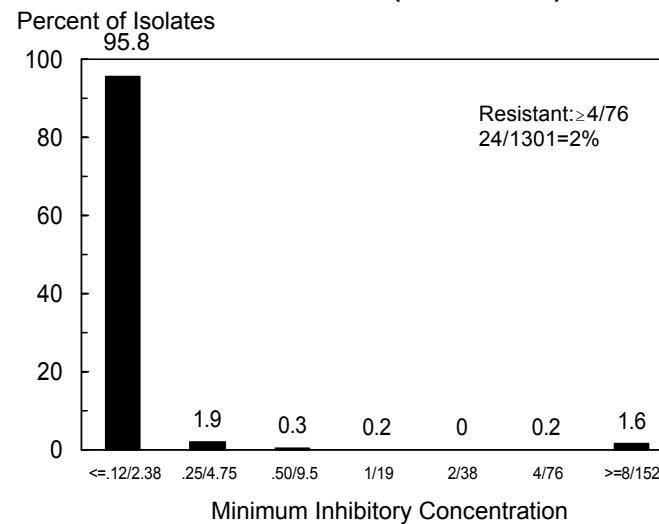
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Figure 2q. MICs for trimethoprim-sulfamethoxazole among non-Typhi *Salmonella* isolates, 1996-2001

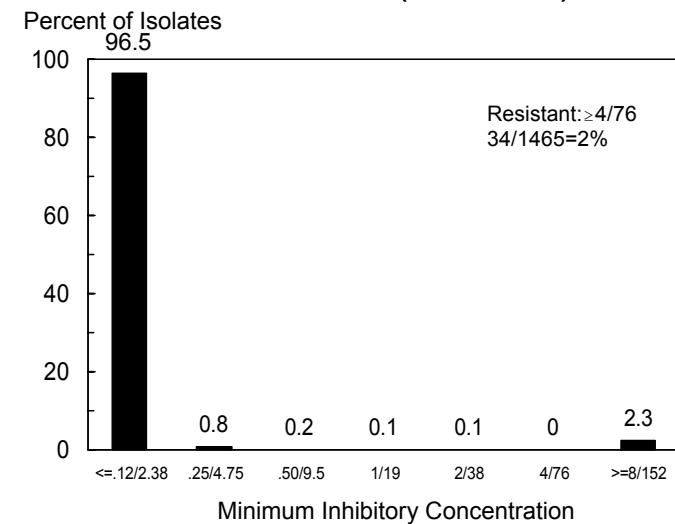
1996 (N=1326)



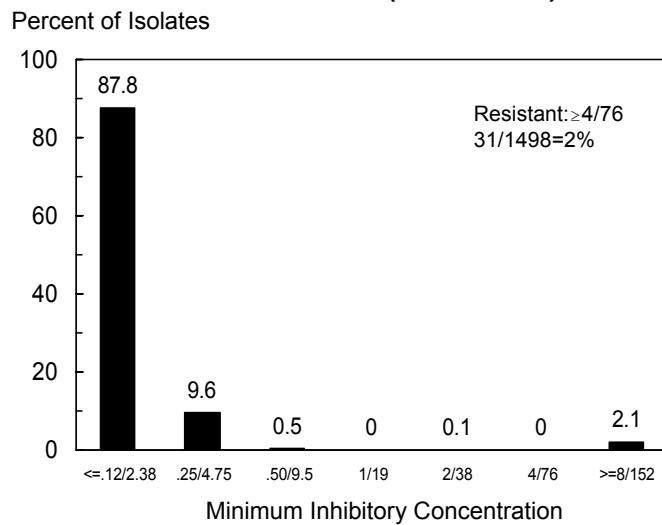
1997 (N=1301)



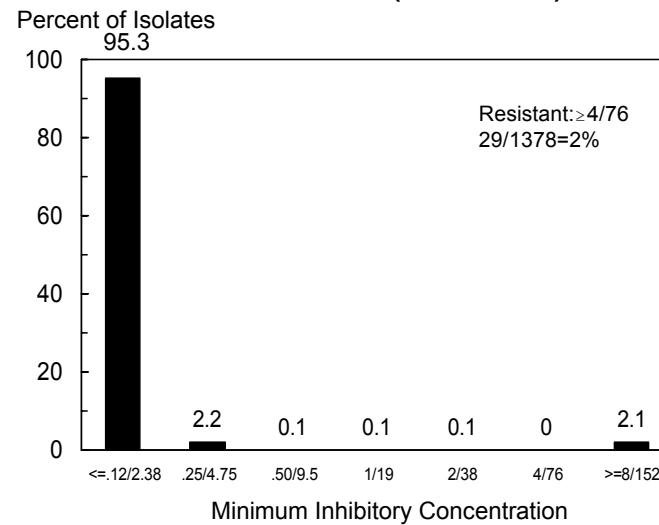
1998 (N=1465)



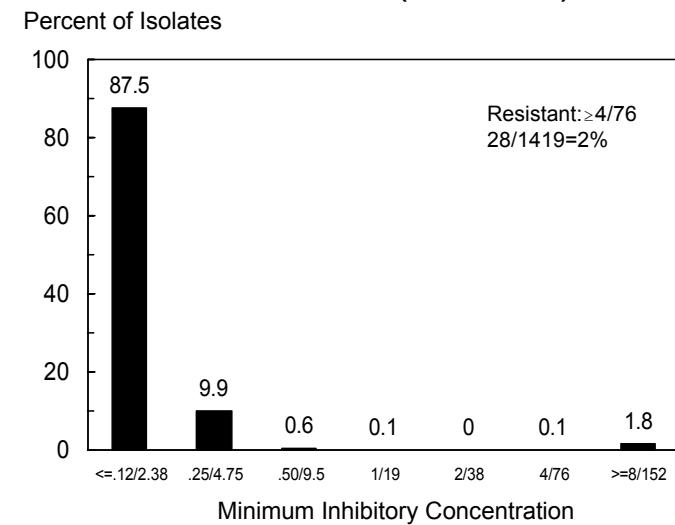
1999 (N=1498)



2000 (N=1378)

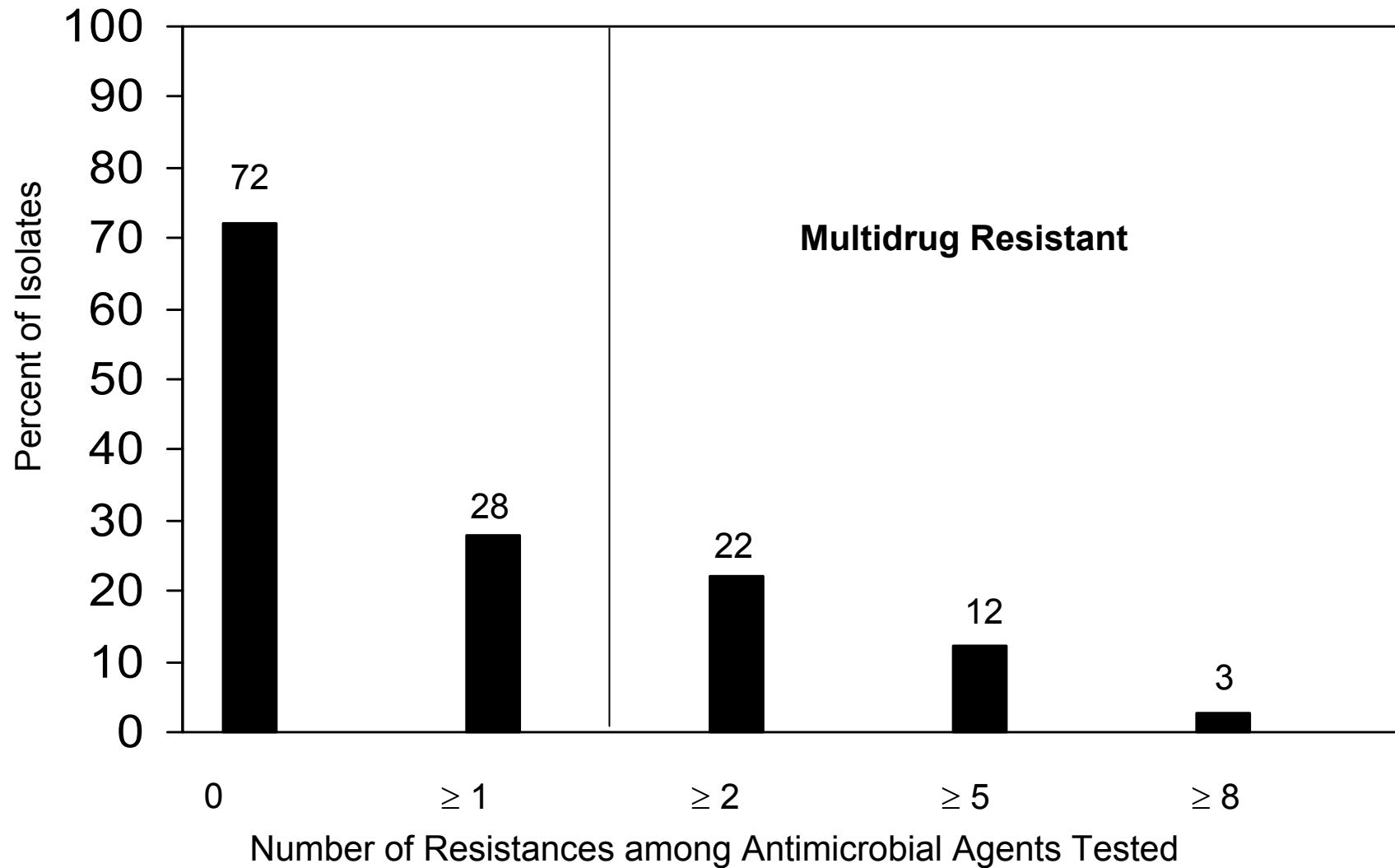


2001 (N=1419)



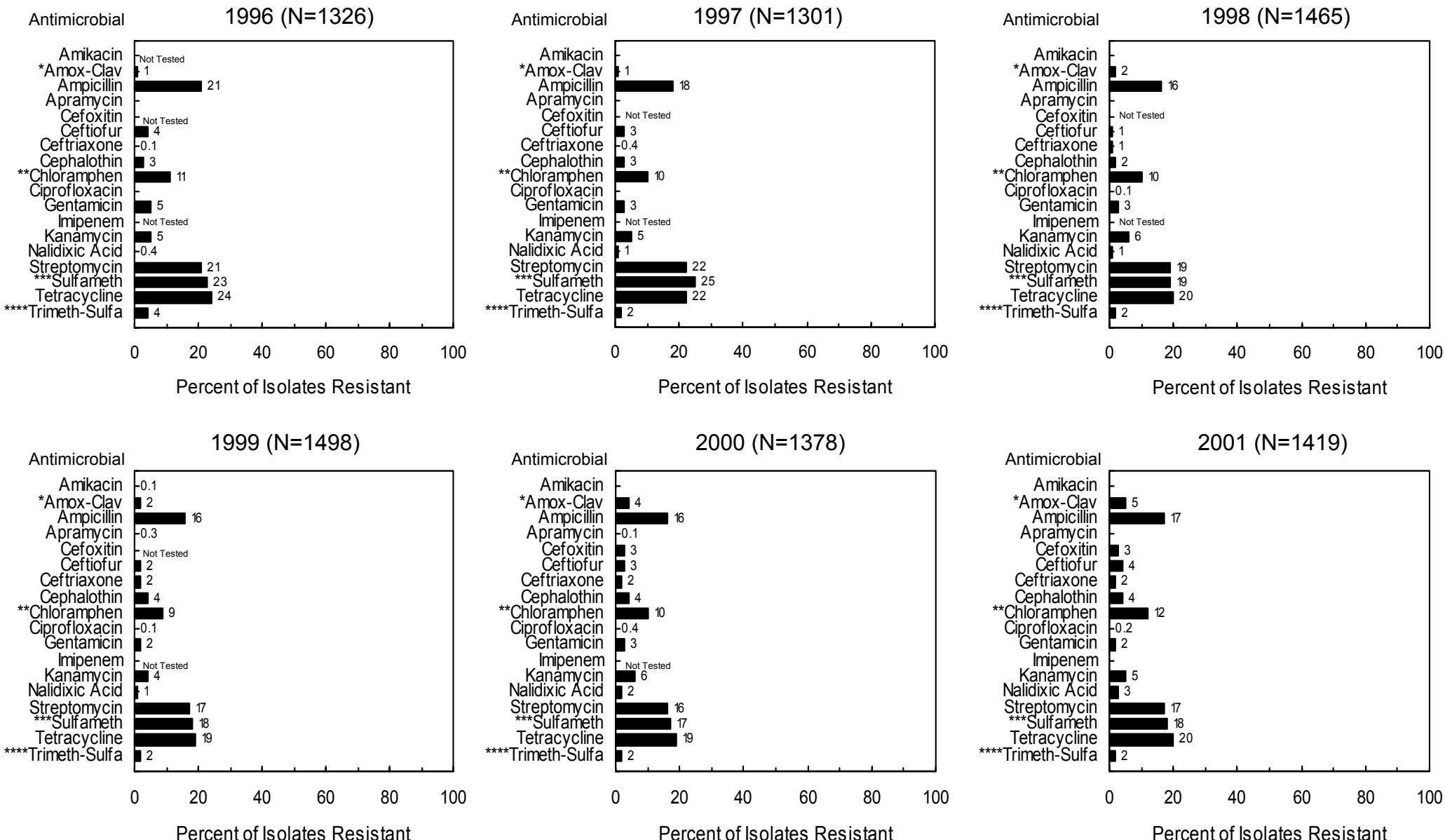
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Figure 3. Percent of non-Typhi *Salmonella* isolates (N=1419) that were pansusceptible, and percent resistant to ≥ 1 , ≥ 2 , ≥ 5 , and ≥ 8 of the 18 antimicrobial agents tested, 2001



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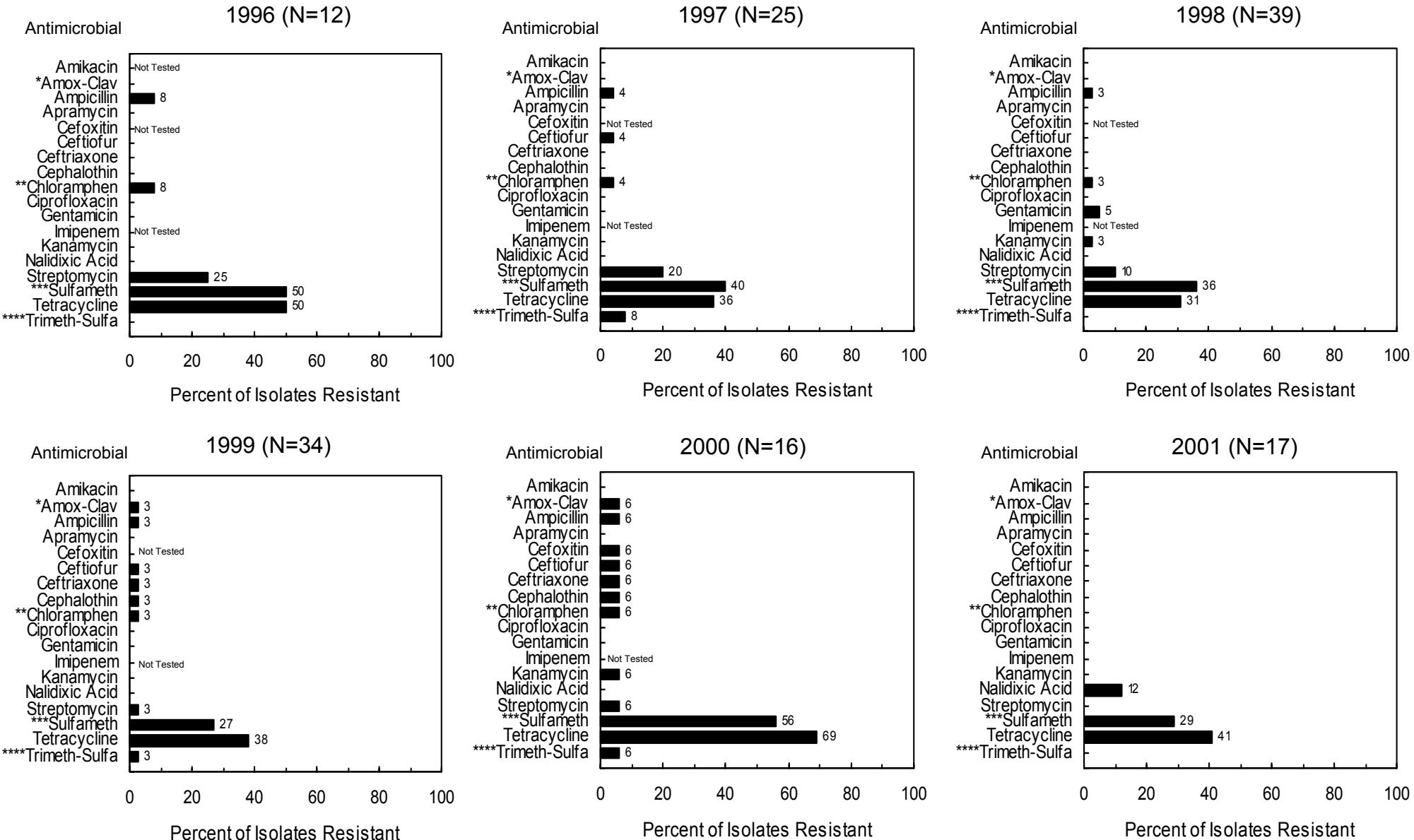
Figure 4. Resistance among non-Typhi *Salmonella* isolates, 1996-2001



*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

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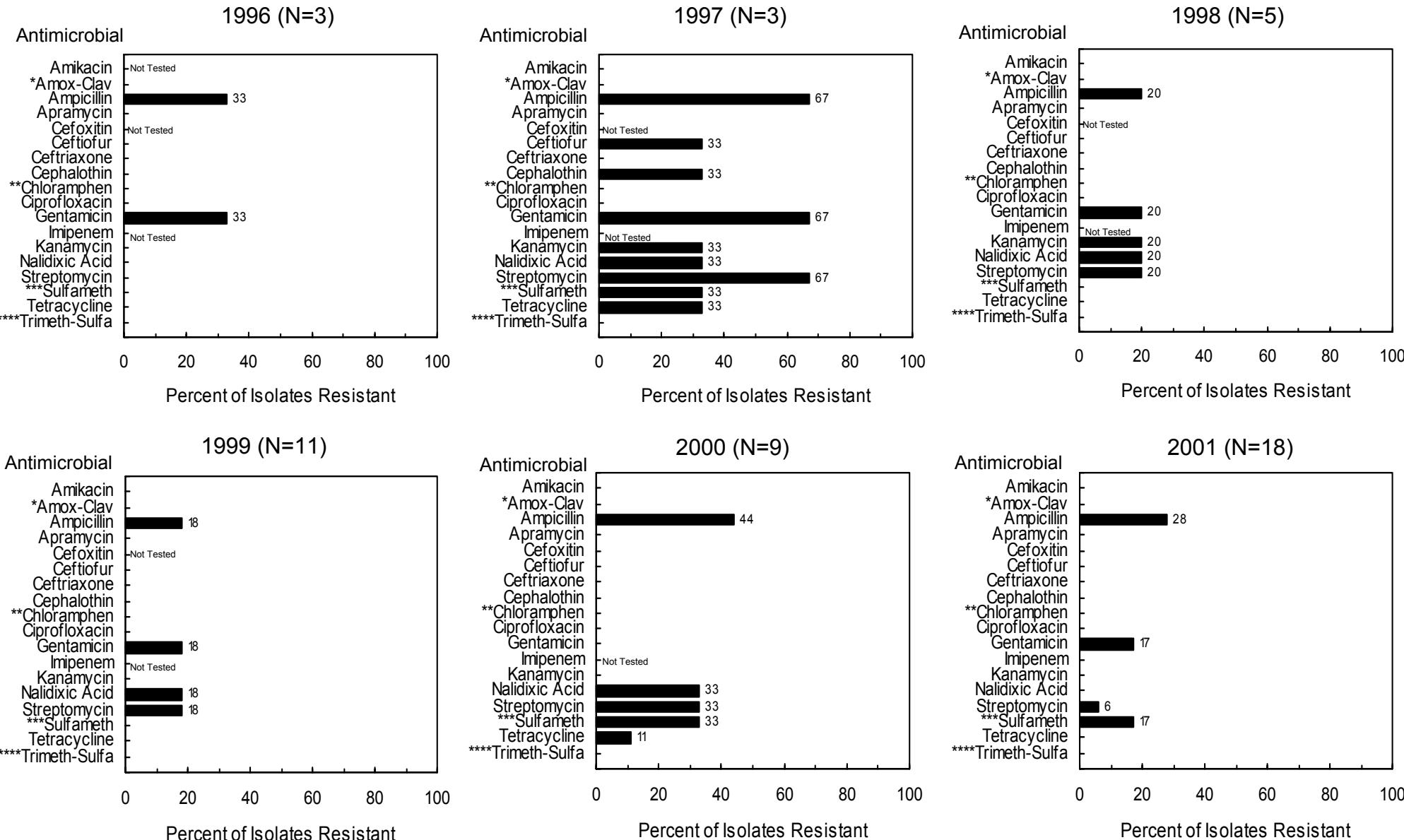
Figure 5a. Resistance among *Salmonella* serotype Agona isolates, 1996-2001



*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

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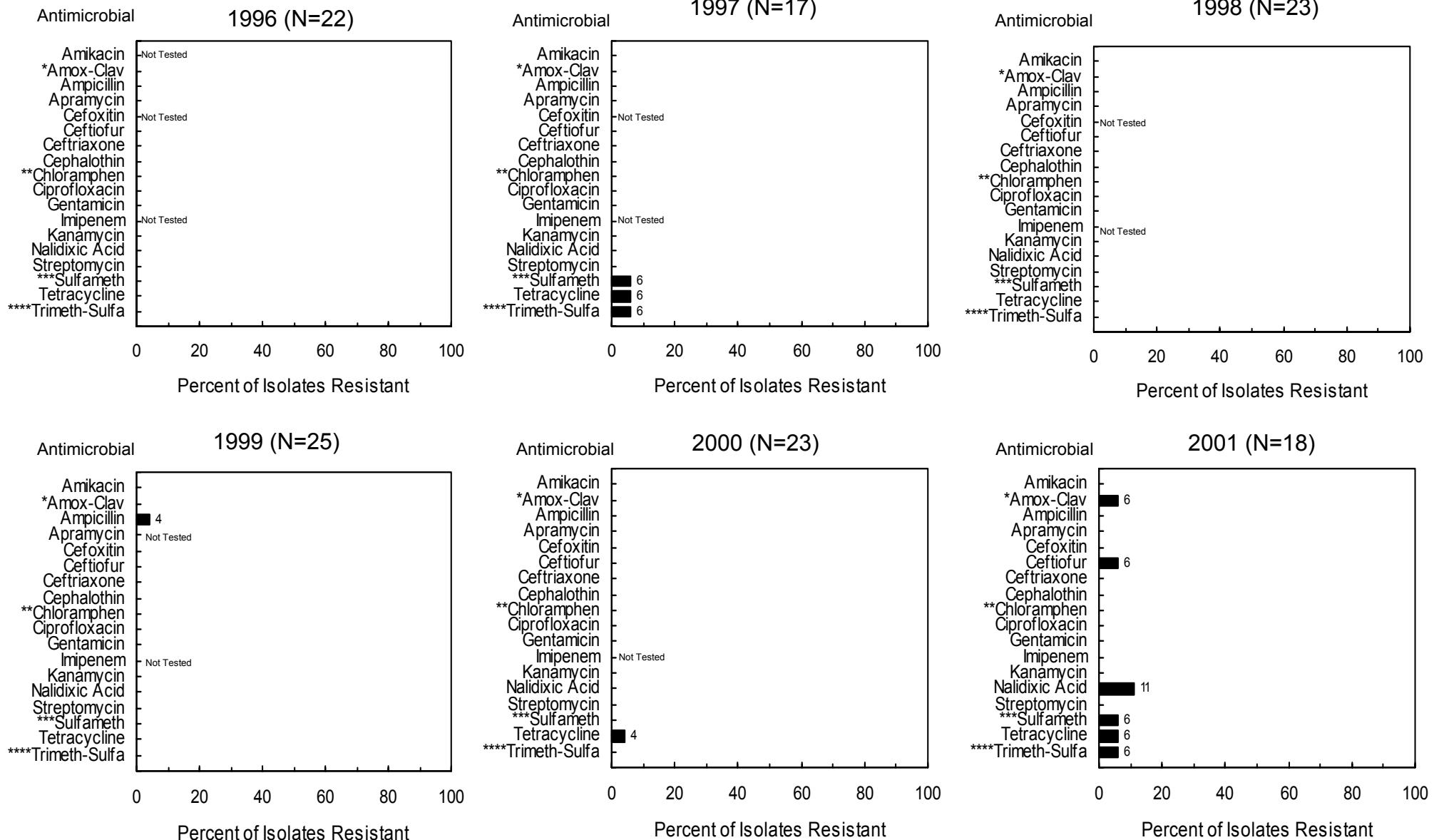
Figure 5b. Resistance among *Salmonella* serotype Berta isolates, 1996-2001



*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

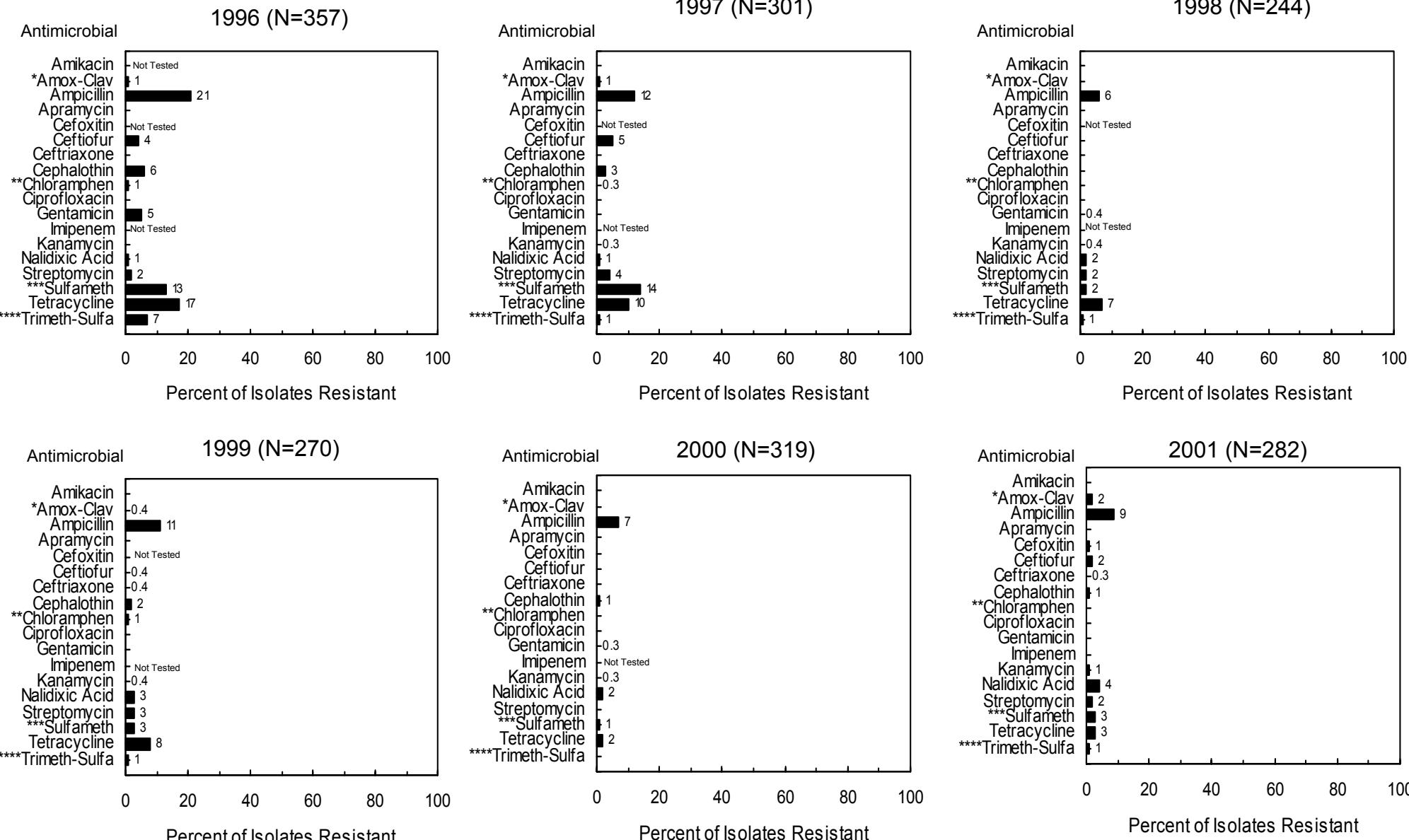
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Figure 5c. Resistance among *Salmonella* serotype Braenderup isolates, 1996-2001



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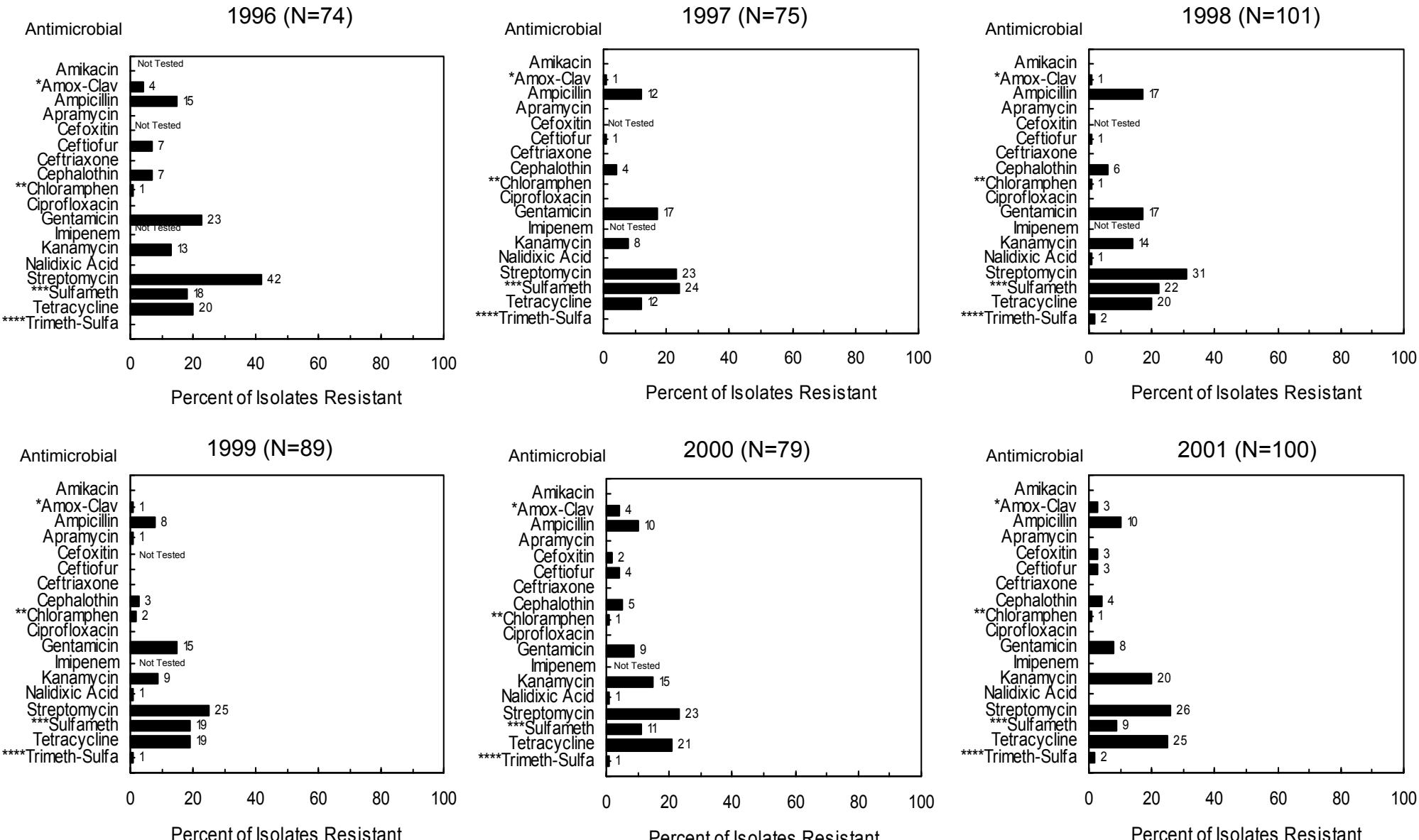
Figure 5d. Resistance among *Salmonella* serotype Enteritidis isolates, 1996-2001



*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

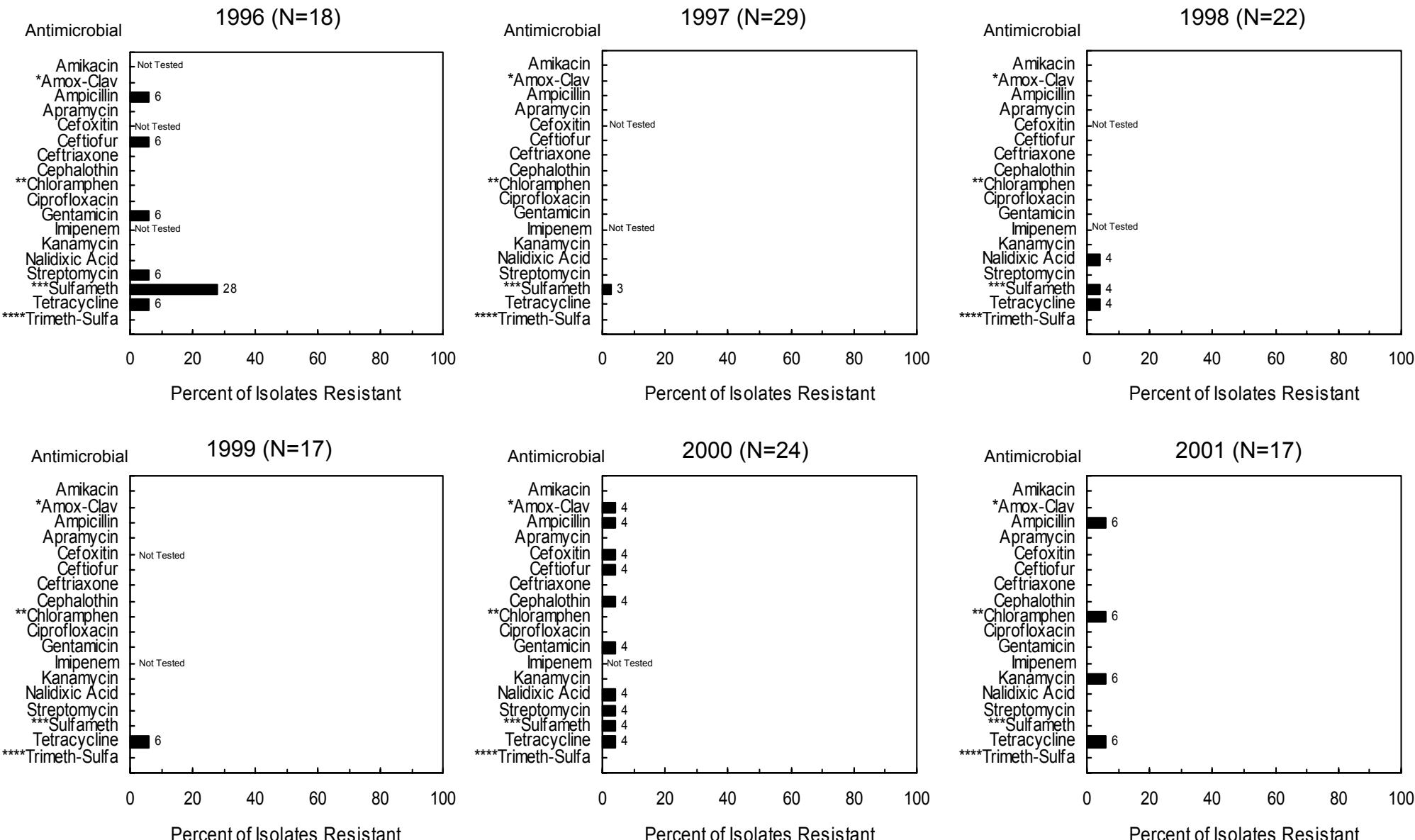
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Figure 5e. Resistance among *Salmonella* serotype Heidelberg isolates, 1996-2001



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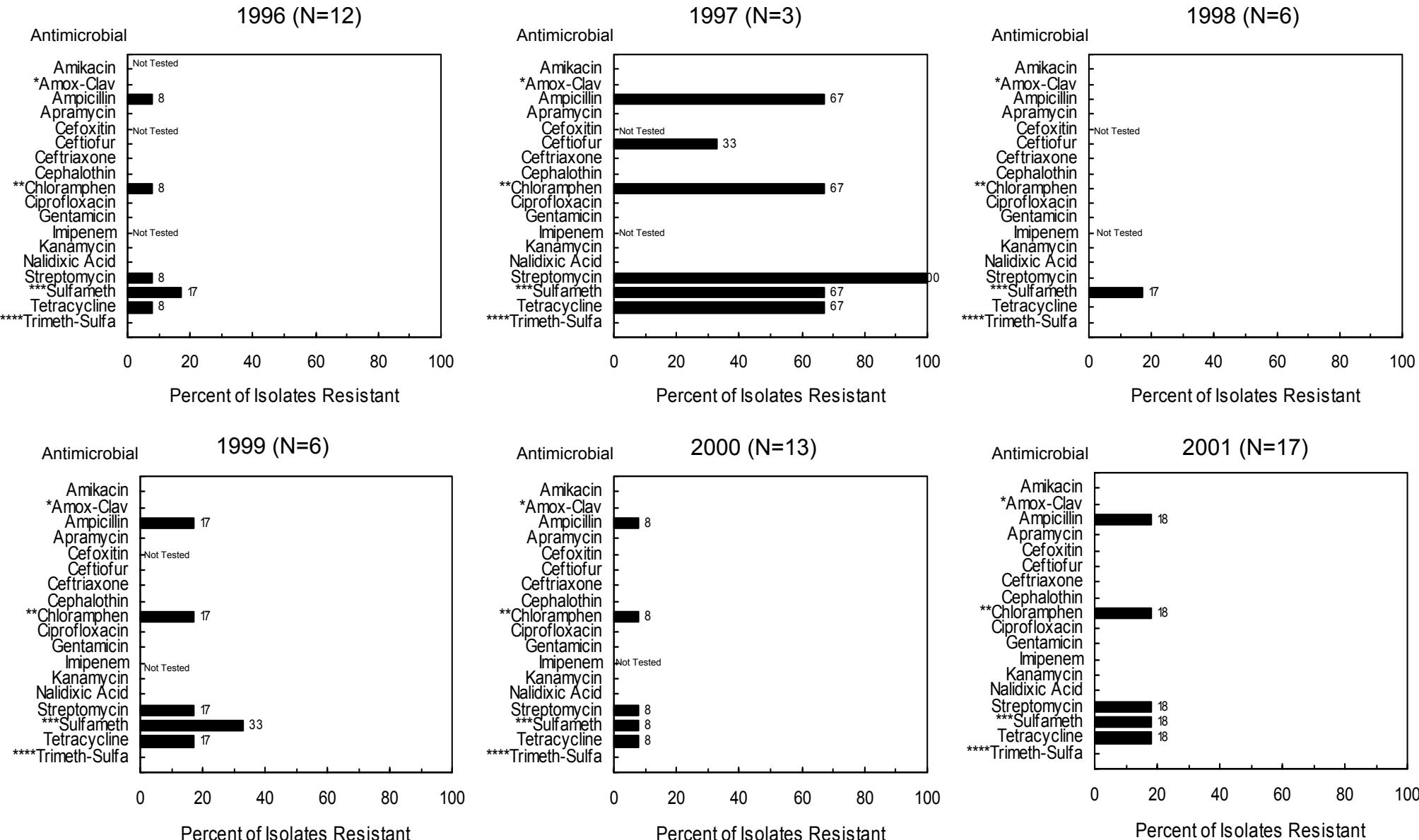
Figure 5f. Resistance among *Salmonella* serotype *Infantis* isolates, 1996-2001



*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

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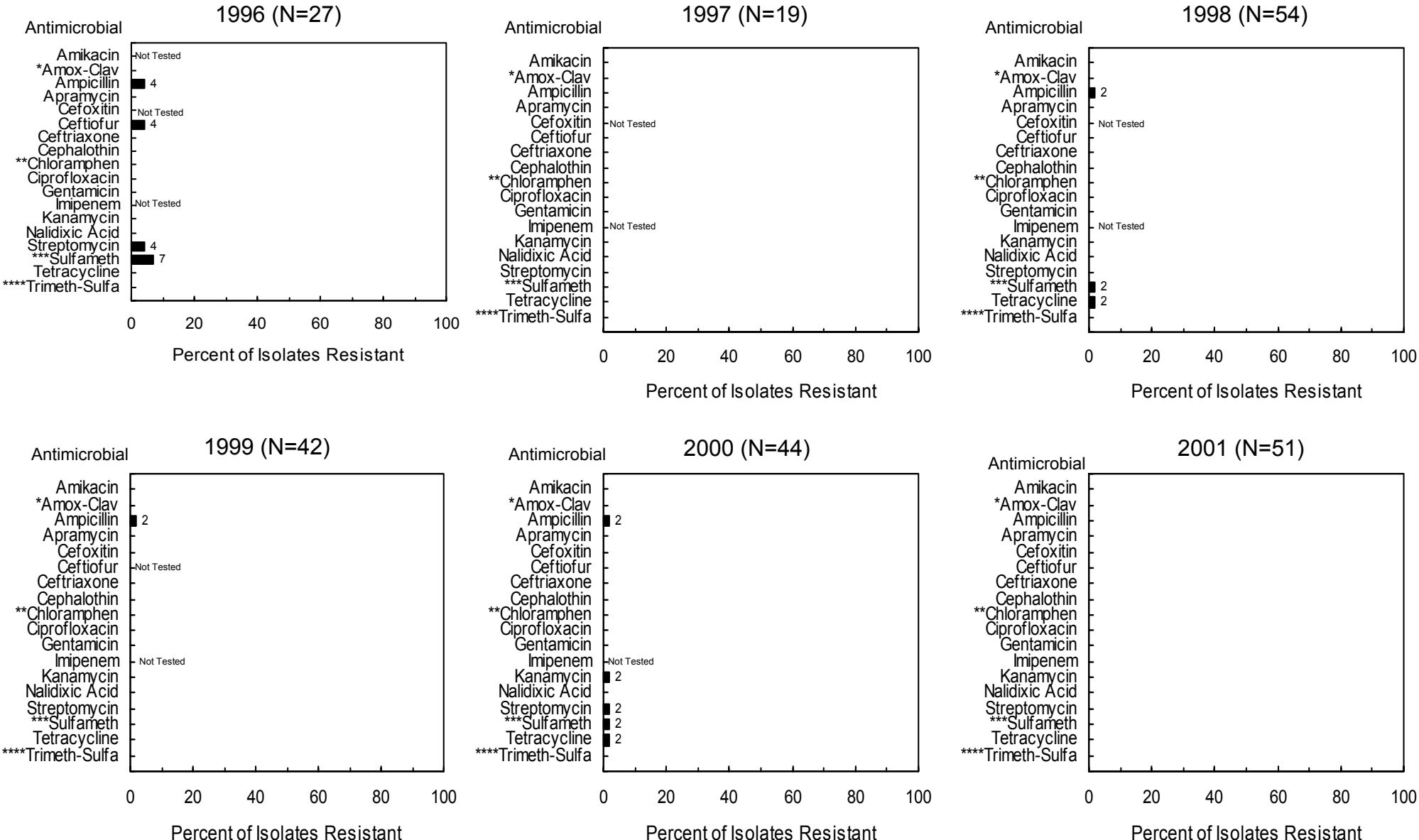
Figure 5g. Resistance among *Salmonella* serotype Java isolates, 1996-2001



*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

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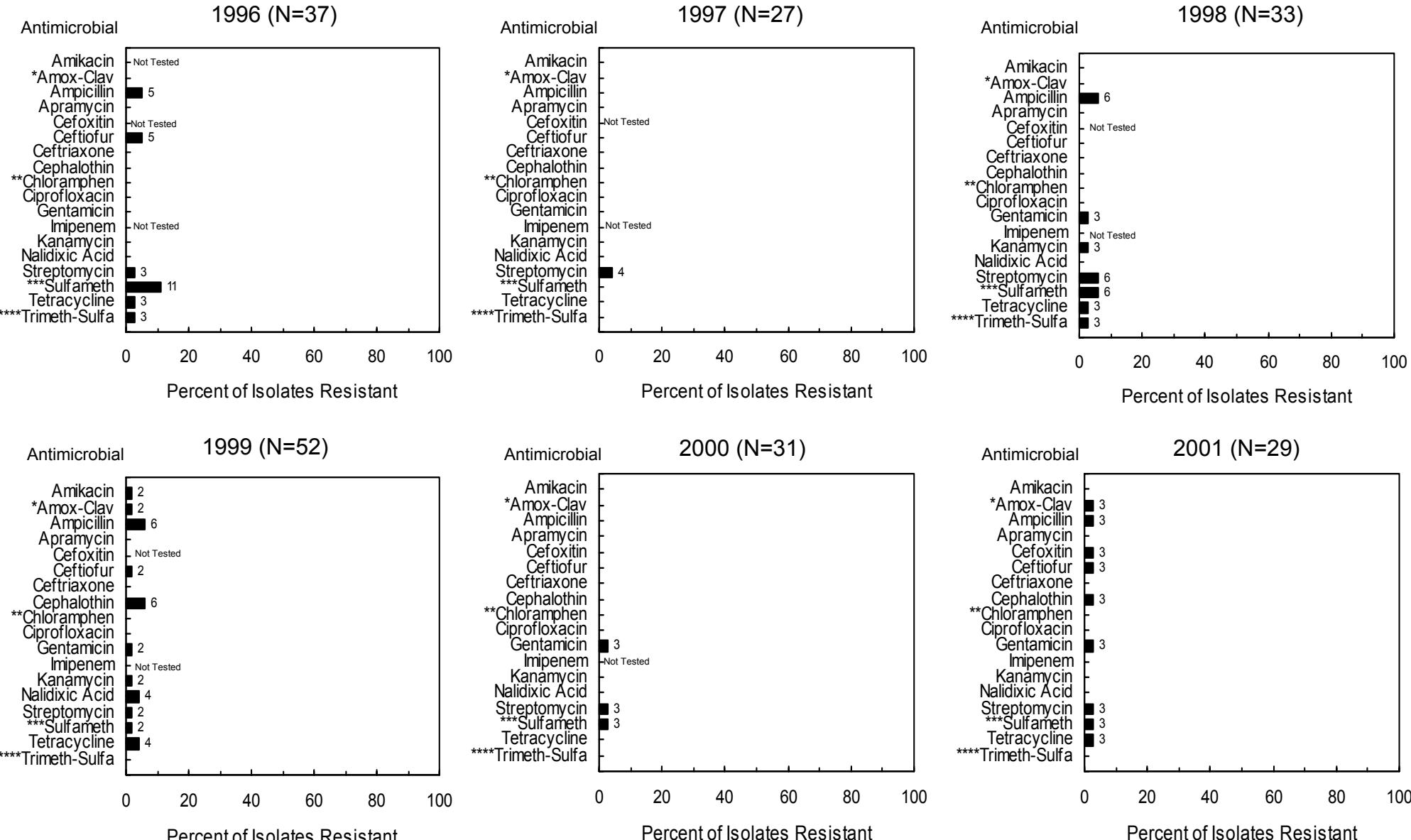
Figure 5h. Resistance among *Salmonella* serotype Javiana isolates, 1996-2001



*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

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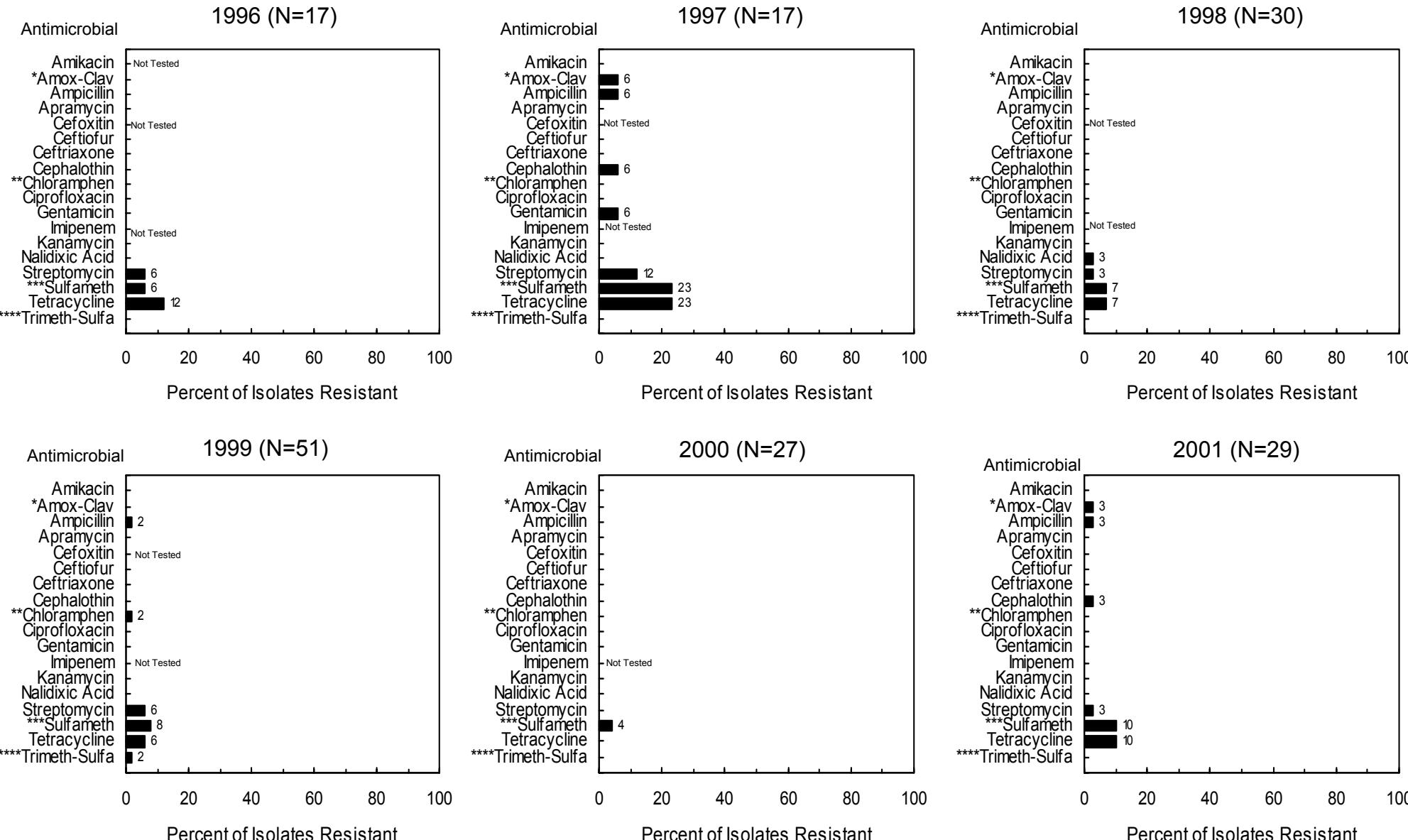
Figure 5i. Resistance among *Salmonella* serotype Montevideo isolates, 1996-2001



*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

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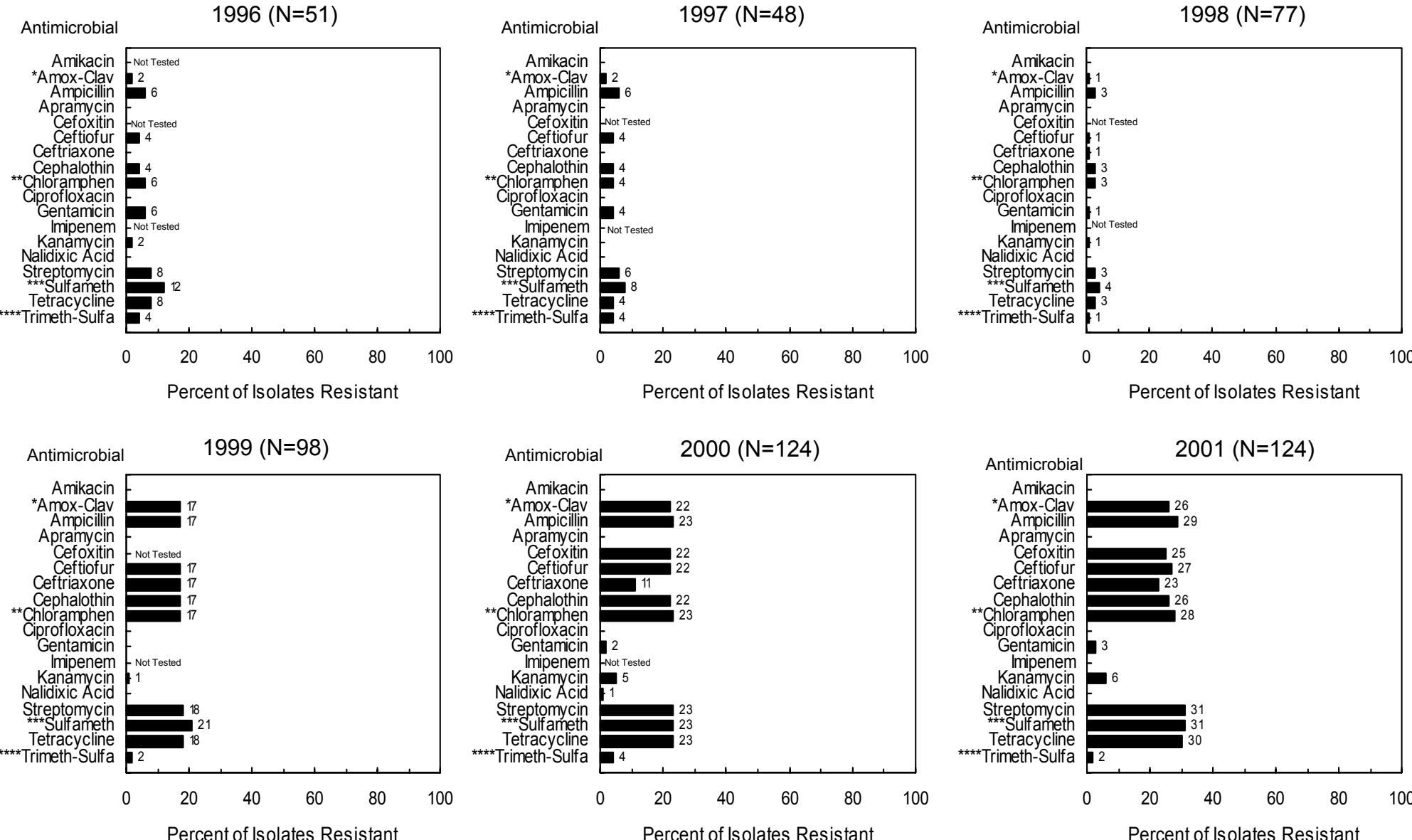
Figure 5j. Resistance among *Salmonella* serotype Muenchen isolates, 1996-2001



*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

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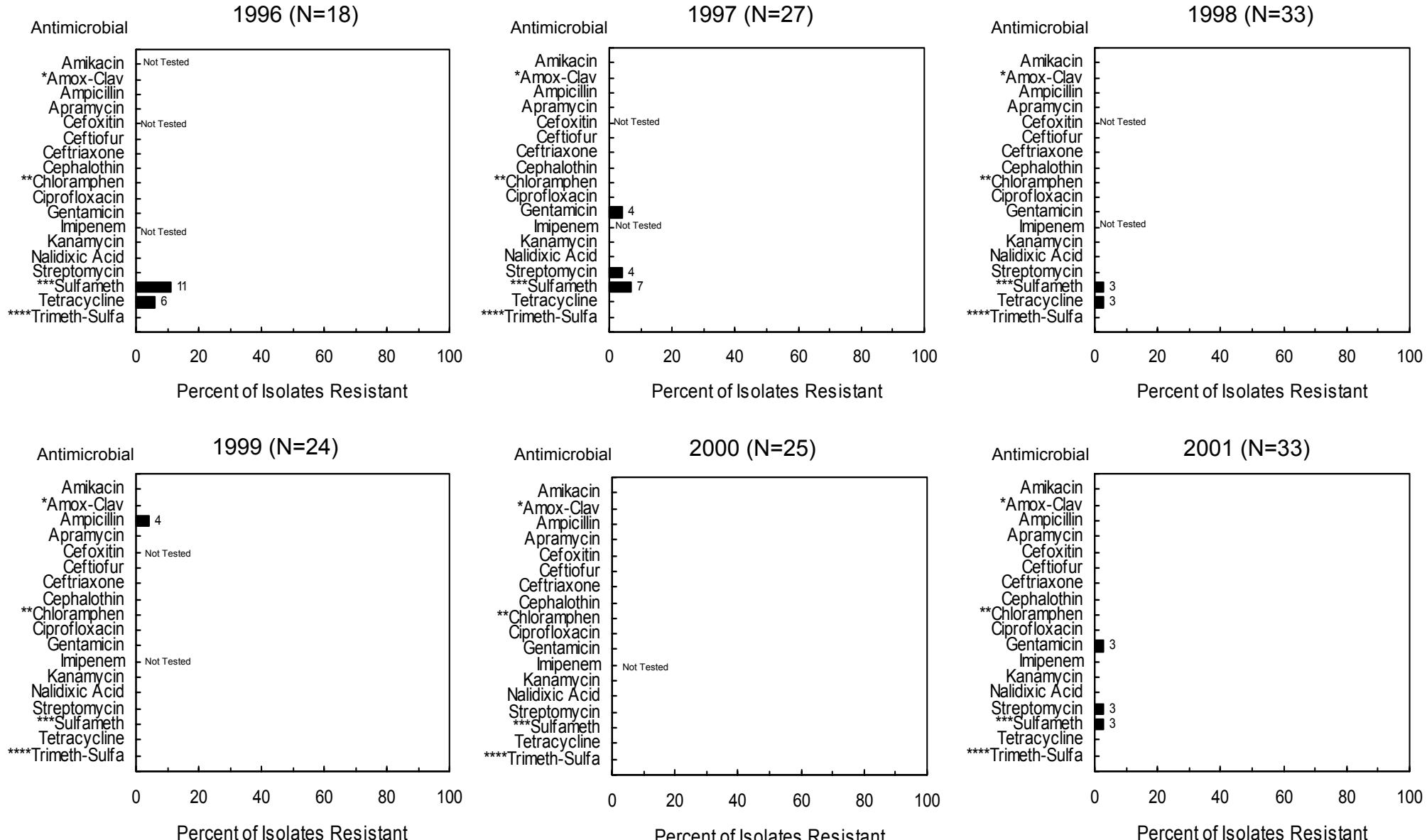
Figure 5k. Resistance among *Salmonella* serotype Newport isolates, 1996-2001



*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

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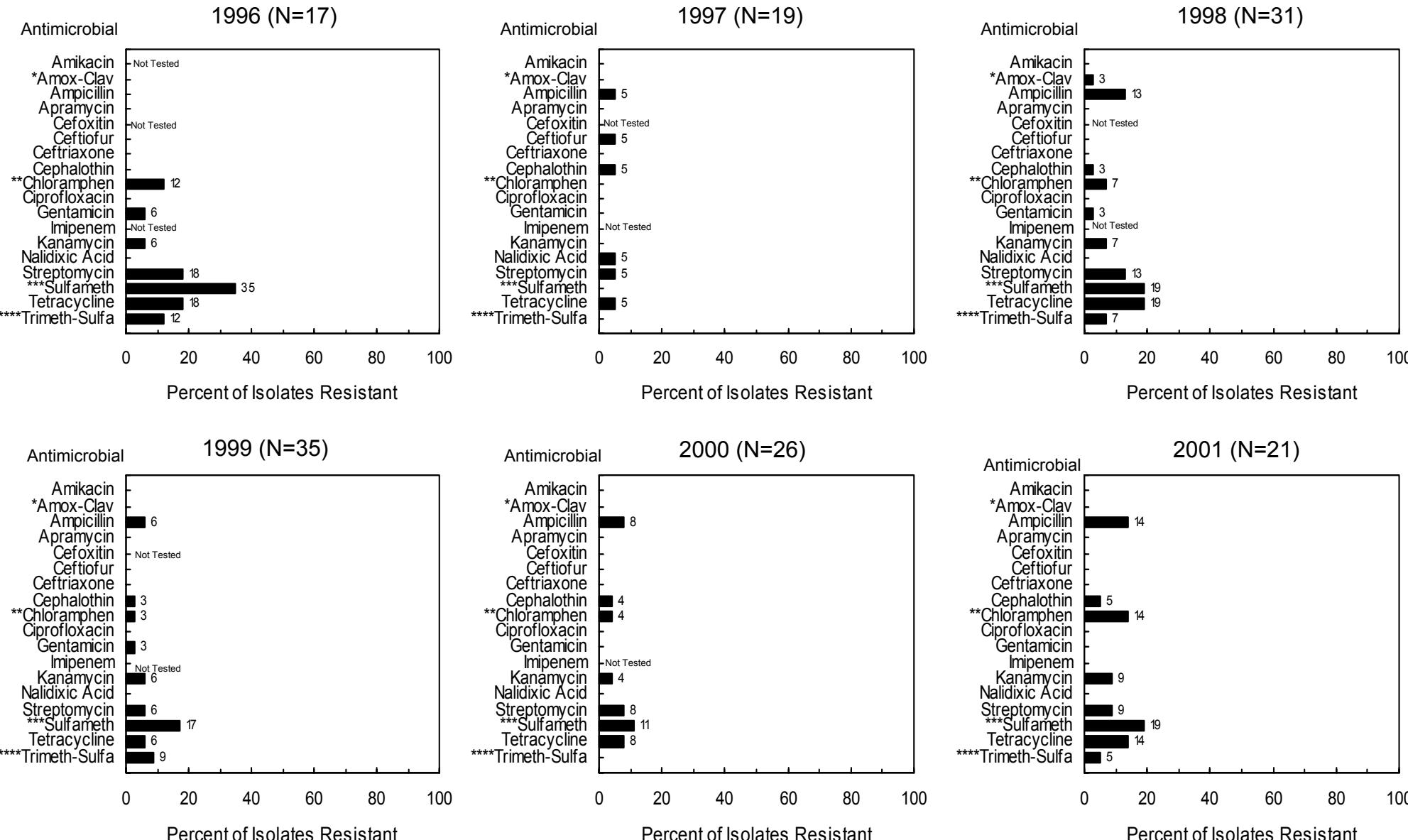
Figure 5l. Resistance among *Salmonella* serotype Oranienburg isolates, 1996-2001



*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

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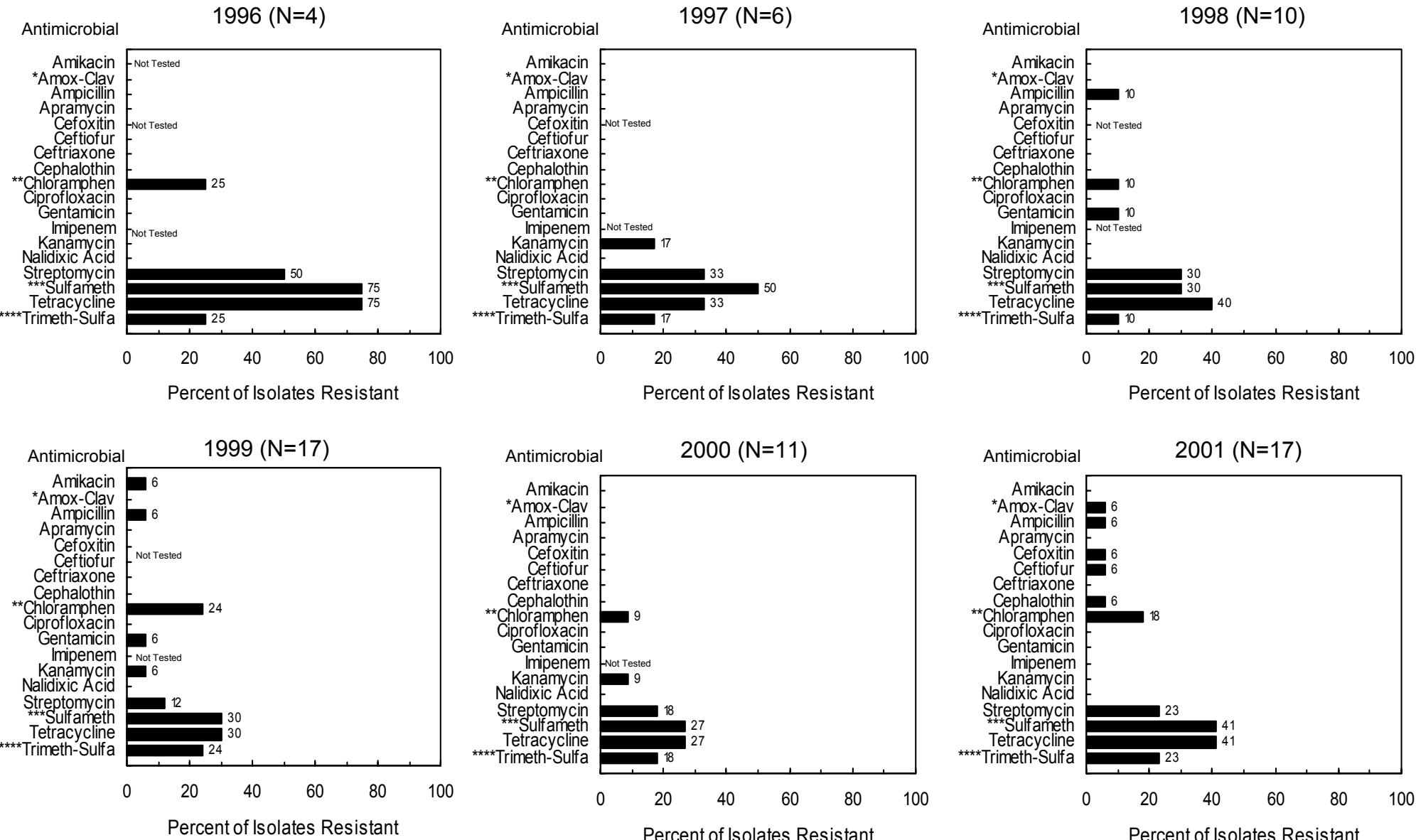
Figure 5m. Resistance among *Salmonella* serotype Saint Paul isolates, 1996-2001



*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

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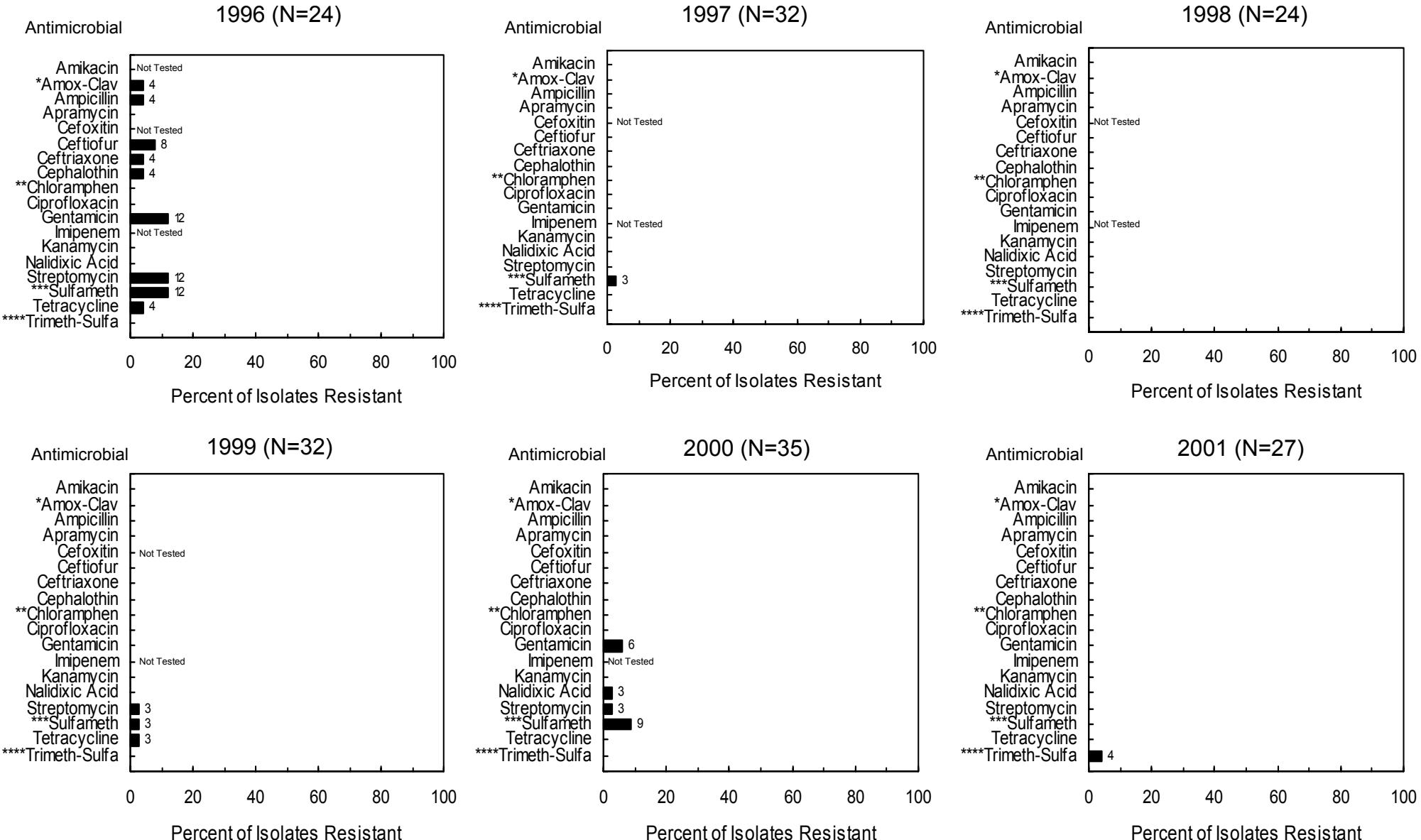
Figure 5n. Resistance among *Salmonella* serotype Stanley isolates, 1996-2001



*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

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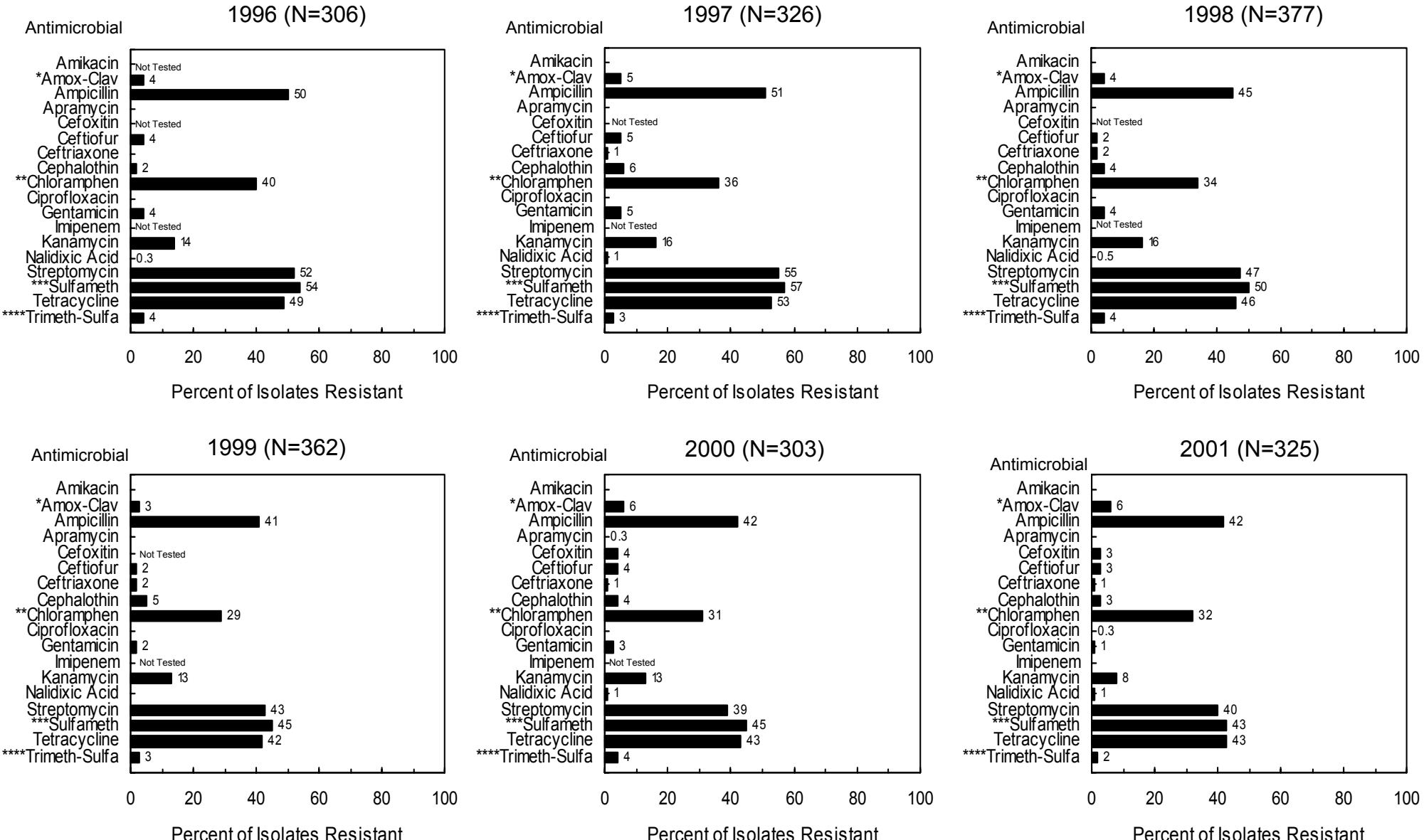
Figure 5o. Resistance among *Salmonella* serotype Thompson isolates, 1996-2001



*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

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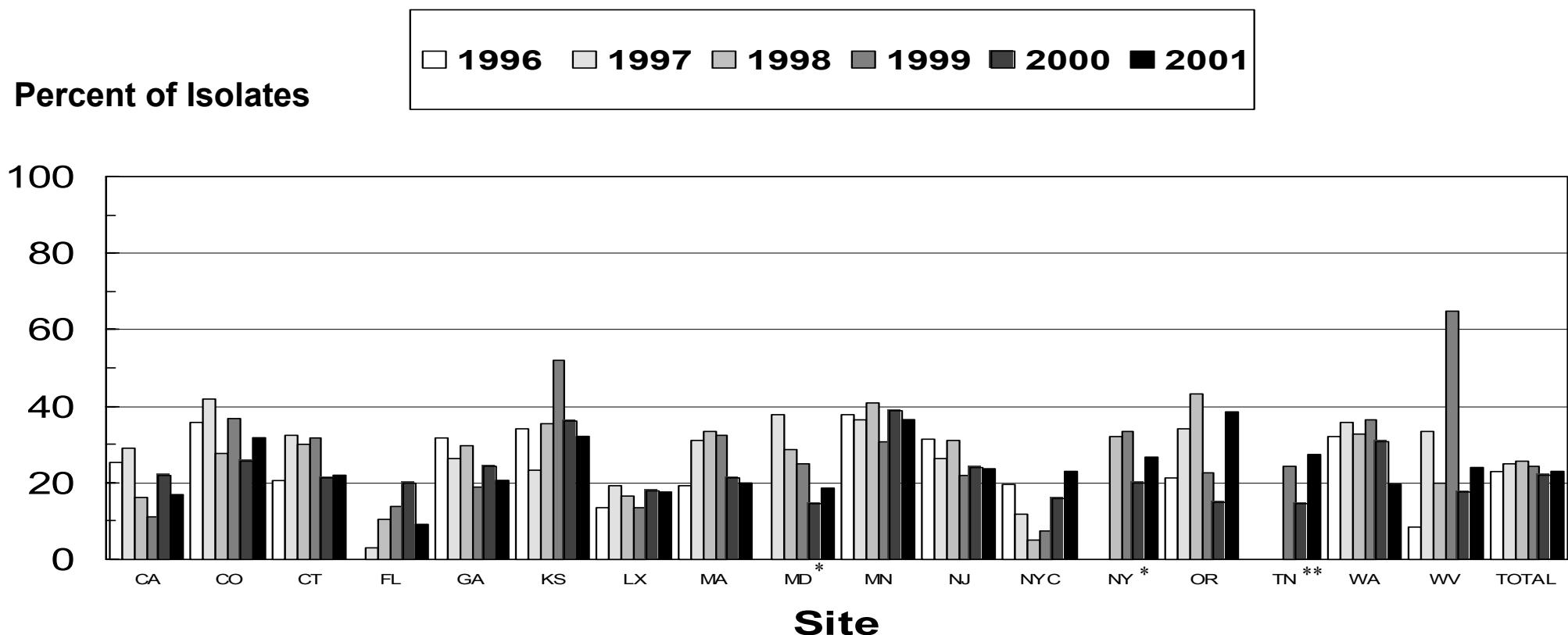
Figure 5p. Resistance among *Salmonella* serotype Typhimurium isolates, 1996-2001



*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

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Figure 6. Percent of non-Typhi *Salmonella* isolates that are serotype Typhimurium, by site, 1996-2001



Percent Typhimurium for all sites:

$$1996 - 306 / 1326 = 23\% \quad 1997 - 326 / 1301 = 25\% \quad 1998 - 377 / 1465 = 26\%$$

$$1999 - 362 / 1498 = 24\% \quad 2000 - 303 / 1378 = 22\% \quad 2001 - 325 / 1419 = 23\%$$

CA=Alameda, Contra Costa, and San Francisco counties

LX=Los Angeles County

NY=excluding New York City

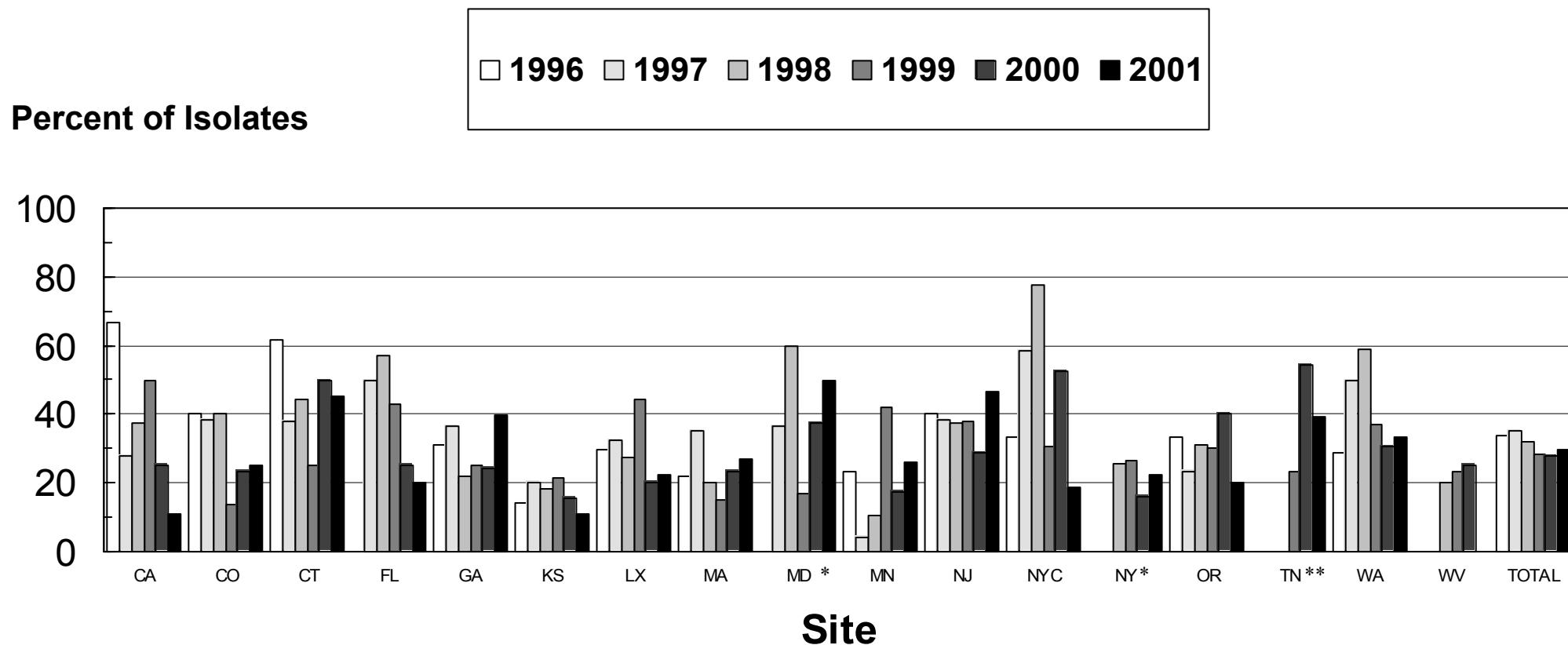
NYC>New York City

* Joined NARMS in 1997

** Joined NARMS in 1999

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Figure 7. Percent of *Salmonella* Typhimurium isolates that are resistant to at least ampicillin, chloramphenicol, streptomycin, sulfamethoxazole, and tetracycline (ACSSuT), by site, 1996–2001



Percent Typhimurium with at least ACSSuT pattern for all sites:

1996 - 103/306 = 34% 1997 - 115/326 = 35% 1998 - 120/377 = 32%

1999 - 102/362 = 28% 2000 - 84/303 = 28% 2001 - 96/325 = 29%

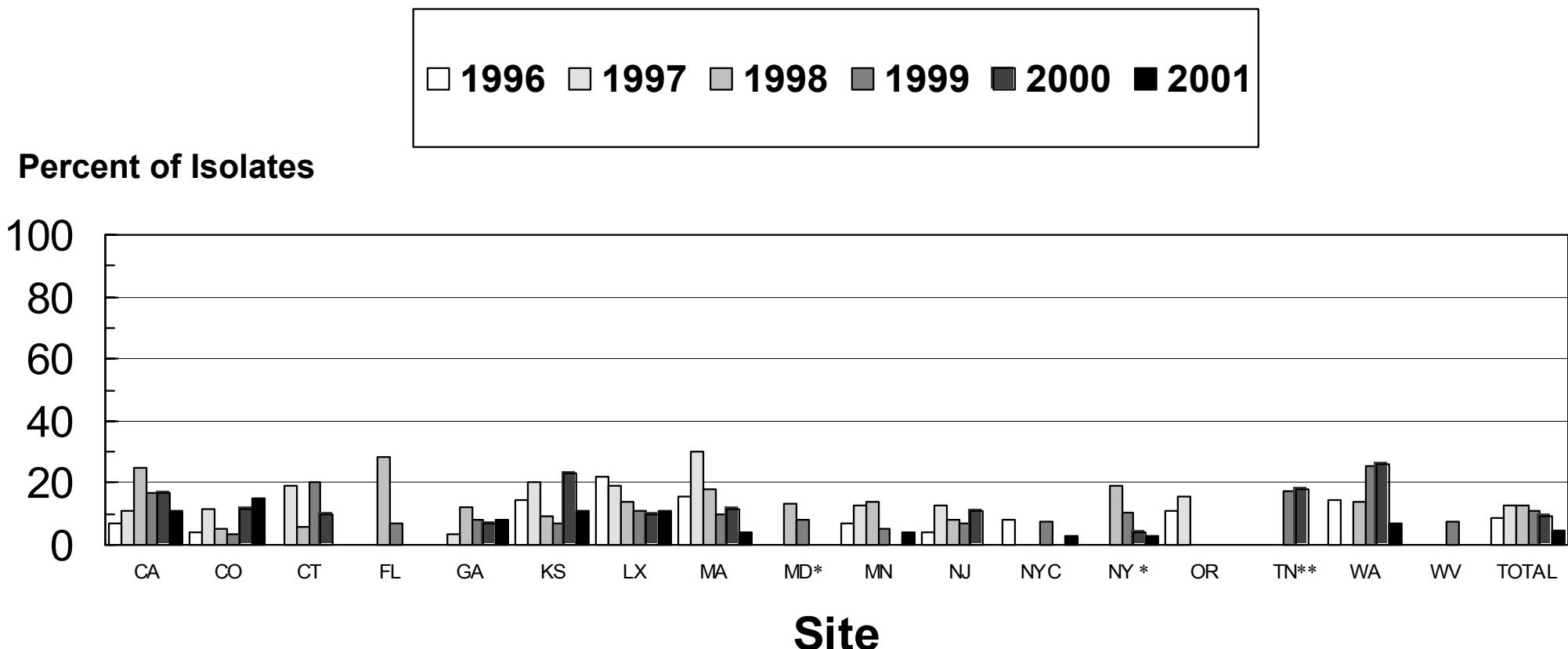
CA=Alameda, Contra Costa, and San Francisco counties
 LX=Los Angeles County
 NY=excluding New York City
 NYC>New York City

* Joined NARMS in 1997

** Joined NARMS in 1999

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Figure 8. Percent of *Salmonella* Typhimurium isolates that are resistant to at least ampicillin, kanamycin, streptomycin, sulfamethoxazole, and tetracycline (AKSSuT), by site, 1996-2001



Percent Typhimurium with at least AKSSuT pattern for all sites:

1996 - 27/306 = 9% 1997 - 41/326 = 13% 1998 - 47/377 = 12%

1999 - 39/362 = 11% 2000 - 28/303 = 9% 2001 - 15/325 = 5%

CA=Alameda, Contra Costa, and San Francisco counties

LX=Los Angeles County

NY=excluding New York City

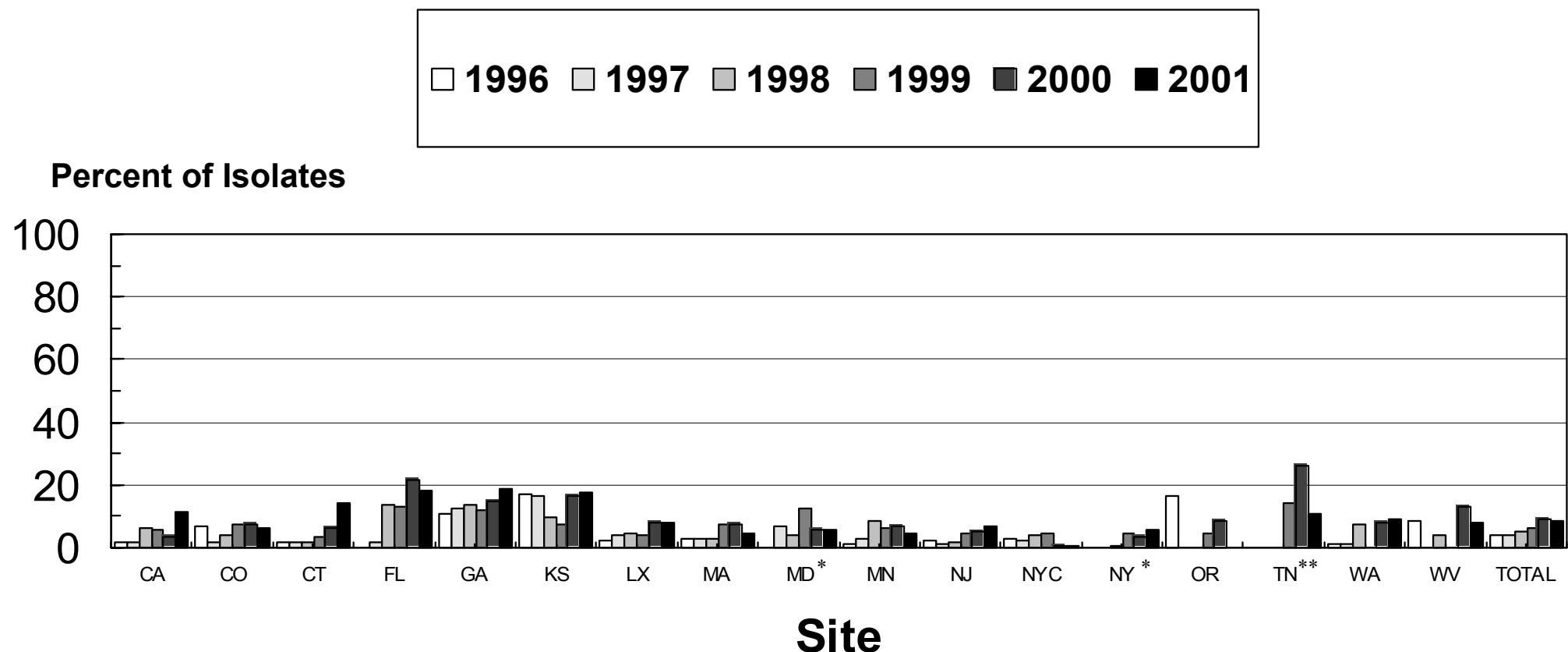
NYC>New York City

* Joined NARMS in 1997

** Joined NARMS in 1999

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Figure 9. Percent of non-Typhi *Salmonella* isolates that are serotype Newport, by site, 1996-2001



Percent Newport for all sites

1996 - 51/1326 = 4% 1997 - 48/1301 = 4% 1998 - 77/1465= 5%

1999 - 98/1498 = 7% 2000 - 124/1378 = 9% 2001 - 124/1419 = 9%

CA=Alameda, Contra Costa, and San Francisco counties

LX=Los Angeles County

NY=excluding New York City

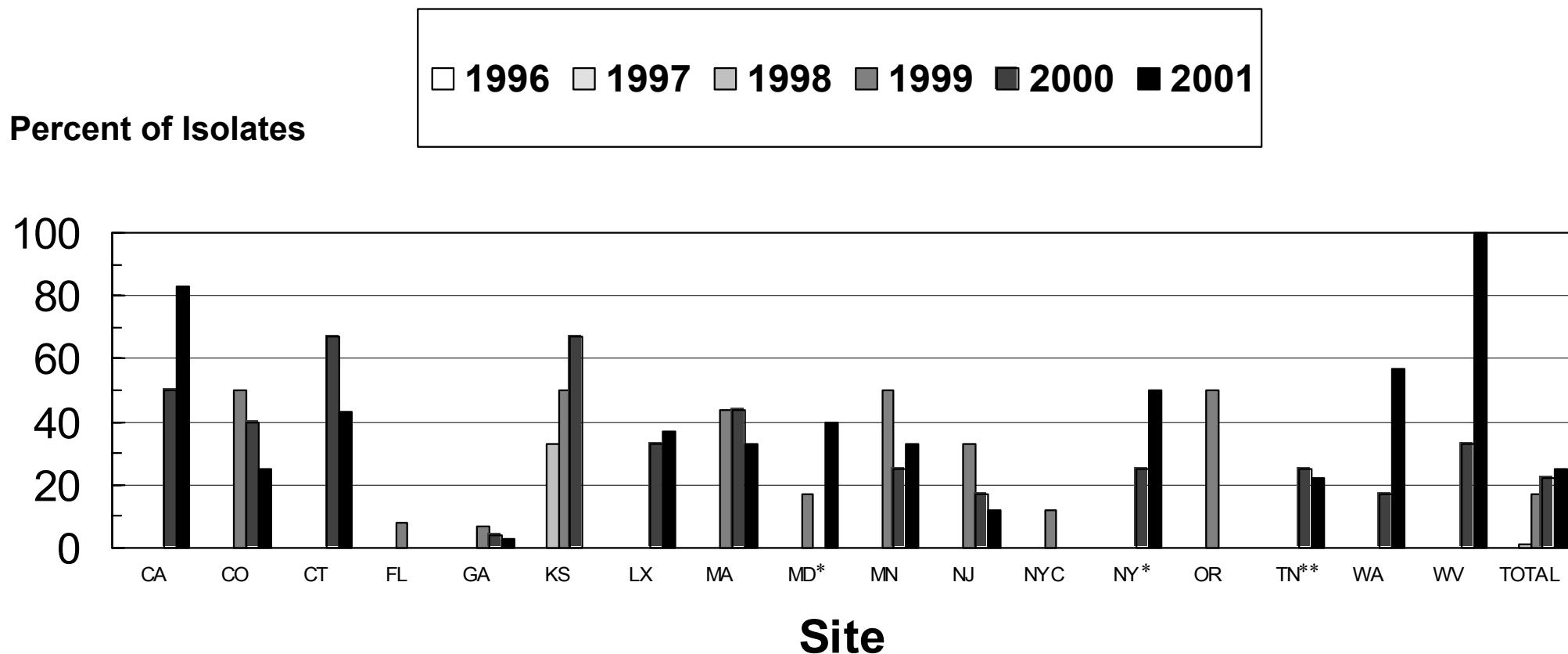
NYC>New York City

* Joined NARMS in 1997

** Joined NARMS in 1999

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Figure 10. Percent of *Salmonella* Newport isolates that are at least MDR-AmpC[‡], by site, 1996-2001



[‡]MDR-AmpC=Resistance to Amoxicillin/Clavulanic Acid, Ampicillin, Cephalothin, Cefoxitin, Ceftiofur, Chloramphenicol, Sulfamethoxazole, Streptomycin and Tetracycline, and Decreased Susceptibility to Ceftriaxone (MIC \geq 16 μ g/ml)

Percent Newport with MDR-AmpC pattern for all sites:

1996 - 0/51 = 0% 1997 - 0/48 = 0% 1998 - 1/77 = 1%
1999 - 17/98 = 17% 2000 - 27/124 = 22% 2001 = 31/124 = 25%

CA=Alameda, Contra Costa, and San Francisco counties
LX=Los Angeles County

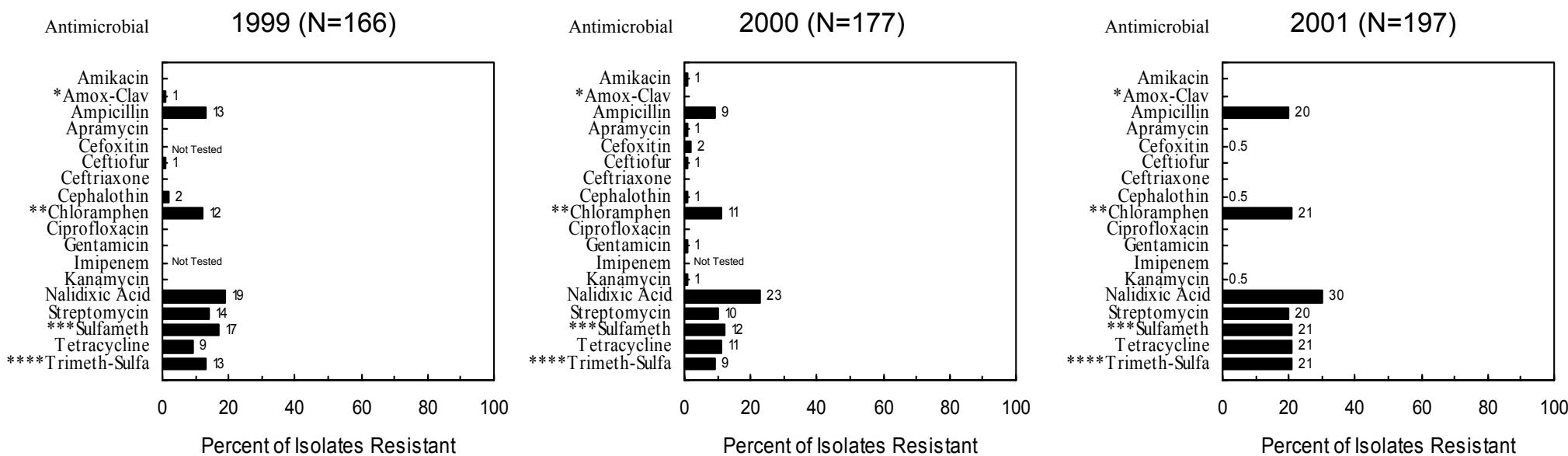
NY=excluding New York City
NYC>New York City

* Joined NARMS in 1997

** Joined NARMS in 1999

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Figure 11. Resistance among *Salmonella* Typhi isolates, 1999-2001

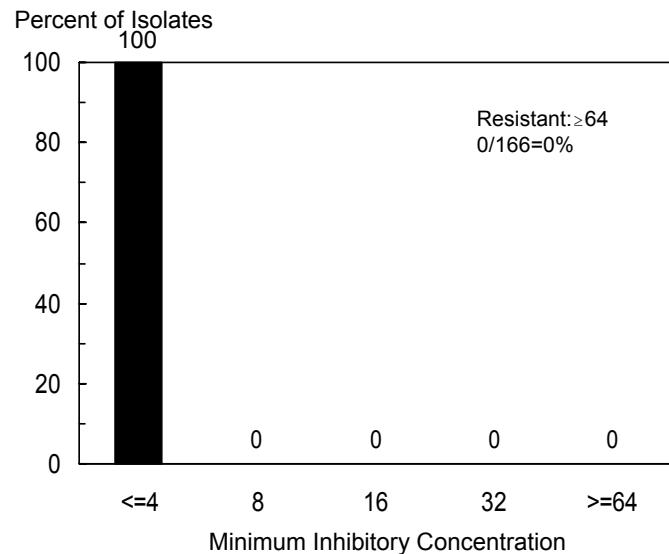


*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

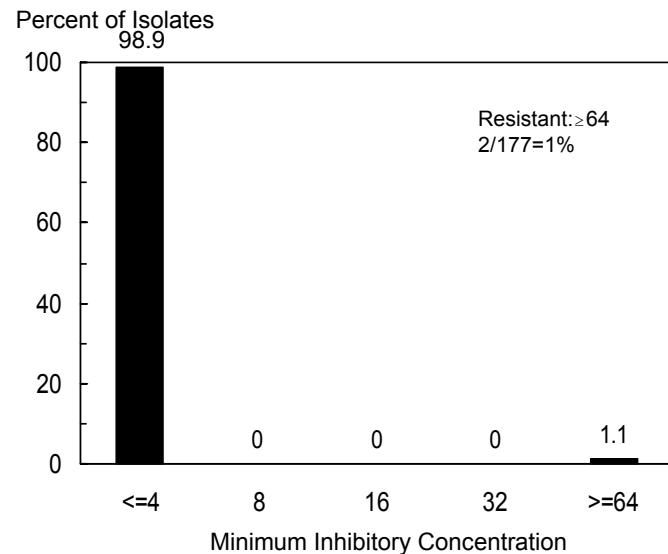
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**Figure 12a. MICs for amikacin among *Salmonella* Typhi isolates,
1999-2001**

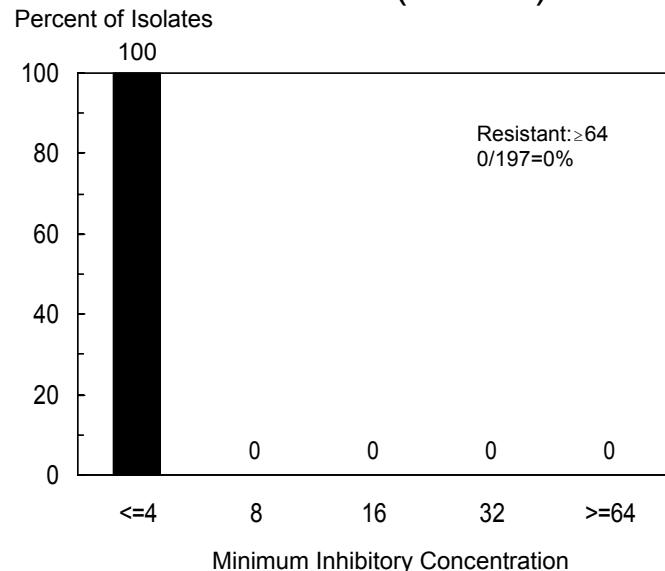
1999 (N=166)



2000 (N=177)



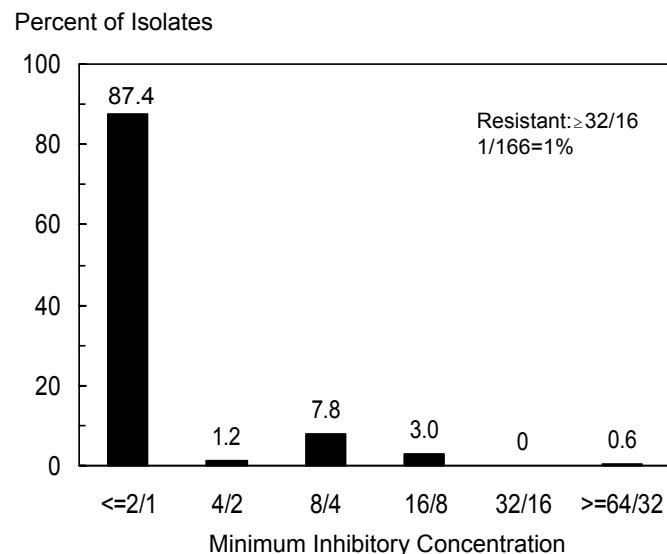
2001 (N=197)



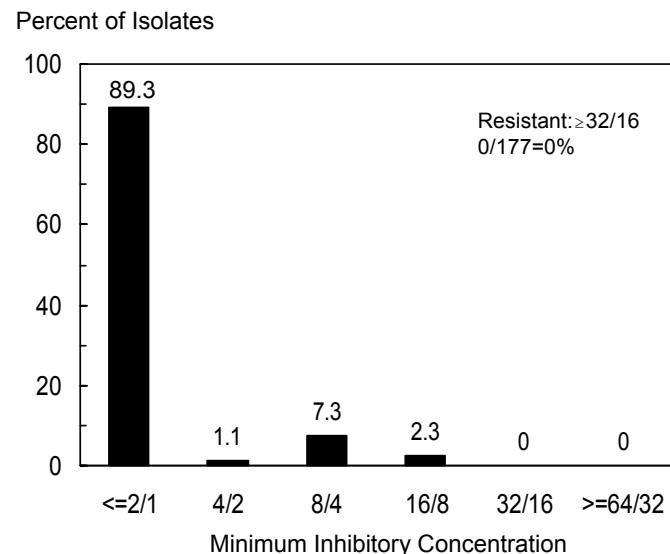
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Figure 12b. MICs for amoxicillin-clavulanic acid among *Salmonella* Typhi isolates, 1999-2001

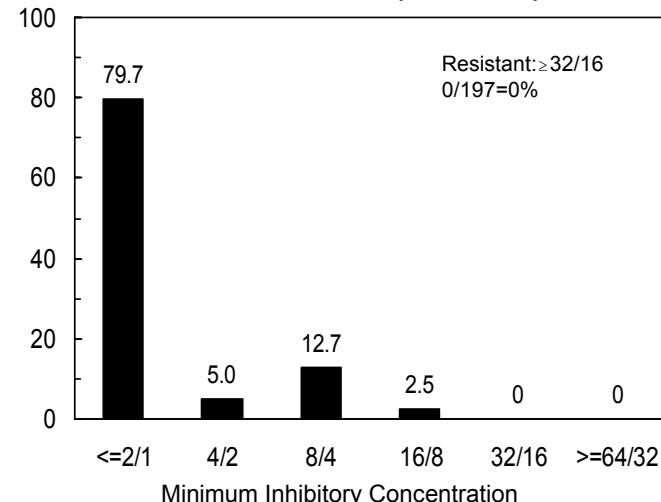
1999 (N=166)



2000 (N=177)



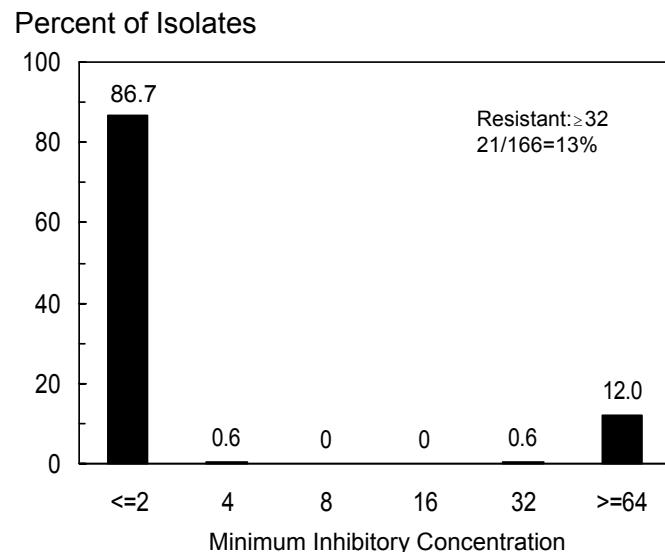
2001 (N=197)



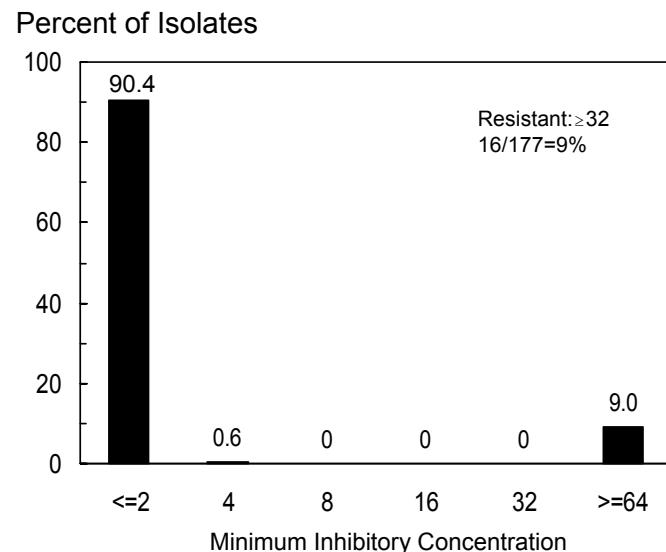
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Figure 12c. MICs for ampicillin among *Salmonella* Typhi isolates, 1999-2001

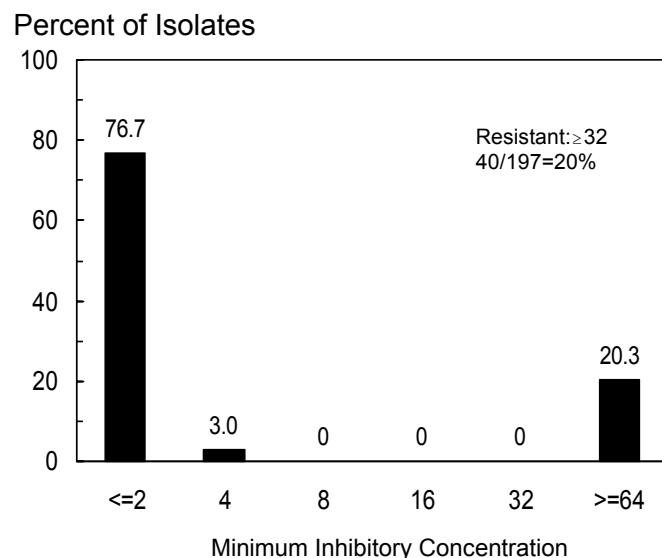
1999 (N=166)



2000 (N=177)



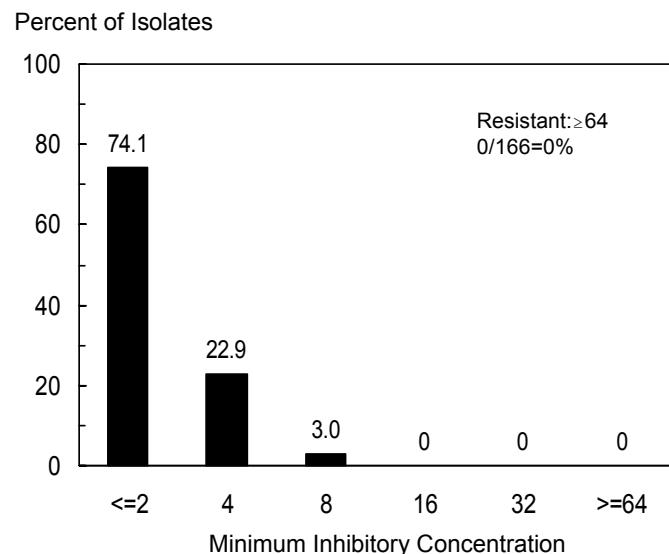
2001 (N=197)



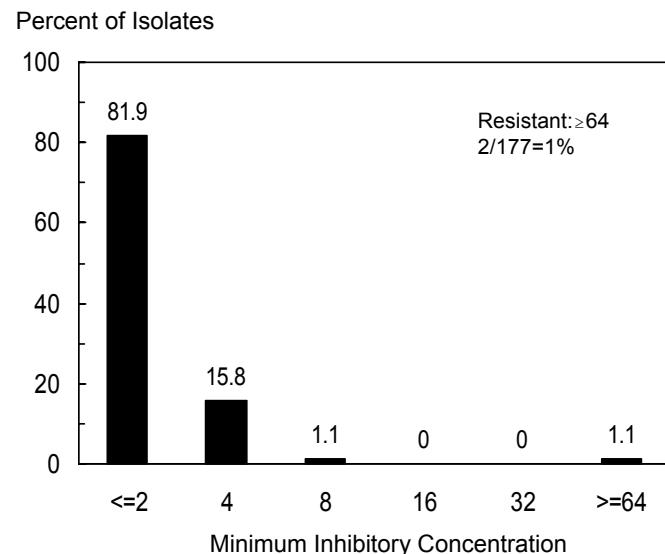
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Figure 12d. MICs for apramycin among *Salmonella* Typhi isolates, 1999-2001

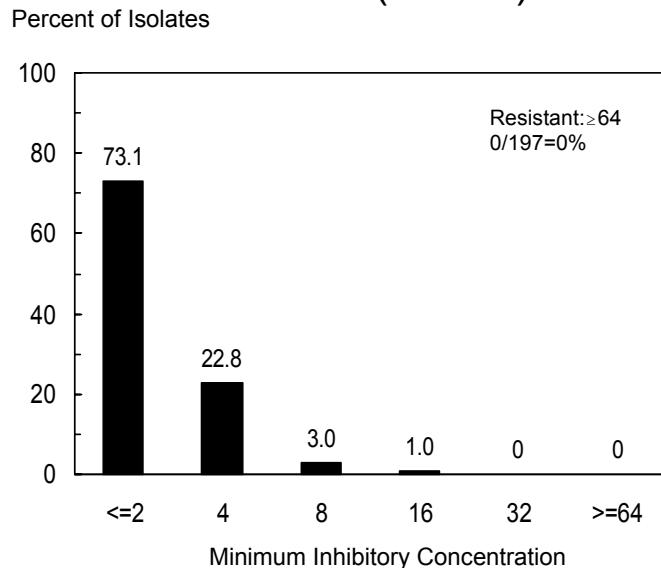
1999 (N=166)



2000 (N=177)



2001 (N=197)

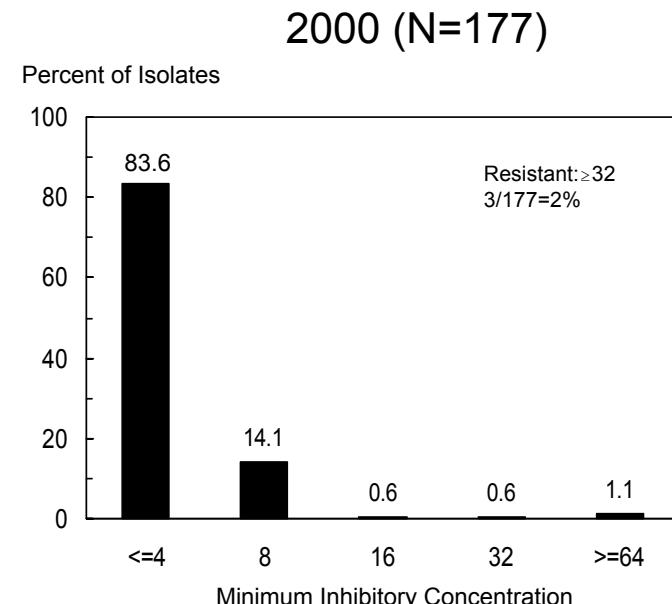
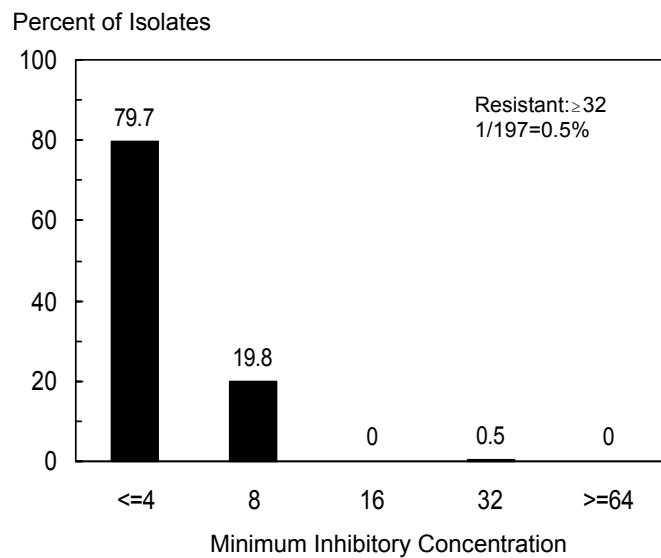


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Figure 12e. MICs for cefoxitin among *Salmonella* Typhi isolates, 1999-2001

Not Tested in 1999

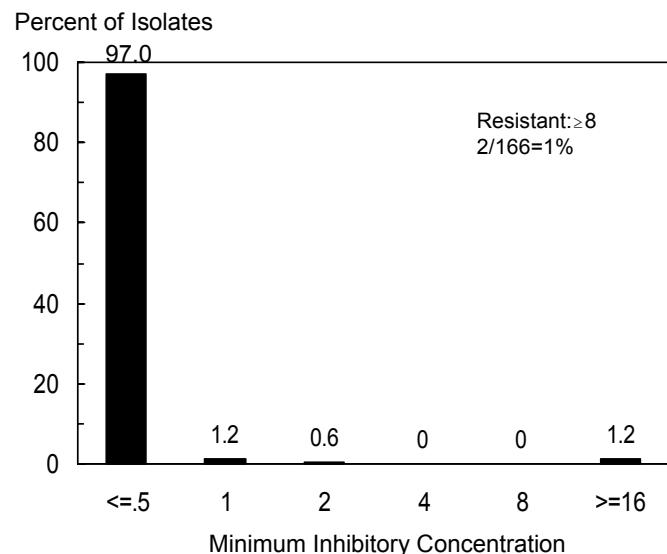
2001 (N=197)



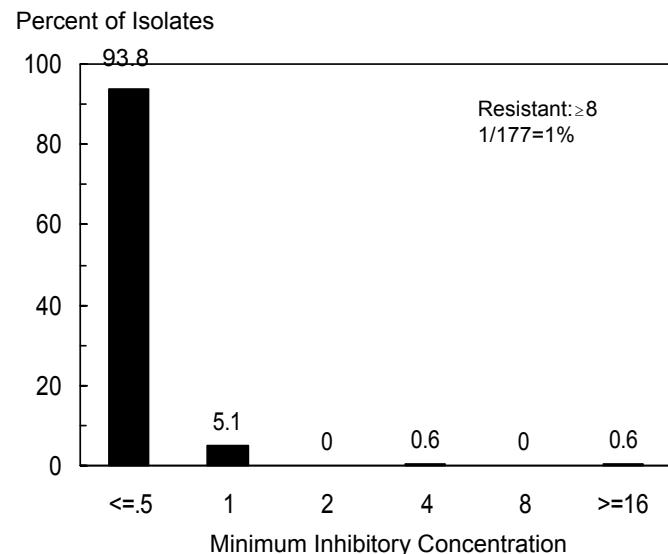
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Figure 12f. MICs for ceftiofur among *Salmonella* Typhi isolates, 1999-2001

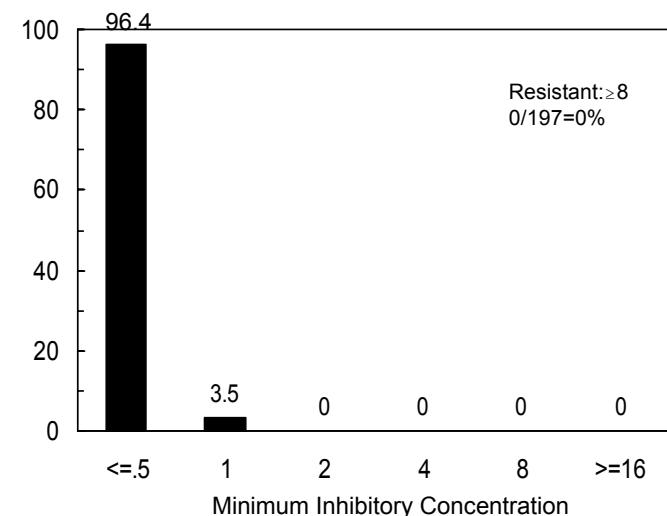
1999 (N=166)



2000 (N=177)



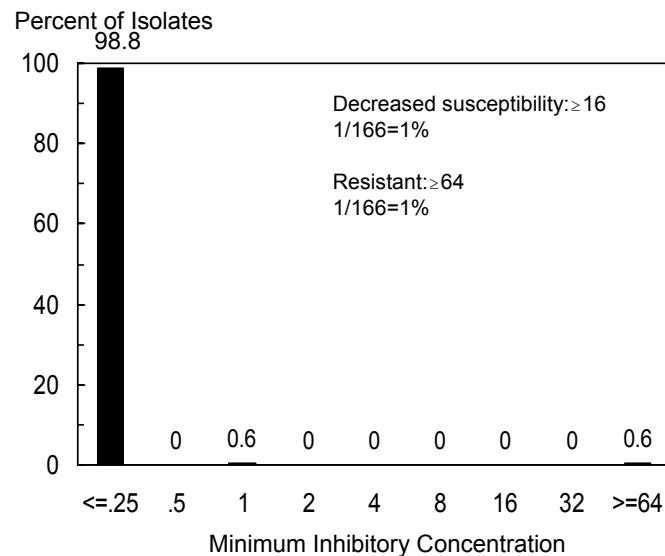
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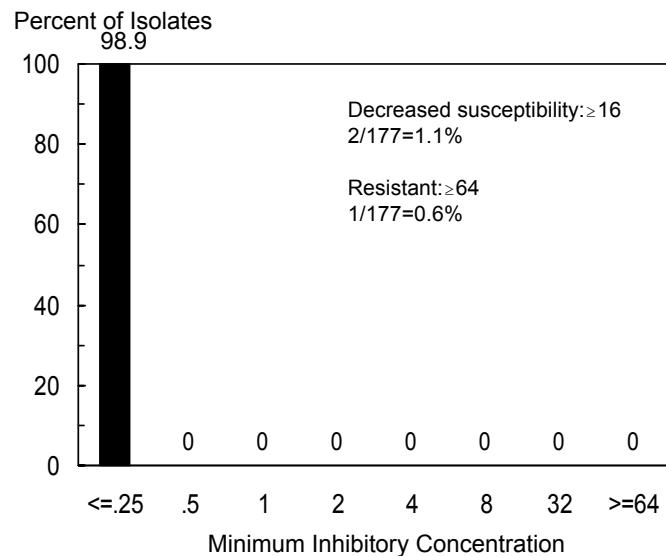
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Figure 12g. MICs for ceftriaxone* among *Salmonella* Typhi isolates, 1999-2001

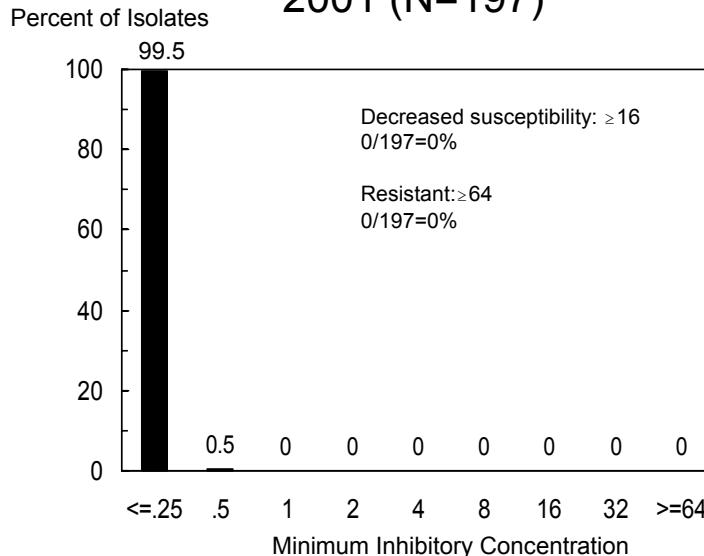
1999 (N=166)



2000 (N=177)



2001 (N=197)

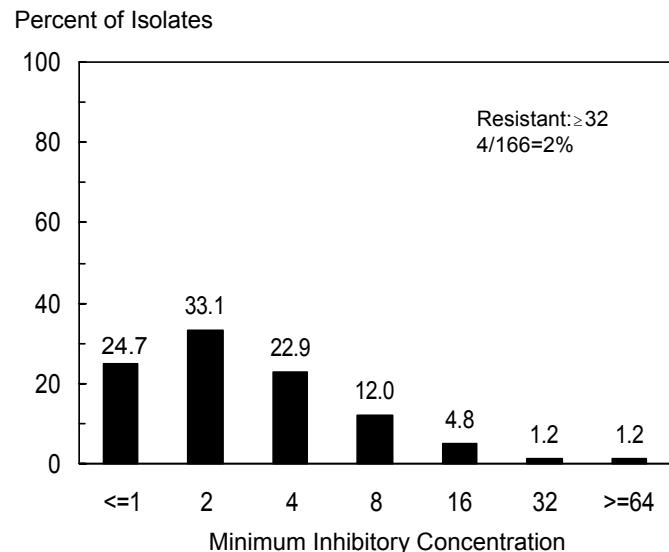


*Sensititre® results only

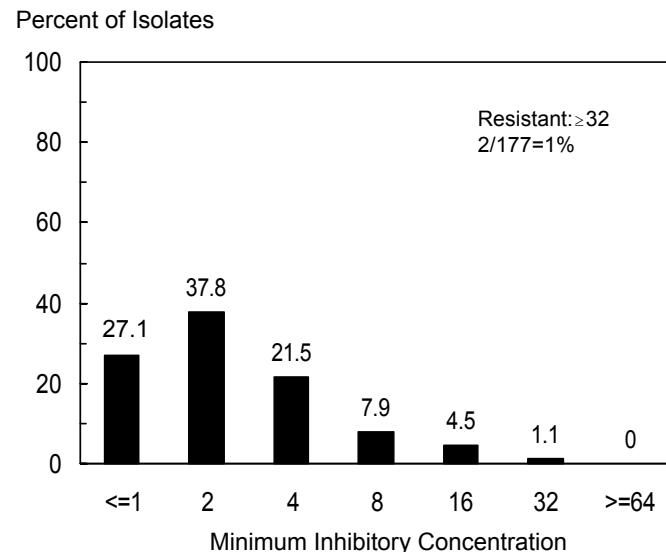
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Figure 12h. MICs for cephalothin among *Salmonella* Typhi isolates, 1999-2001

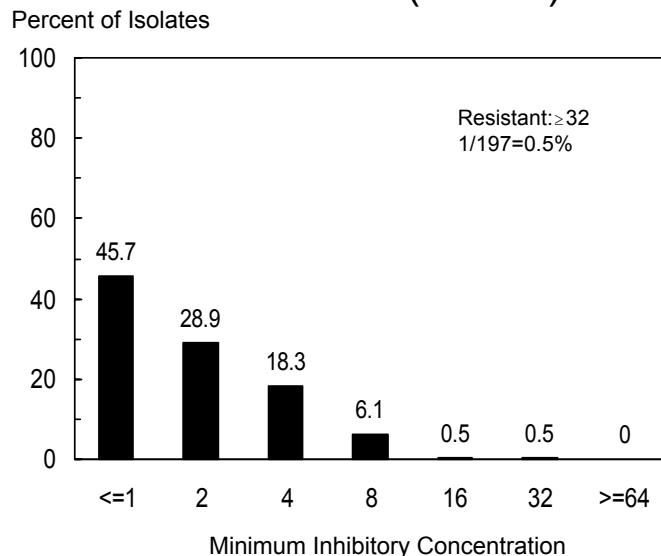
1999 (N=166)



2000 (N=177)



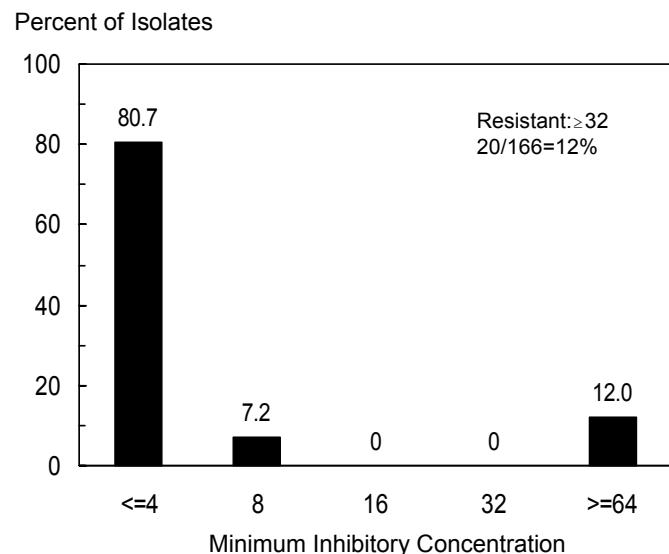
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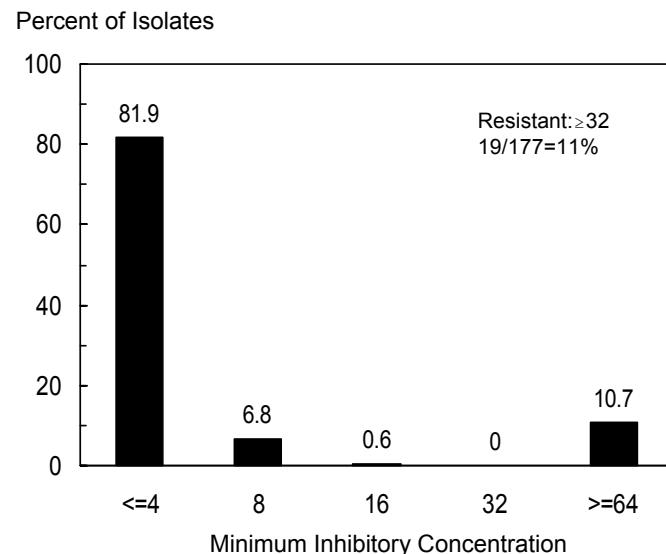
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Figure 12i. MICs for chloramphenicol among *Salmonella* Typhi Isolates, 1999-2001

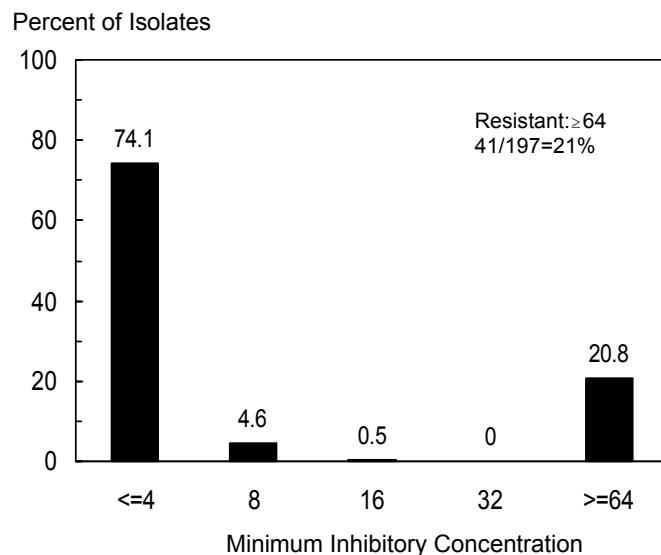
1999 (N=166)



2000 (N=177)



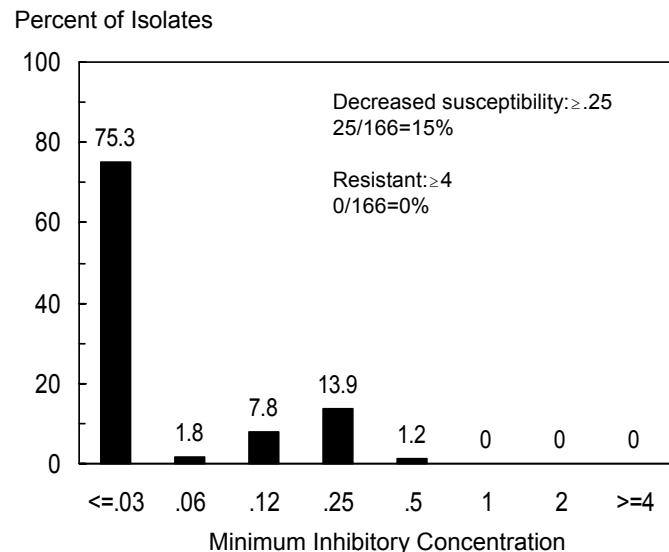
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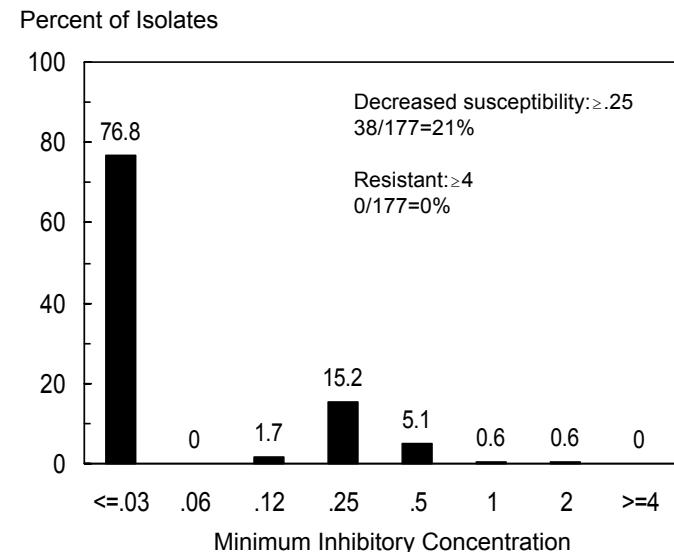
National Antimicrobial Resistance Monitoring System For Enteric Bacteria

**Figure 12j. MICs for ciprofloxacin among *Salmonella* Typhi isolates,
1999-2001**

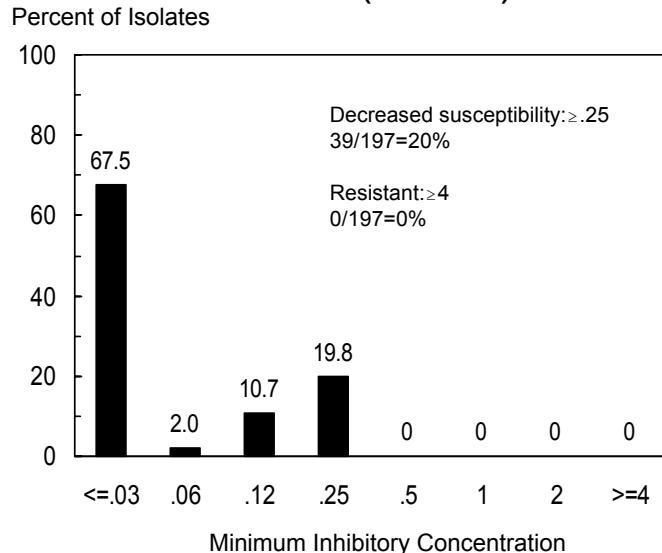
1999 (N=166)



2000 (N=177)



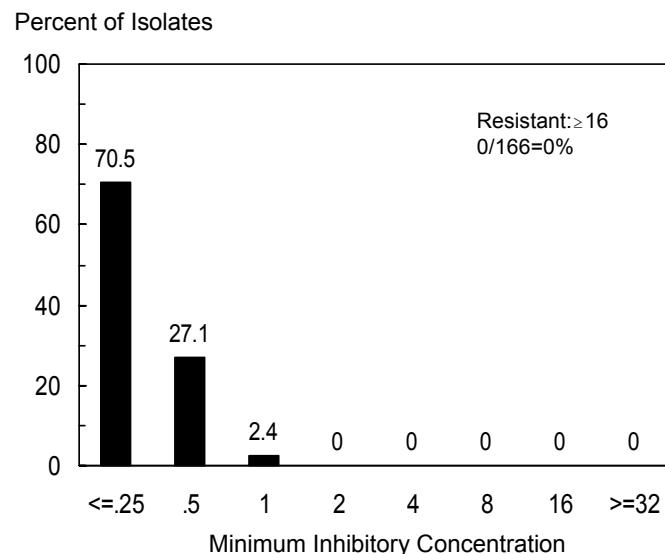
2001 (N=197)



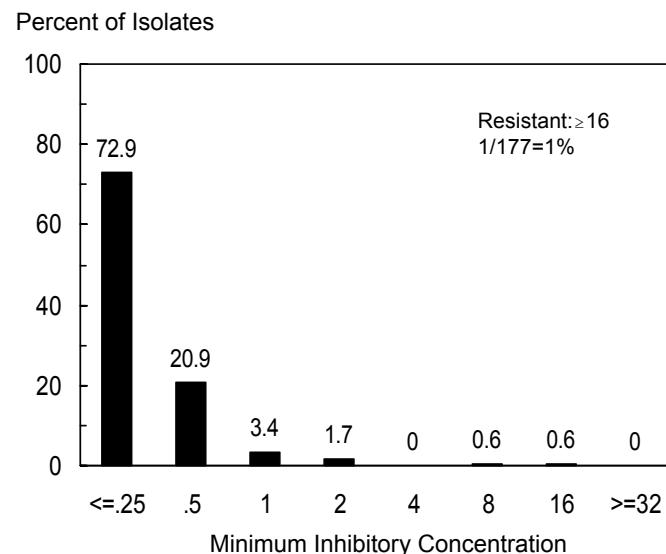
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**Figure 12k. MICs for gentamicin among *Salmonella* Typhi isolates,
1999-2001**

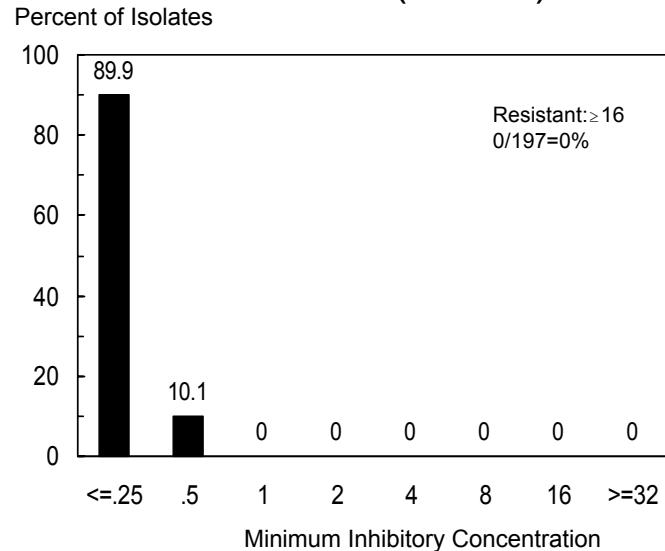
1999 (N=166)



2000 (N=177)



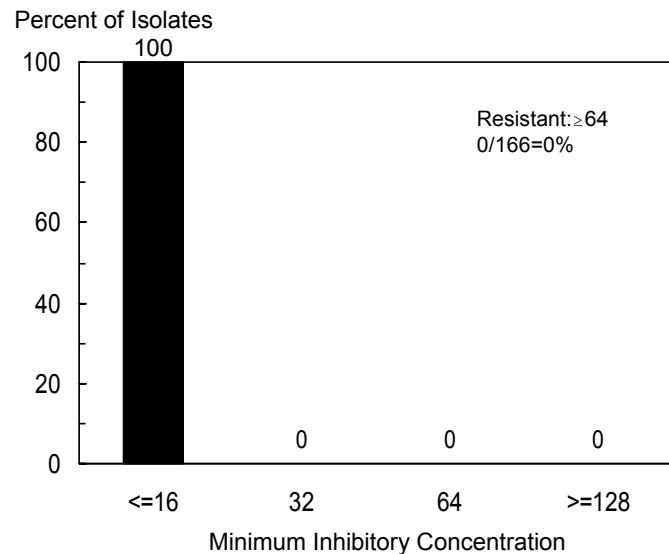
2001 (N=197)



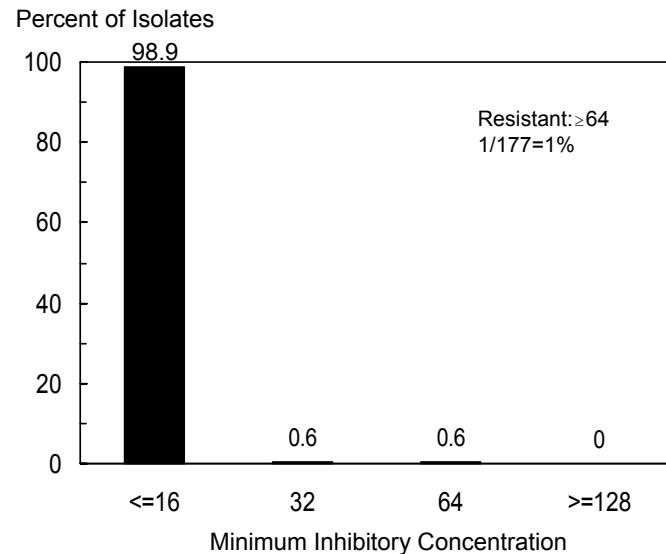
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Figure 12I. MICs for kanamycin among *Salmonella* Typhi isolates, 1999-2001

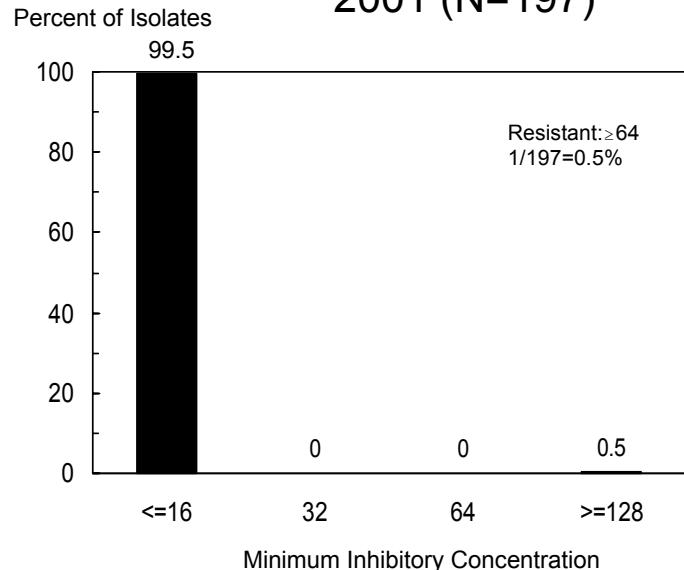
1999 (N=166)



2000 (N=177)



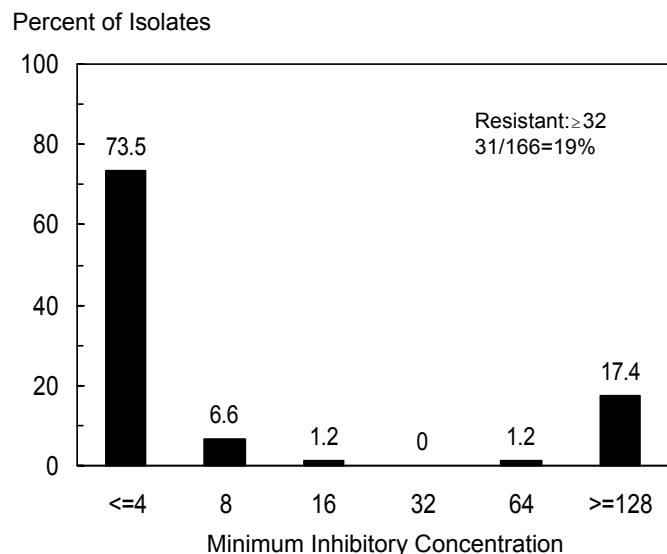
2001 (N=197)



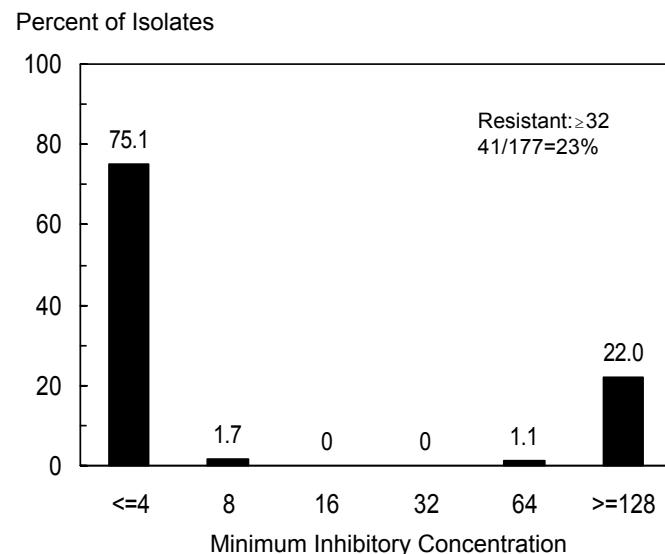
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Figure 12m. MICs for nalidixic acid among *Salmonella* Typhi isolates, 1999-2001

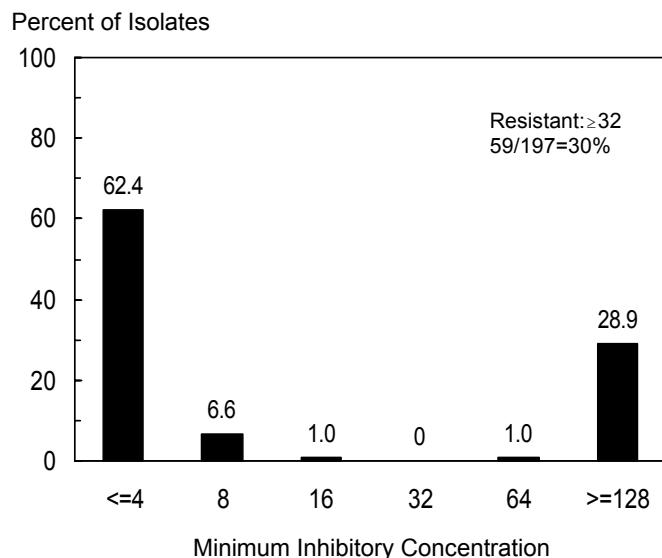
1999 (N=166)



2000 (N=177)



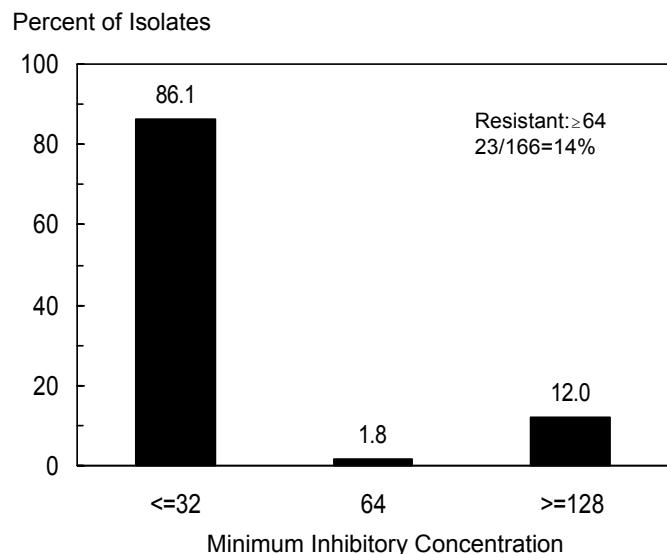
2001 (N=197)



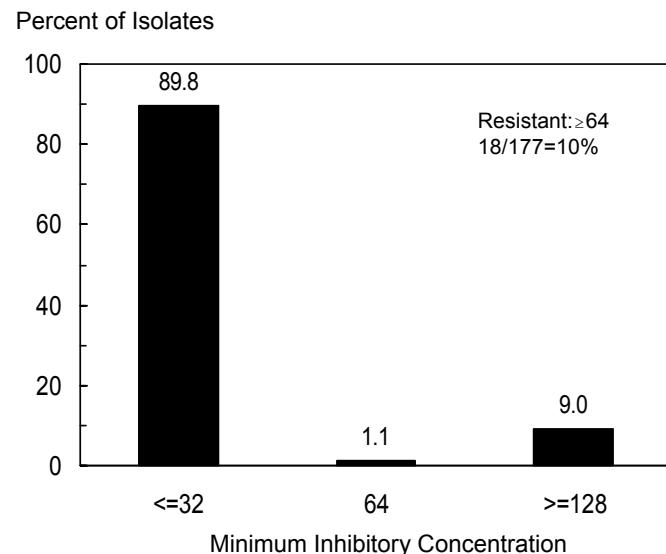
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Figure 12n. MICs for streptomycin among *Salmonella* Typhi isolates, 1999-2001

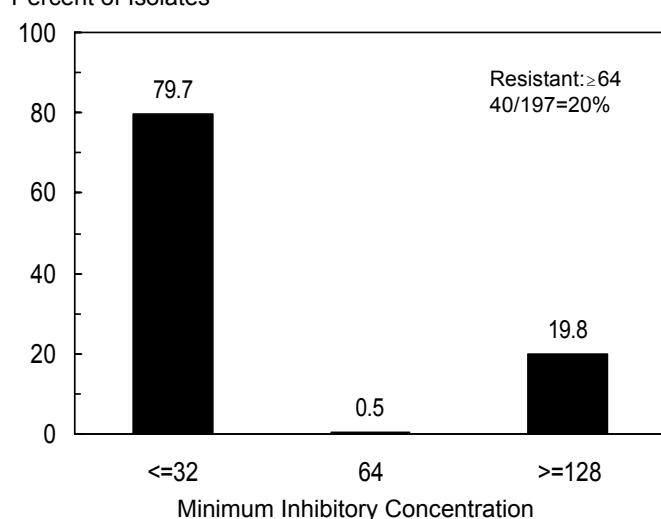
1999 (N=166)



2000 (N=177)



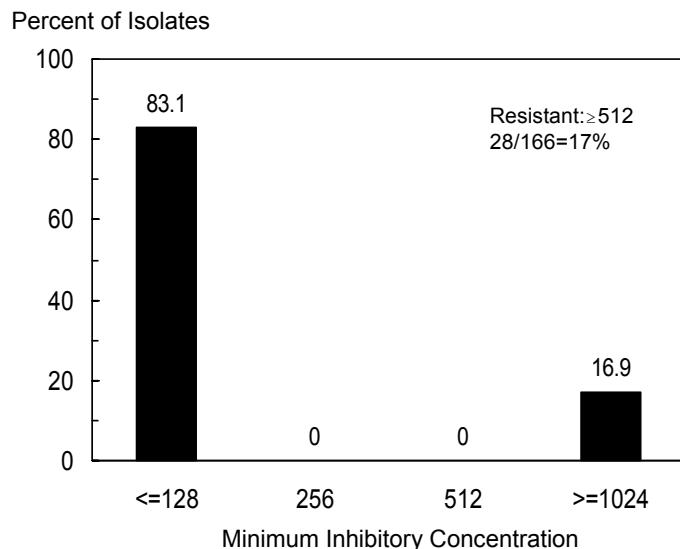
2001 (N=197)



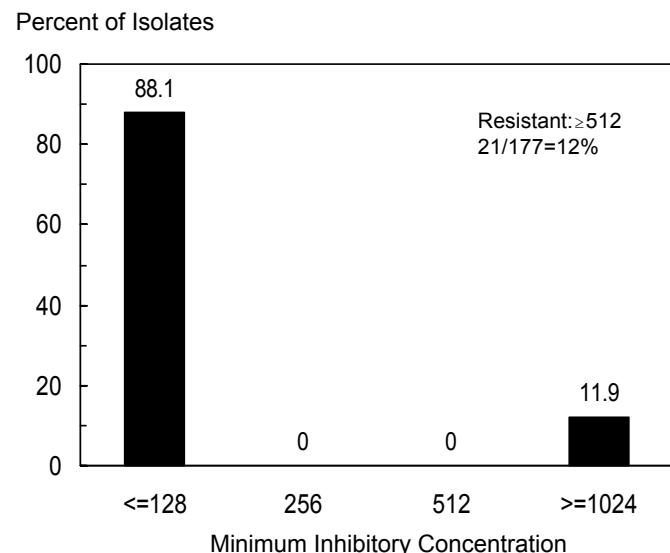
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Figure 12o. MICs for sulfamethoxazole among *Salmonella* Typhi isolates, 1999-2001

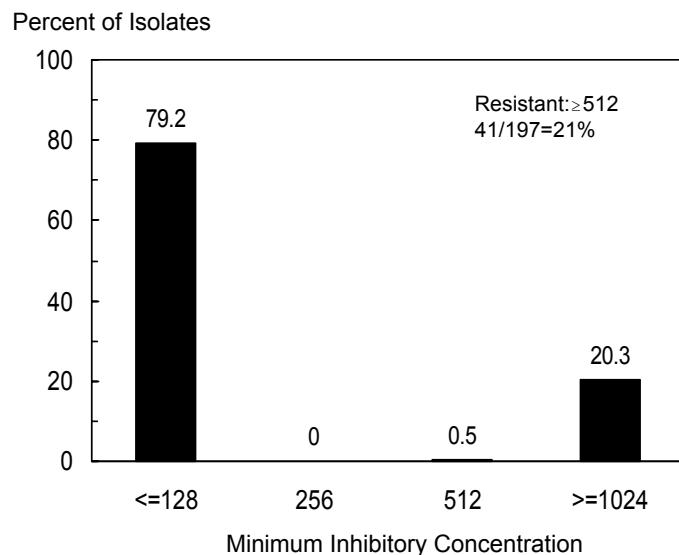
1999 (N=166)



2000 (N=177)



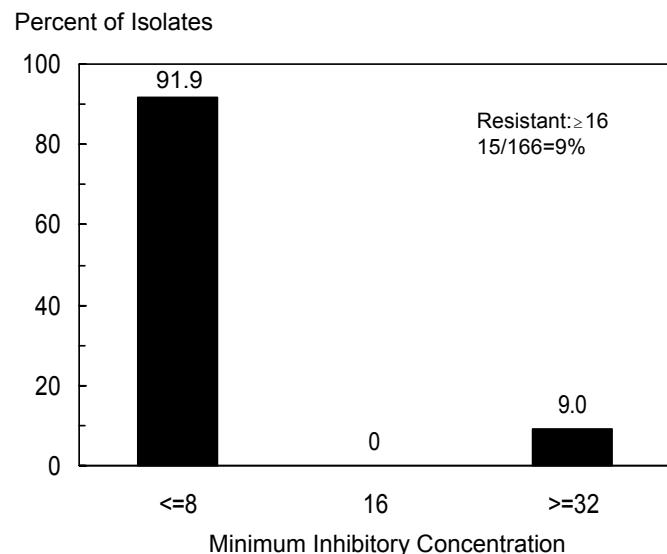
2001 (N=197)



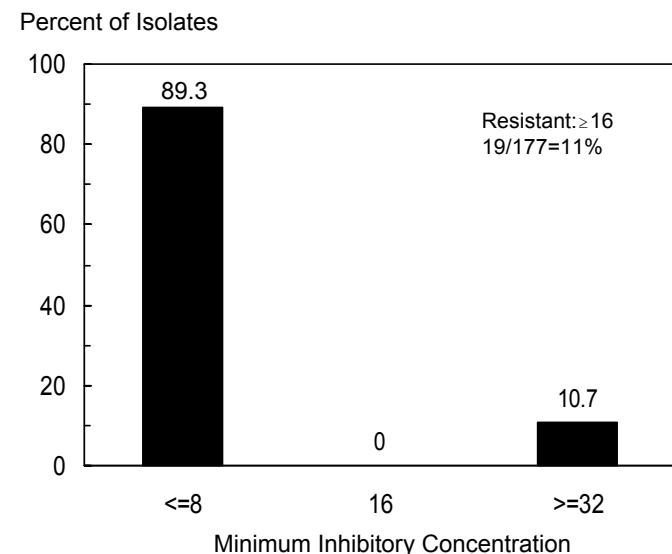
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Figure 12p. MICs for tetracycline among *Salmonella* Typhi isolates, 1999-2001

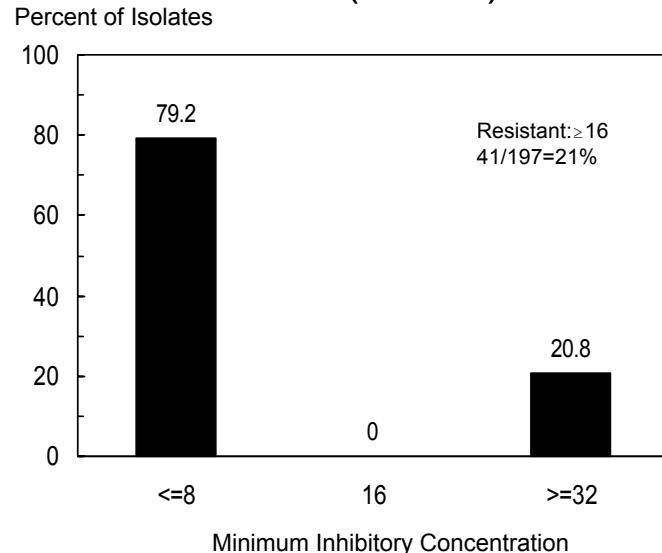
1999 (N=166)



2000 (N=177)



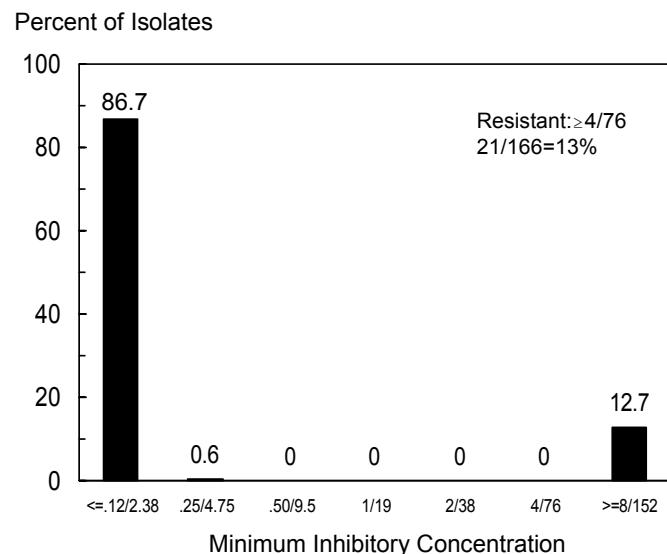
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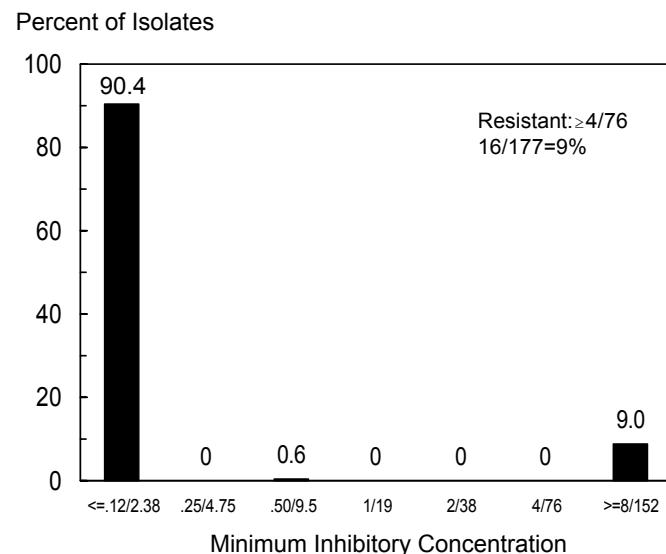
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Figure 12q. MICs for trimethoprim-sulfamethoxazole among *Salmonella* Typhi isolates, 1999 - 2001

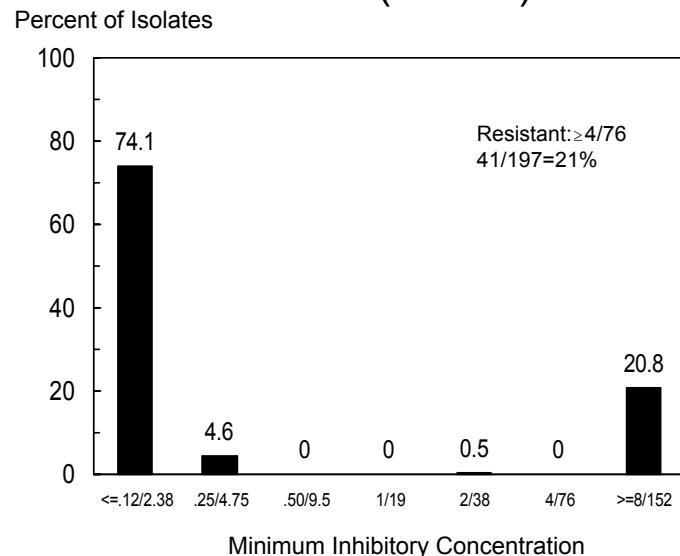
1999 (N=166)



2000 (N=177)

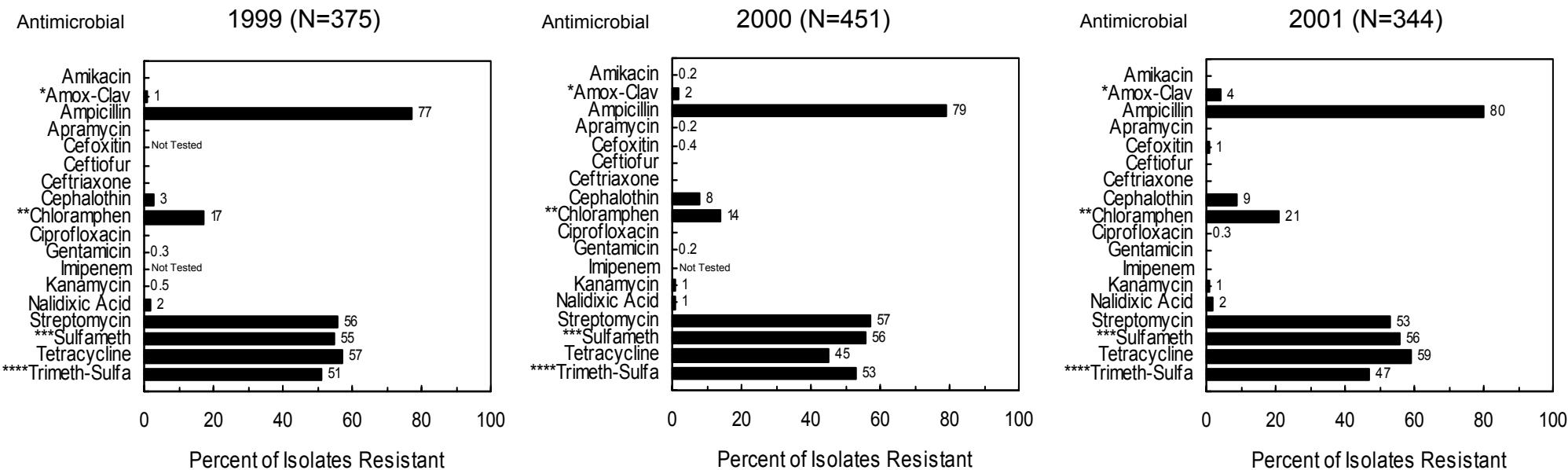


2001 (N=197)



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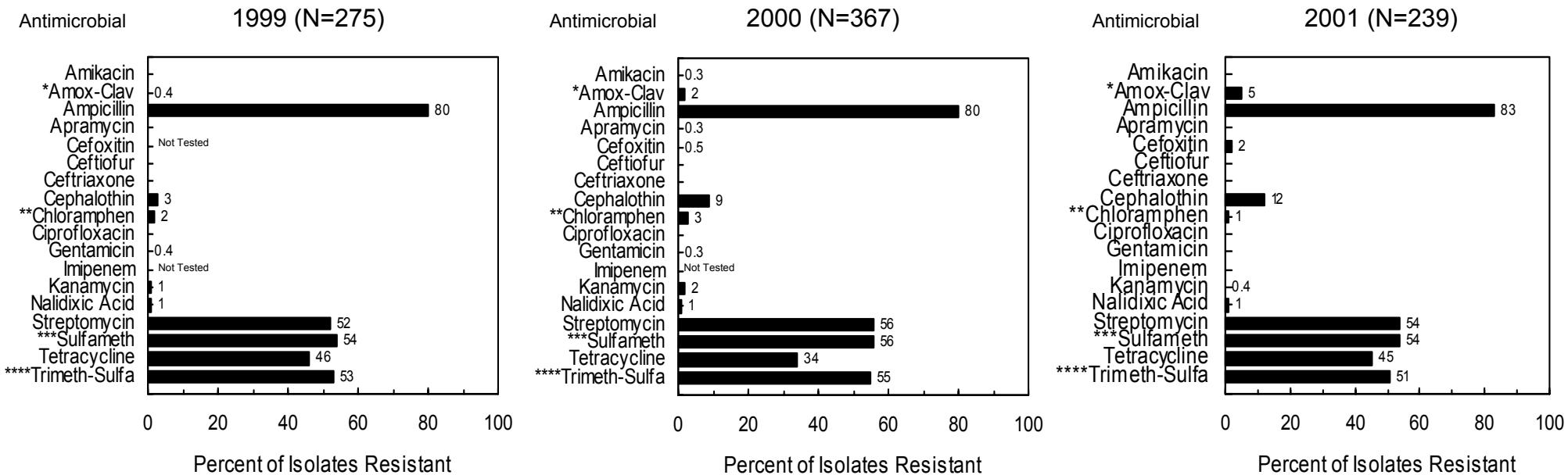
Figure 13. Resistance among *Shigella* isolates, 1999-2001



*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

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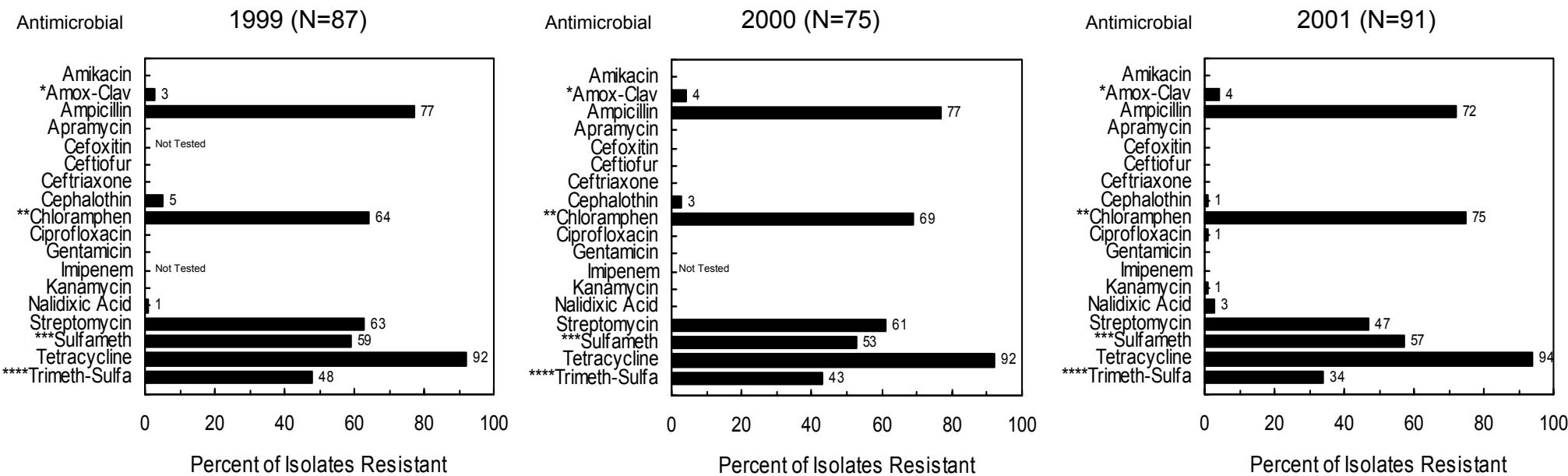
Figure 14a. Resistance among *Shigella sonnei* isolates, 1999-2001



*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

National Antimicrobial Resistance Monitoring System For Enteric Bacteria

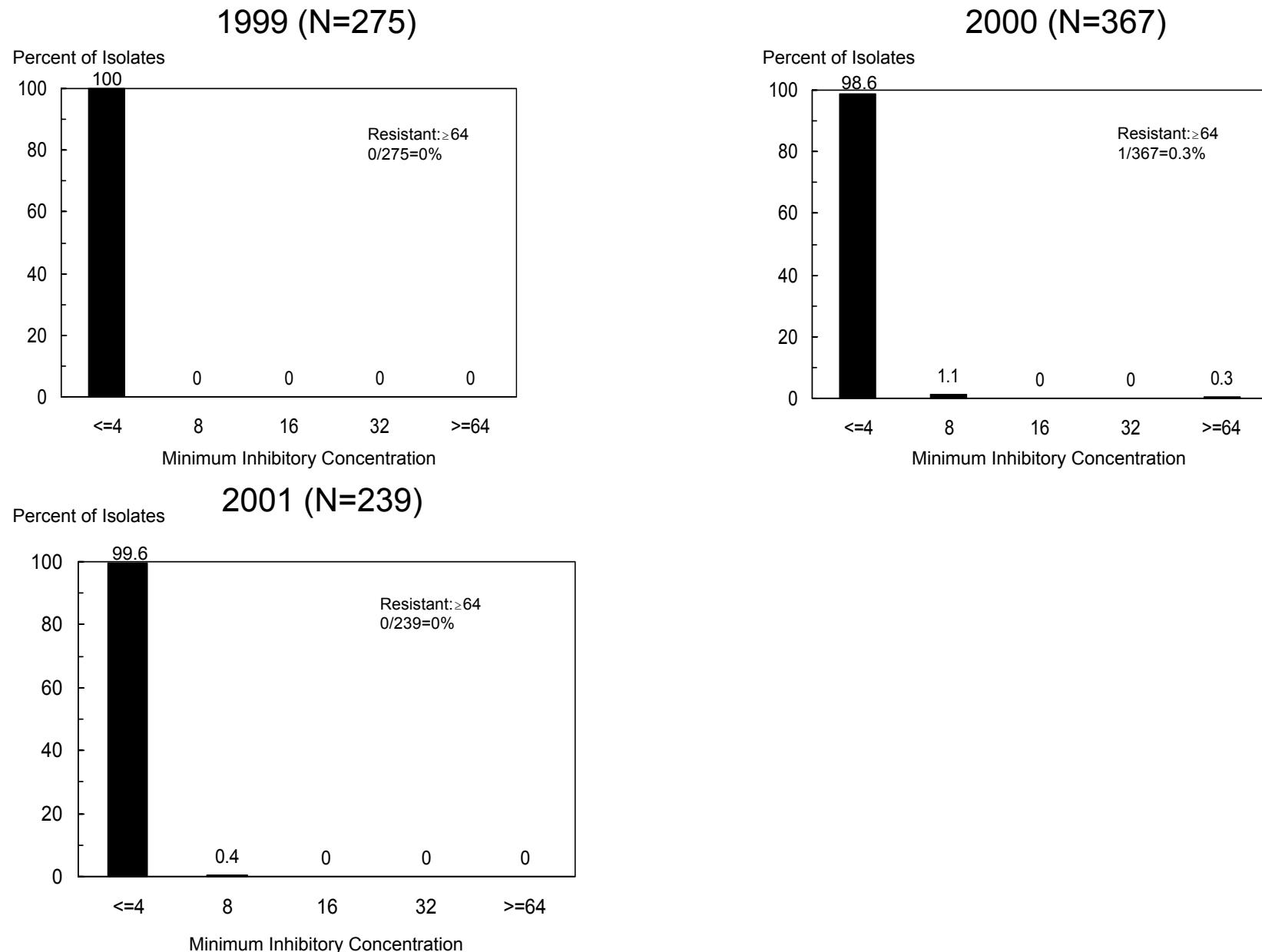
Figure 14b. Resistance among *Shigella flexneri* isolates, 1999-2001



*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

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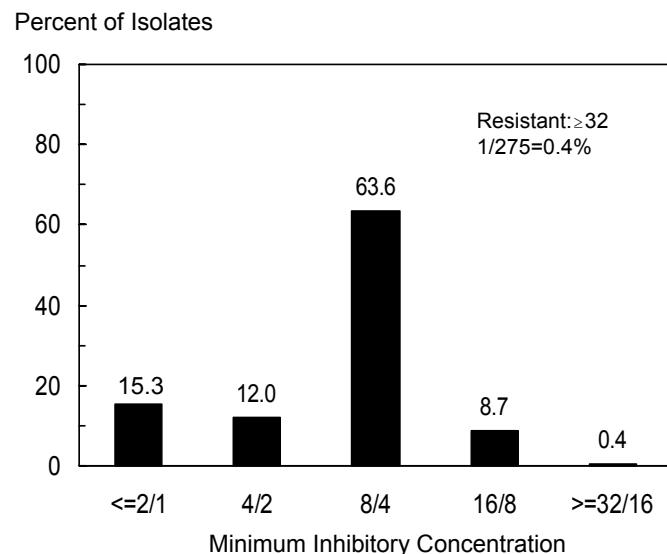
Figure 15a. MICs for amikacin among *Shigella sonnei* isolates, 1999-2001



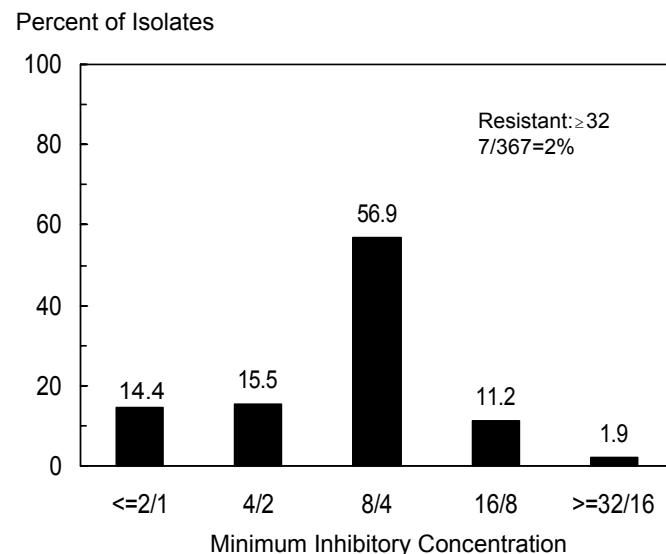
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Figure 15b. MICs for amoxicillin-clavulanic acid among *Shigella sonnei* isolates, 1999-2001

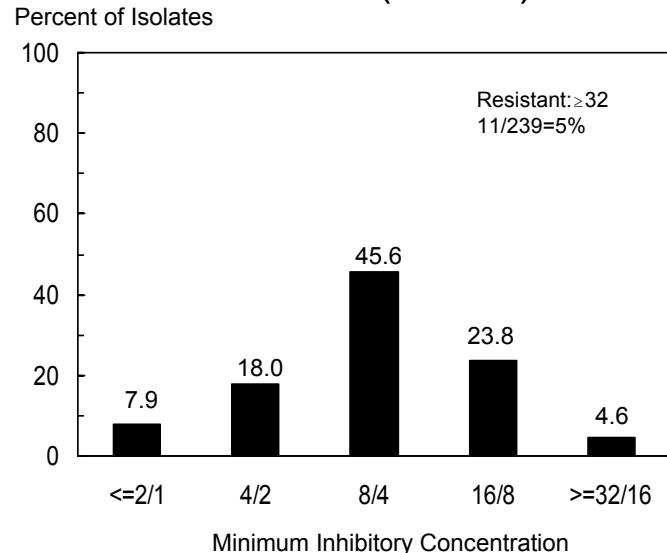
1999 (N=275)



2000 (N=367)



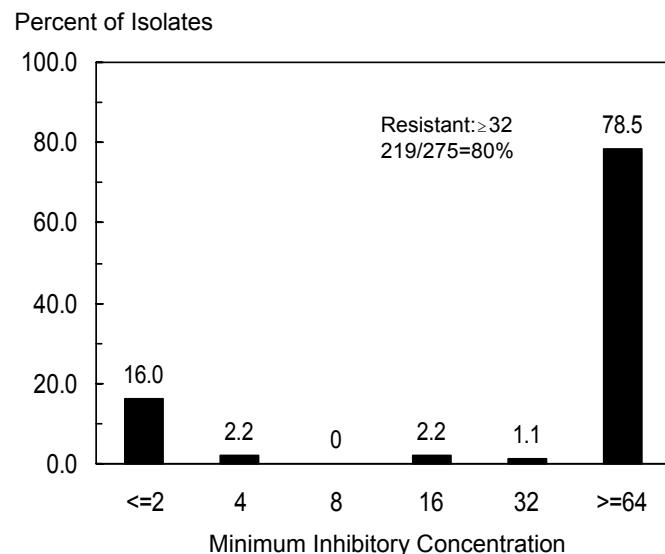
2001 (N=239)



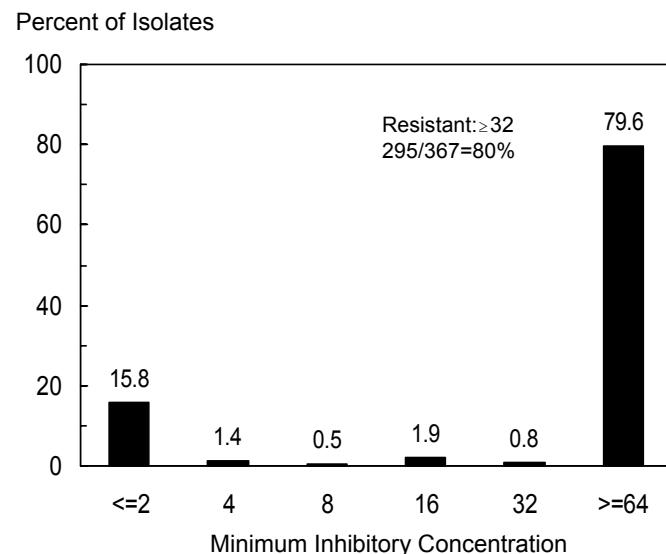
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Figure 15c. MICs for ampicillin among *Shigella sonnei* isolates, 1999-2001

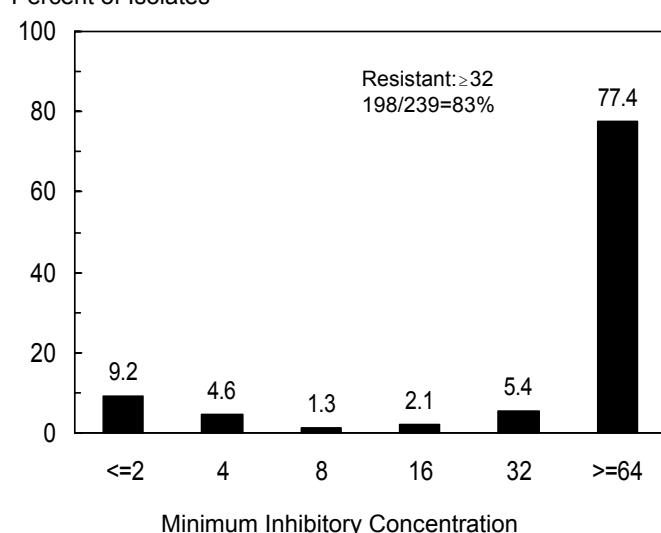
1999 (N=275)



2000 (N=367)



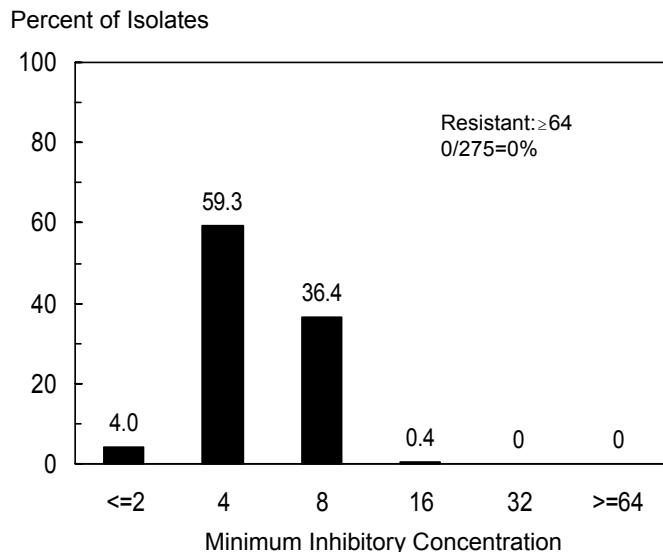
2001 (N=239)



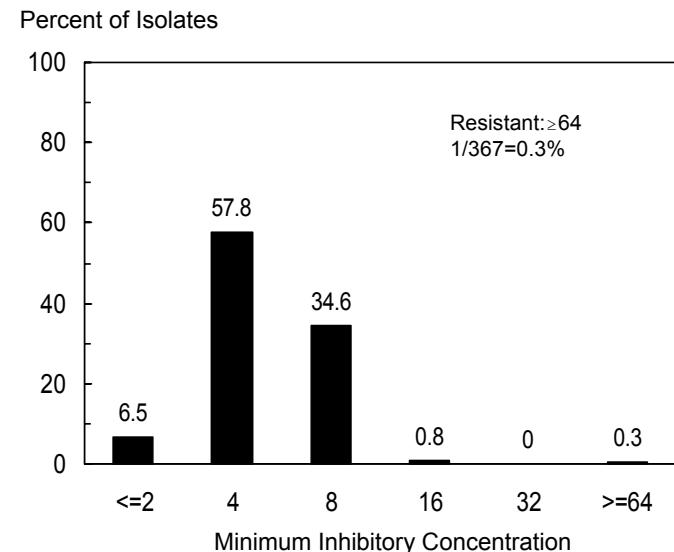
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Figure 15d. MICs for apramycin among *Shigella sonnei* isolates, 1999-2001

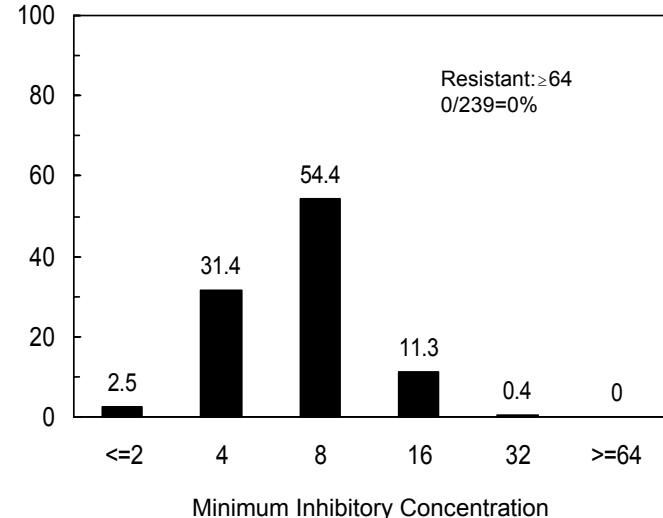
1999 (N=275)



2000 (N=367)



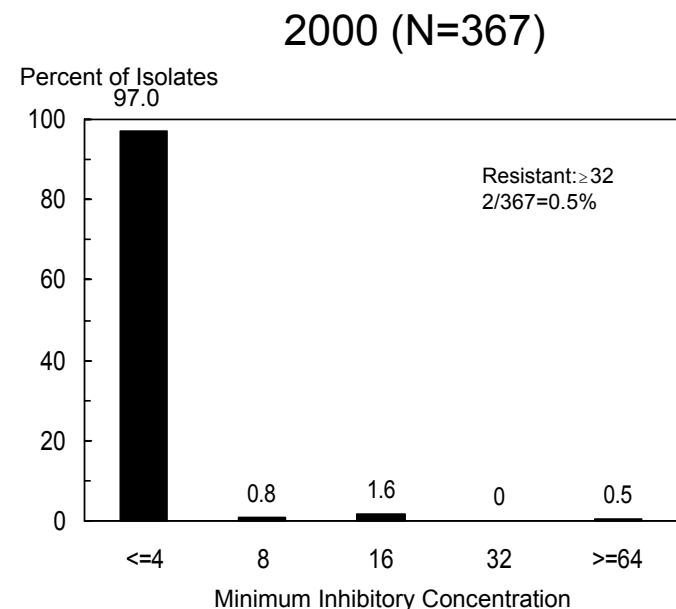
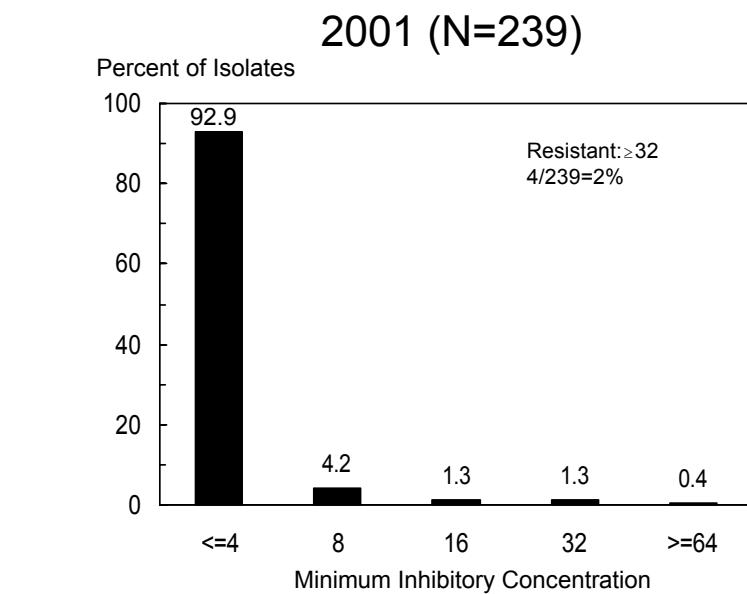
2001 (N=239)



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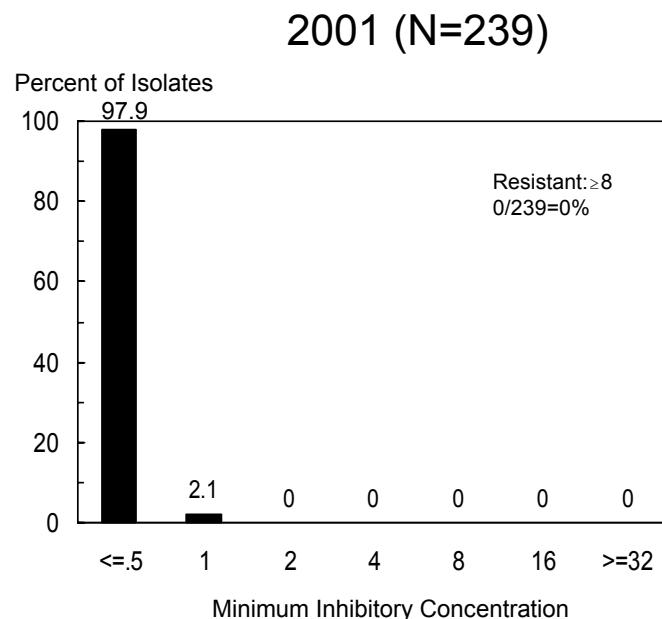
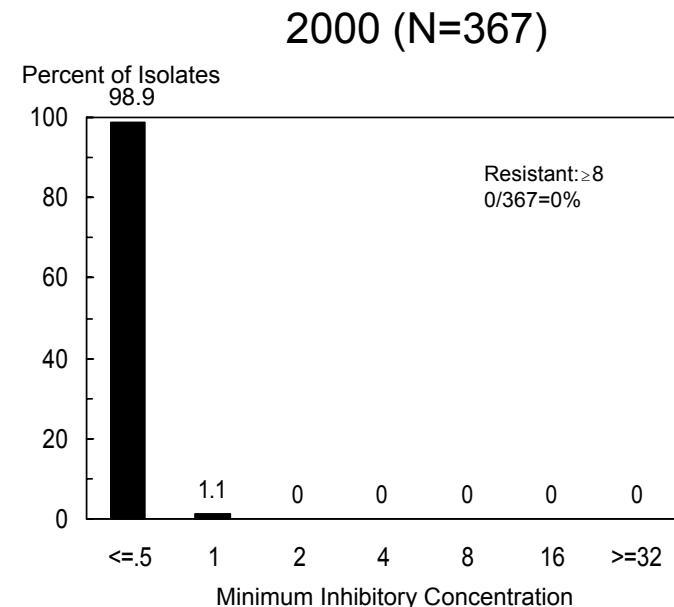
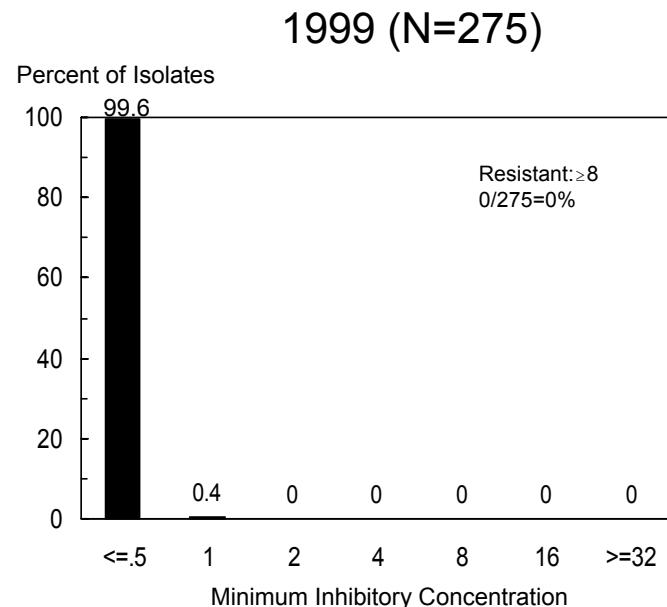
Figure 15e. MICs for cefoxitin among *Shigella sonnei* isolates, 1999-2001

Not Tested in 1999



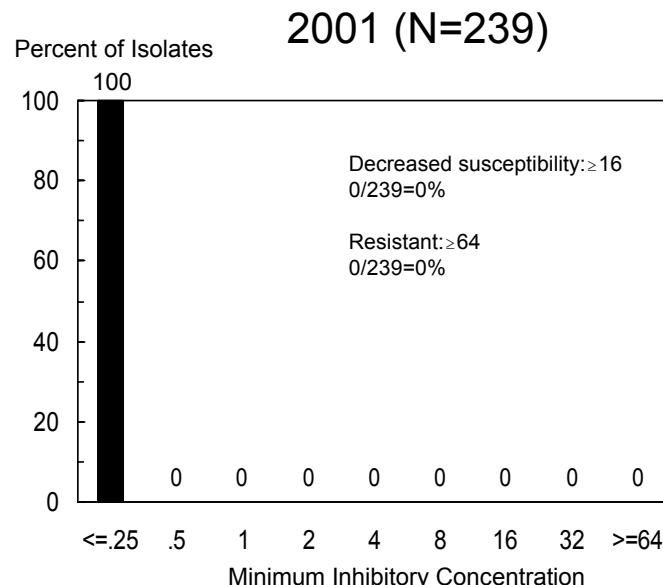
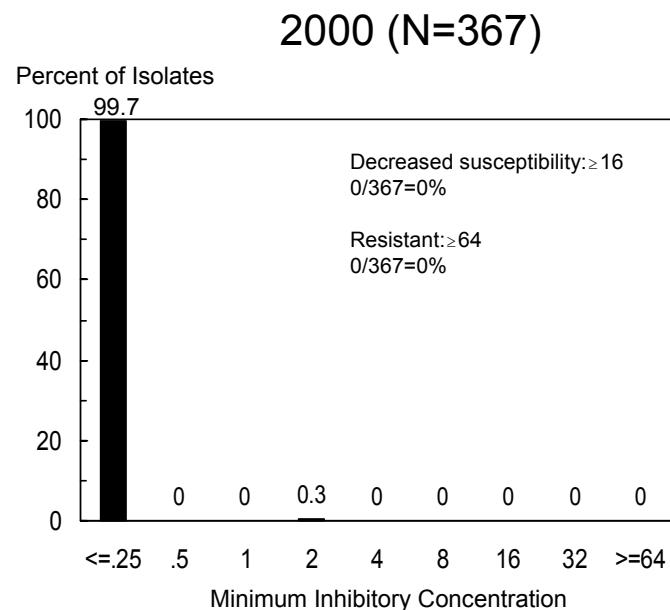
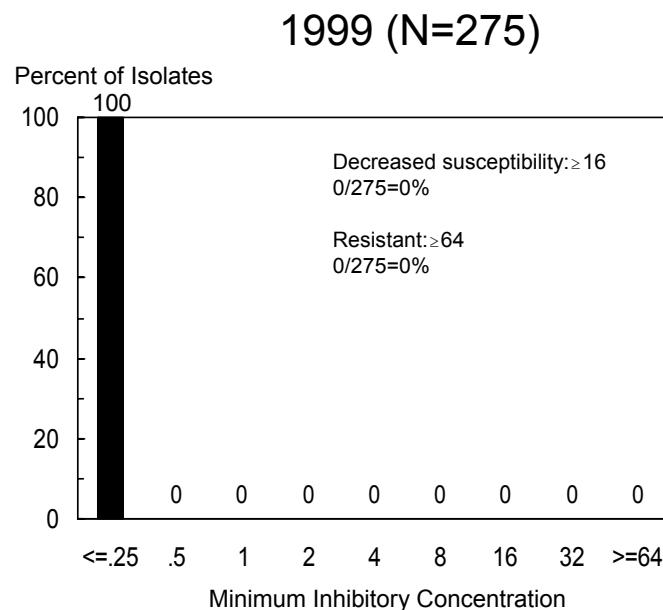
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Figure 15f. MICs for ceftiofur among *Shigella sonnei* isolates, 1999-2001



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Figure 15g. MICs for ceftriaxone* among *Shigella sonnei* isolates, 1999-2001

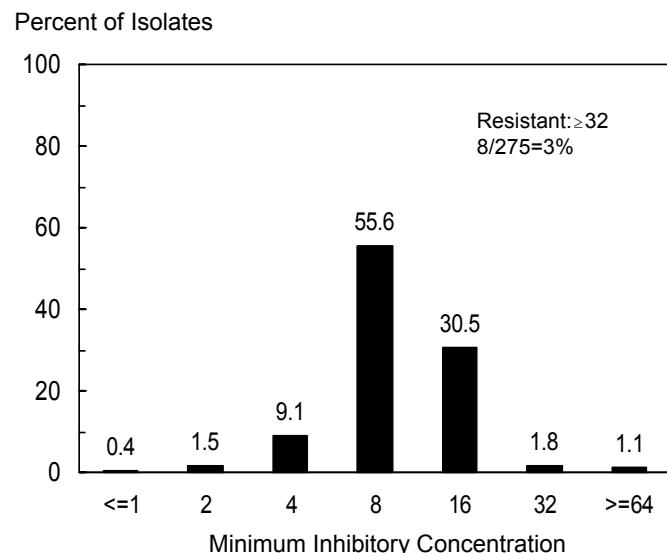


*Sensititre® results only

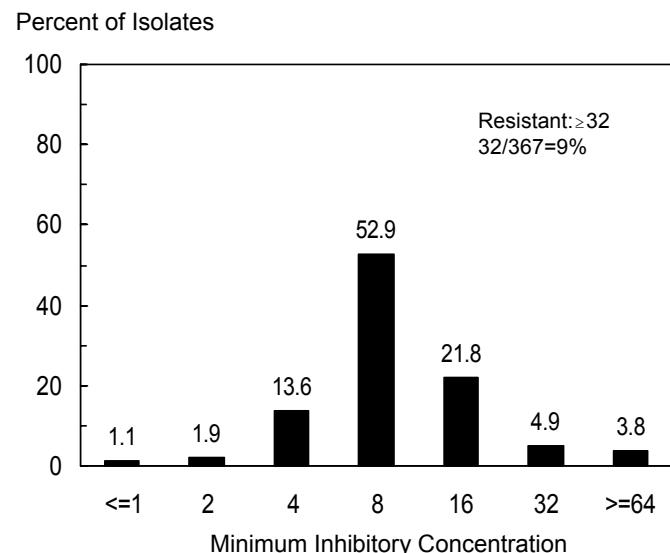
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Figure 15h. MICs for cephalothin among *Shigella sonnei* isolates, 1999-2001

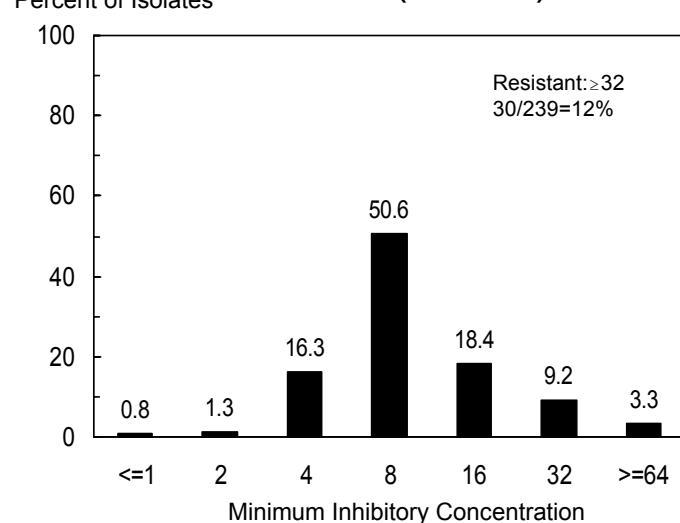
1999 (N=275)



2000 (N=367)

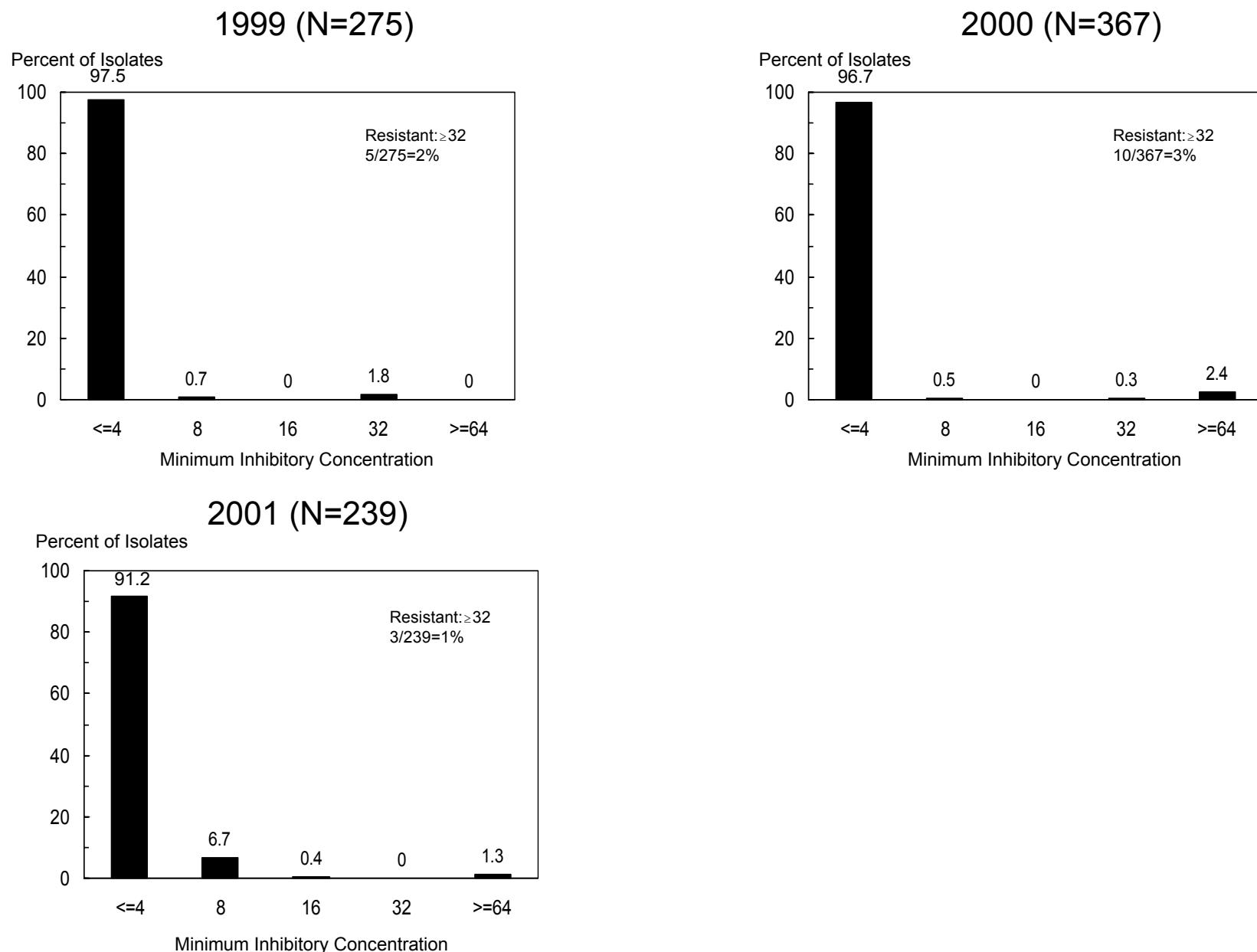


2001 (N=239)



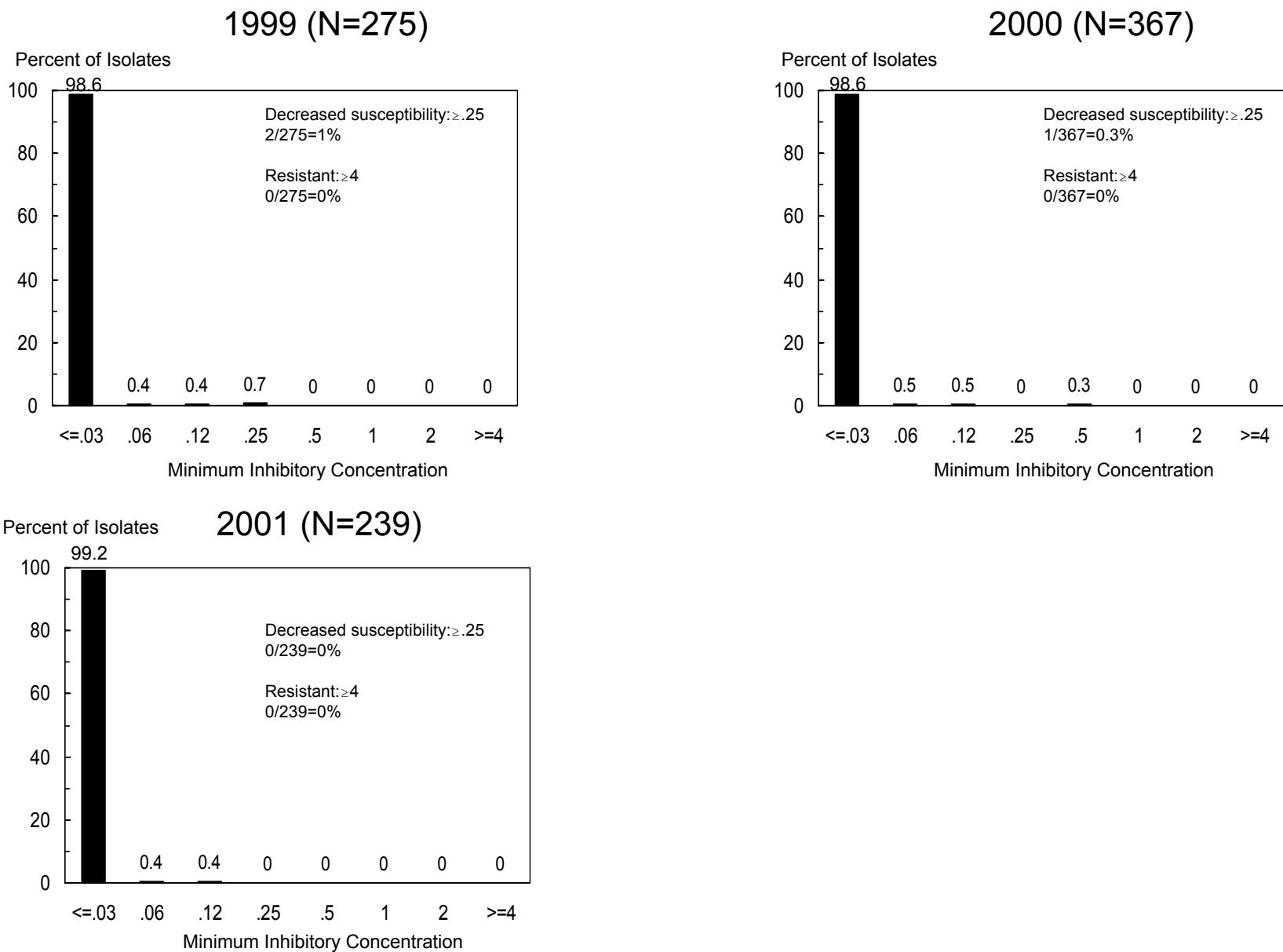
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Figure 15i. MICs for chloramphenicol among *Shigella sonnei* isolates, 1999-2001



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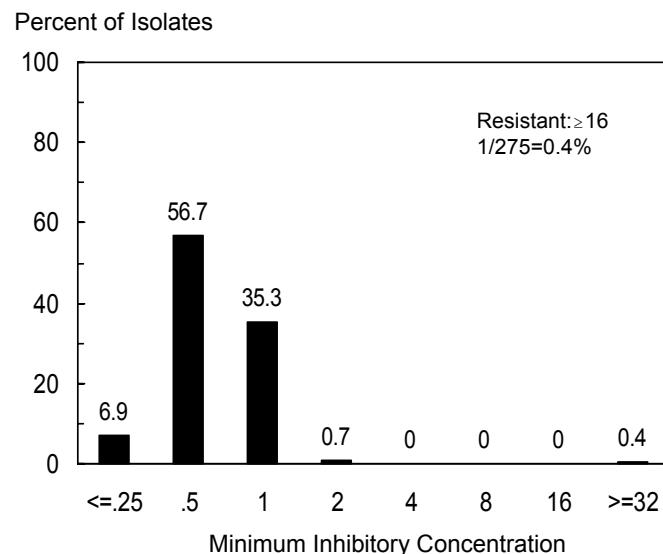
Figure 15j. MICs for ciprofloxacin among *Shigella sonnei* isolates, 1999-2001



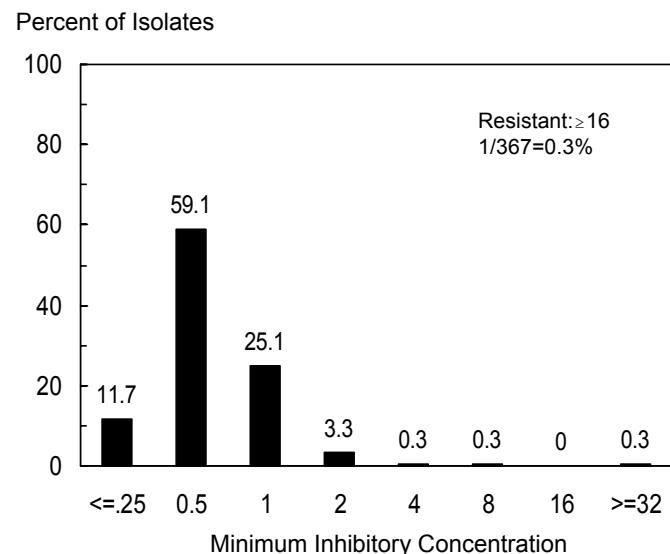
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Figure 15k. MICs for gentamicin among *Shigella sonnei* isolates, 1999-2001

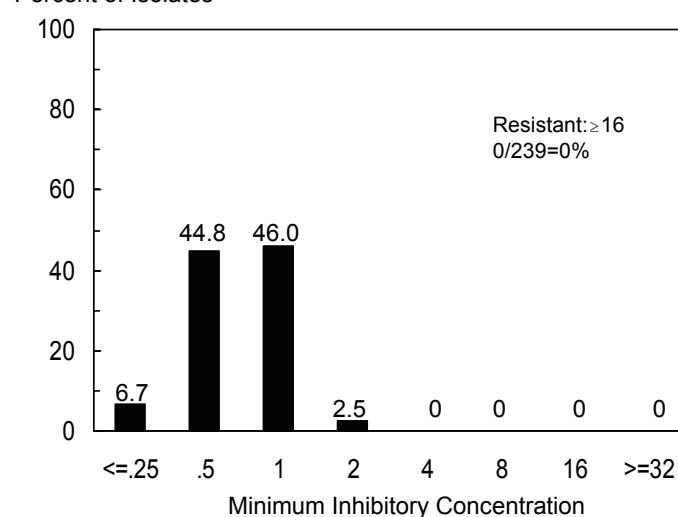
1999 (N=275)



2000 (N=367)

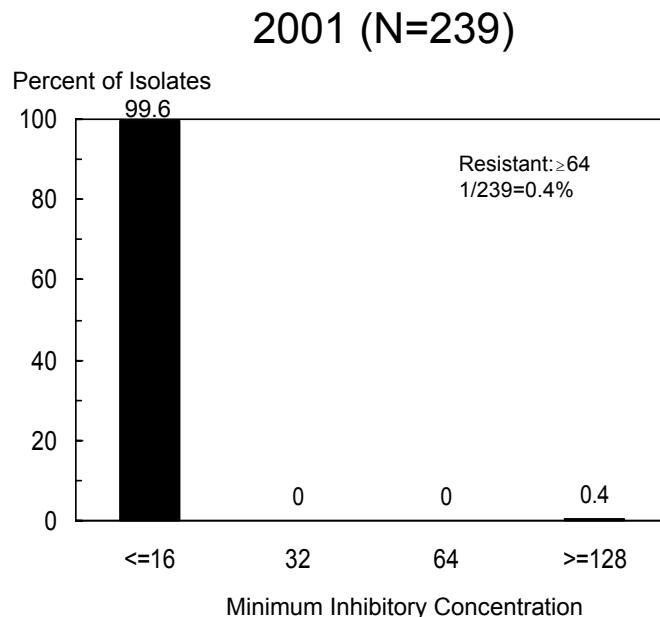
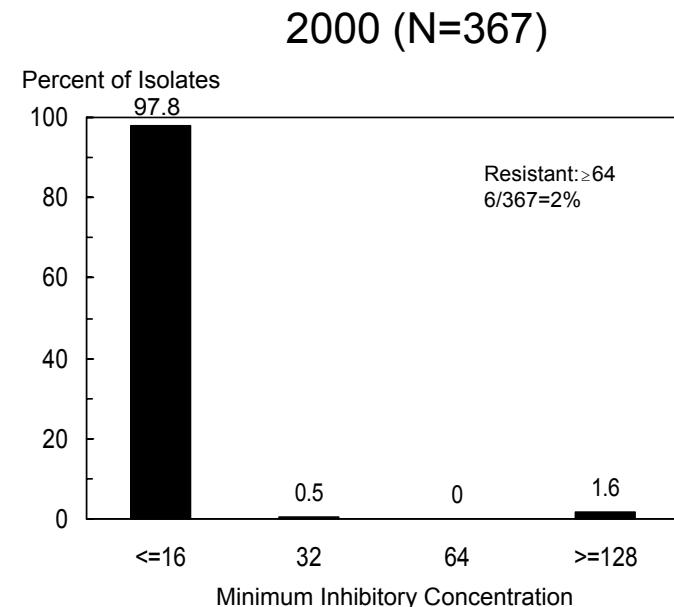
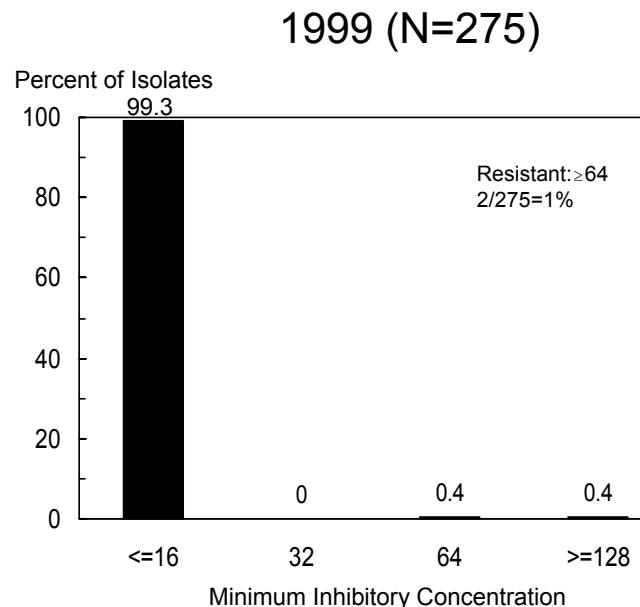


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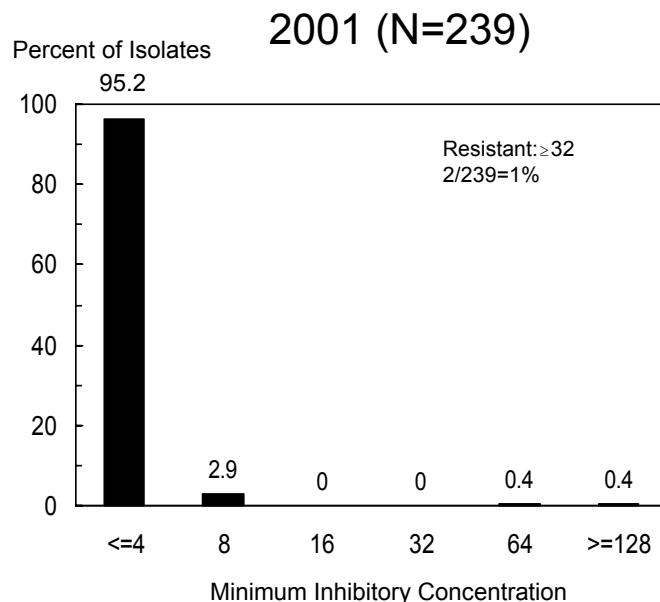
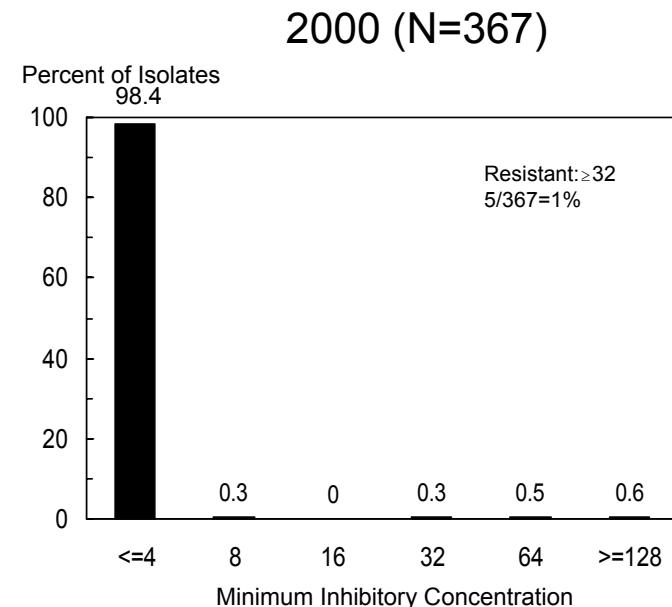
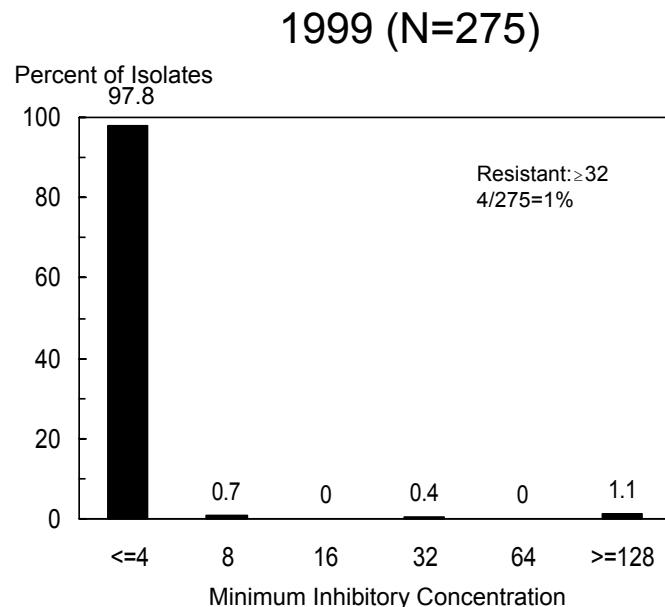
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Figure 15I. MICs for kanamycin among *Shigella sonnei* isolates, 1999-2001



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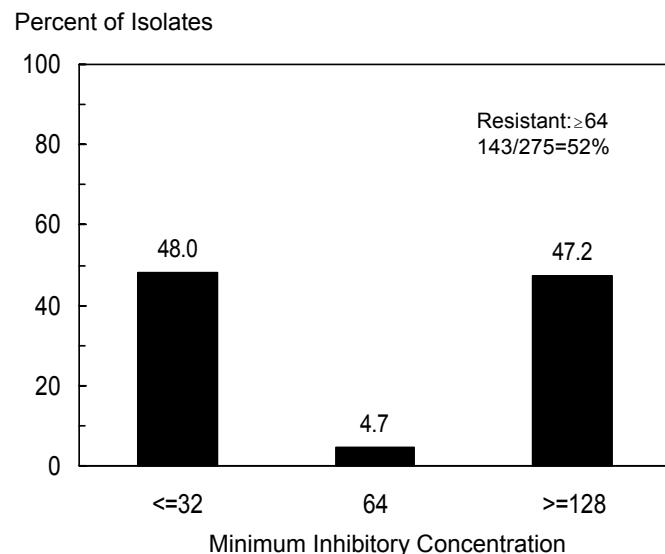
Figure 15m. MICs for nalidixic acid among *Shigella sonnei* isolates, 1999-2001



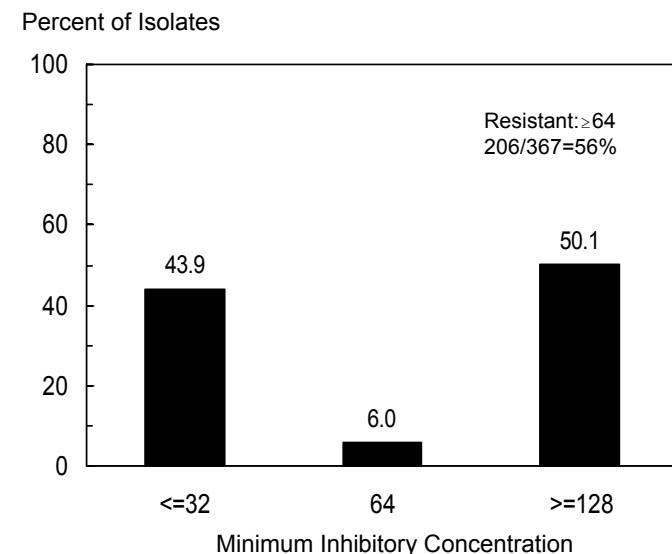
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Figure 15n. MICs for streptomycin among *Shigella sonnei* isolates, 1999-2001

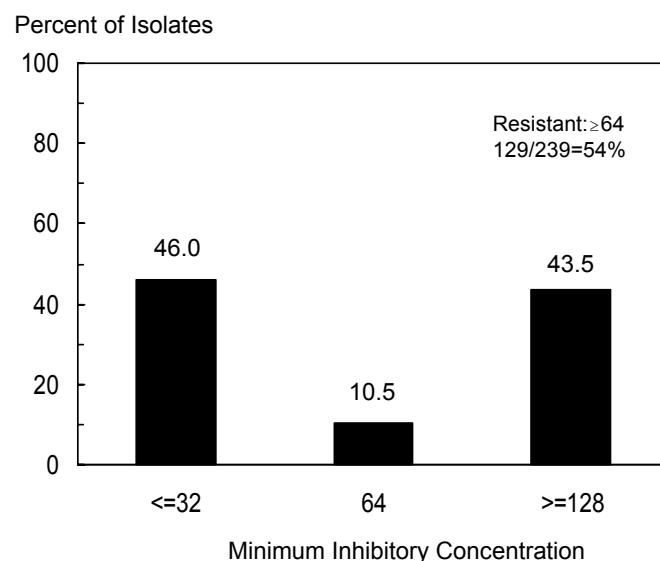
1999 (N=275)



2000 (N=367)



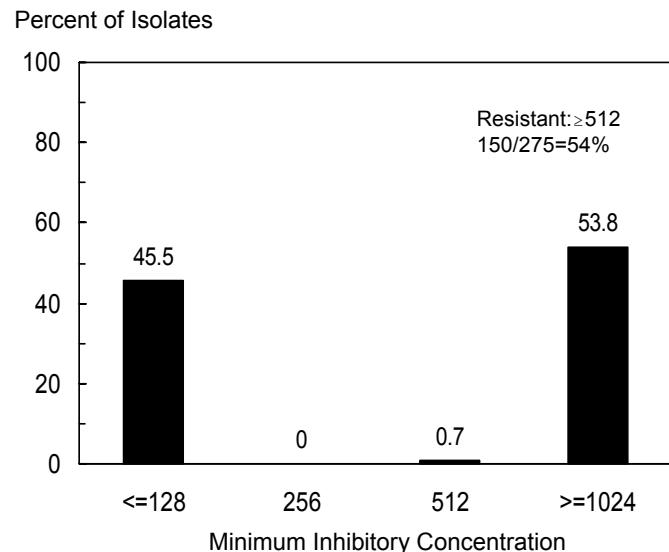
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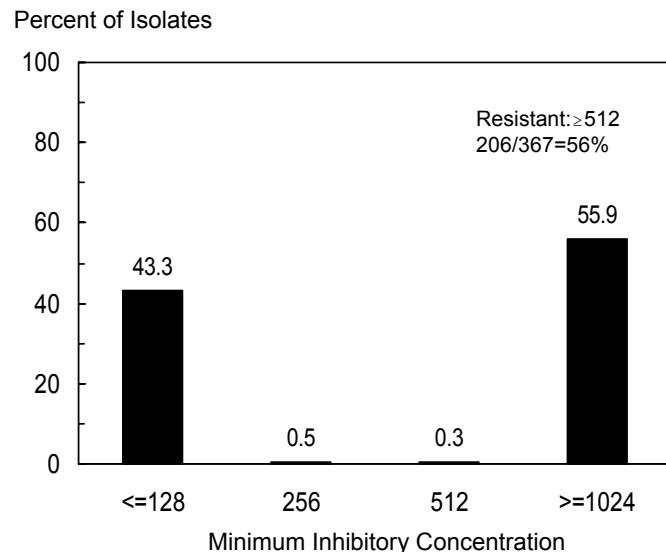
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Figure 15o. MICs for sulfamethoxazole among *Shigella sonnei* isolates, 1999-2001

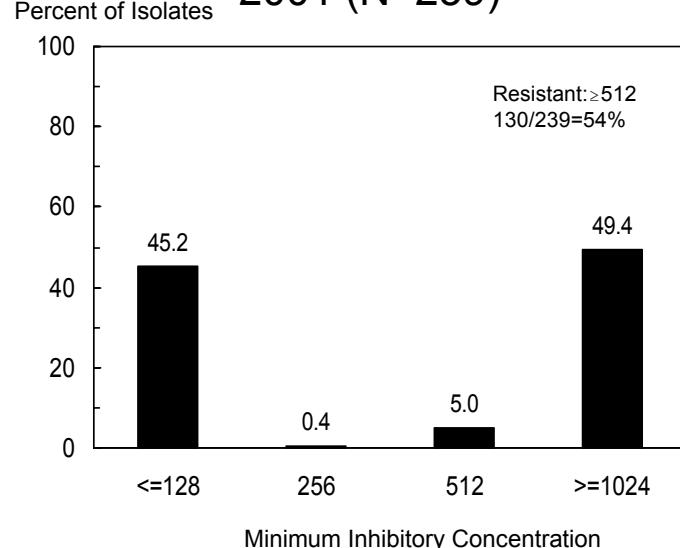
1999 (N=275)



2000 (N=367)



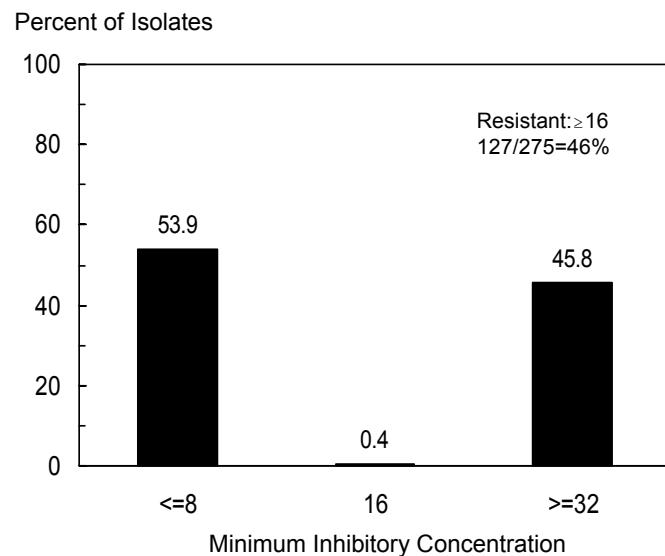
2001 (N=239)



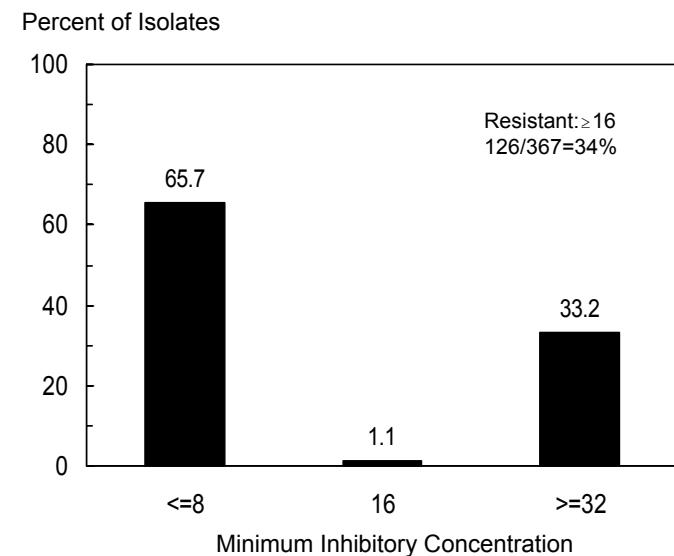
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Figure 15p. MICs for tetracycline among *Shigella sonnei* isolates, 1999-2001

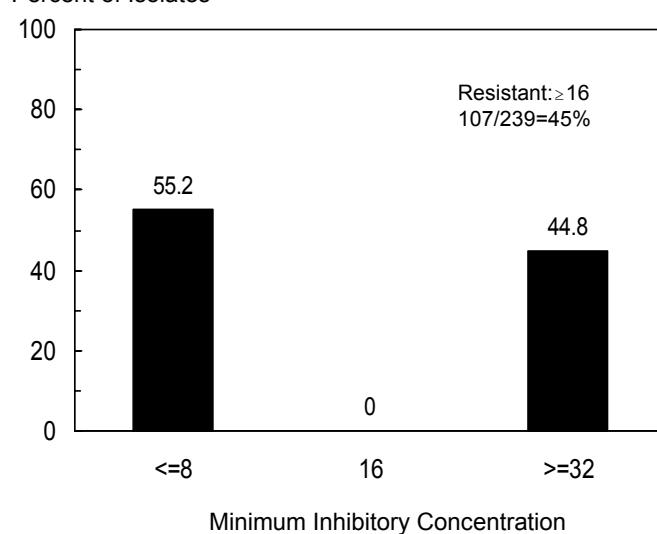
1999 (N=275)



2000 (N=367)

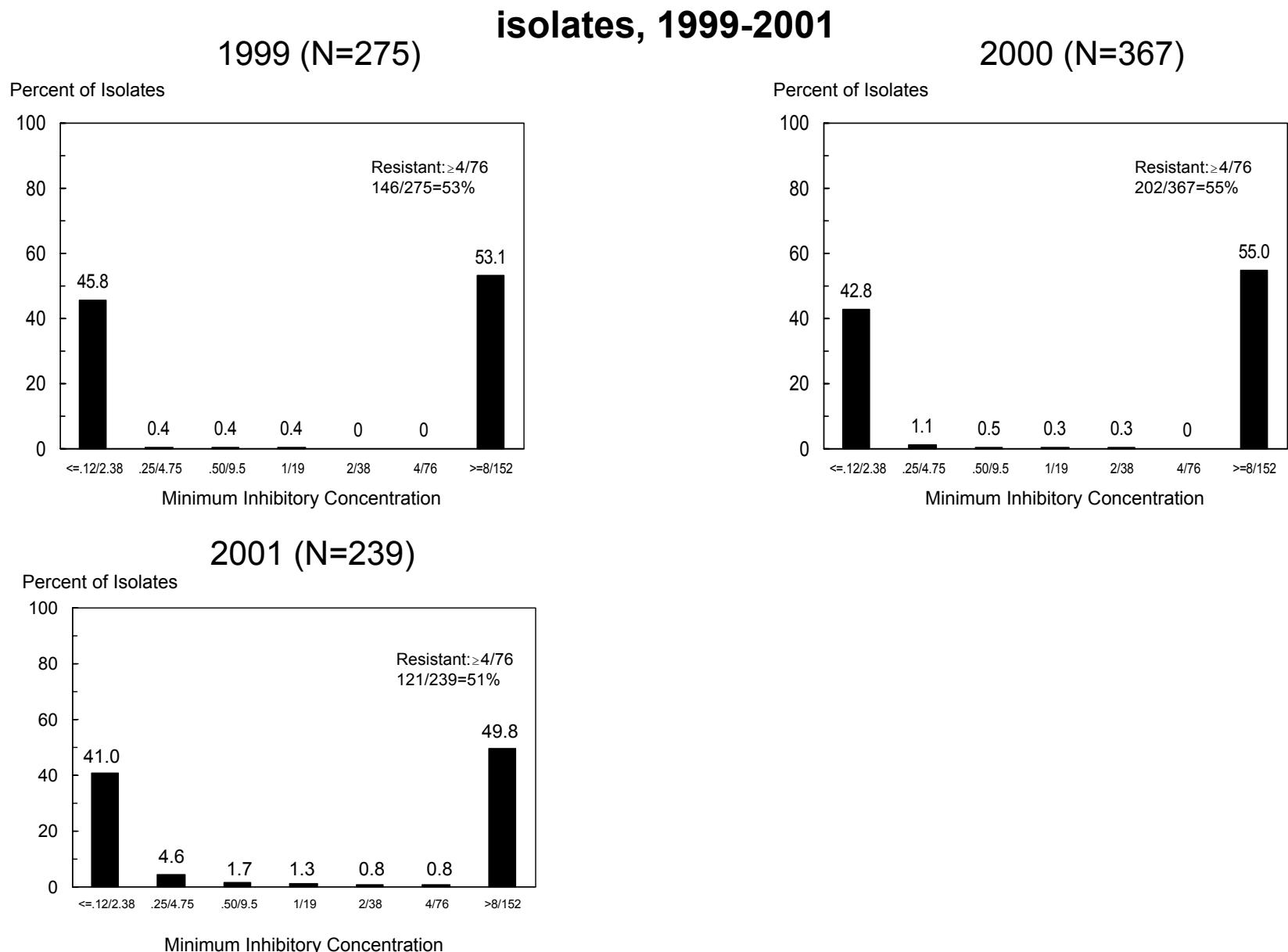


2001 (N=239)



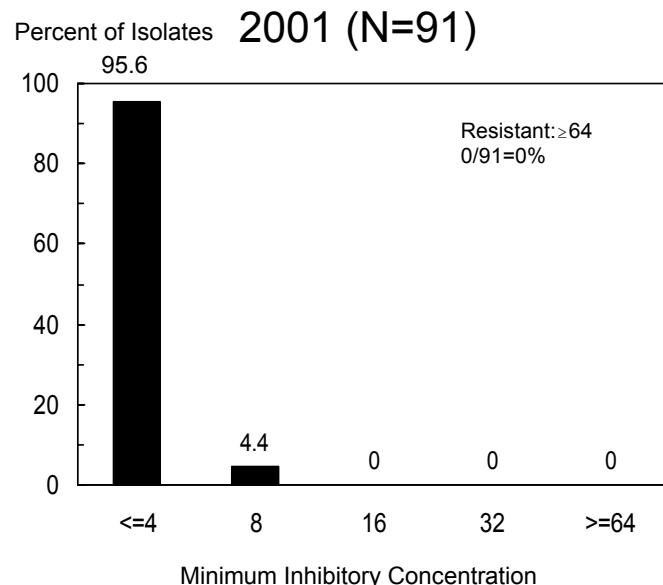
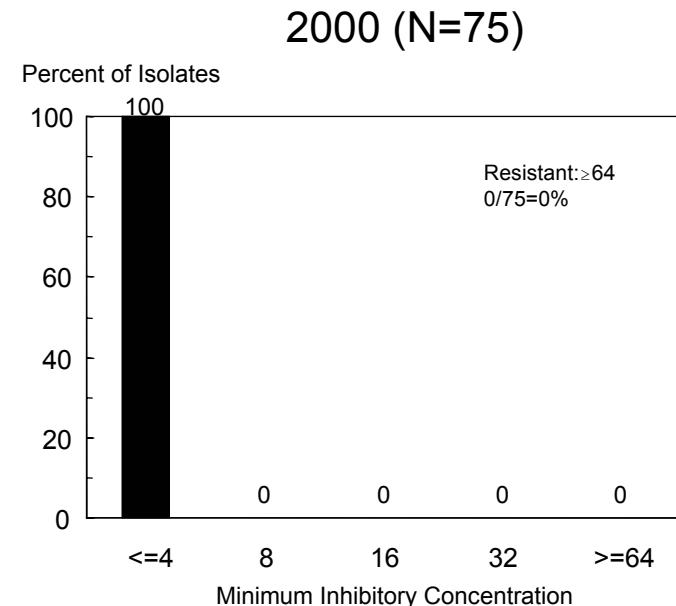
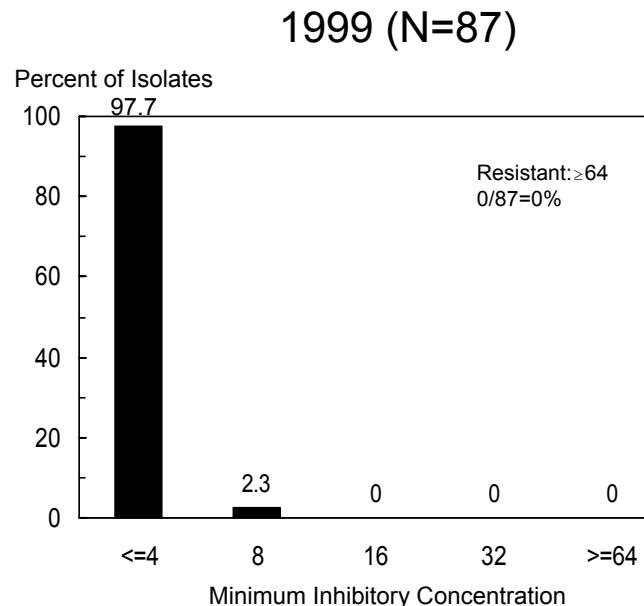
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Figure 15q. MICs for trimethoprim-sulfamethoxazole among *Shigella sonnei*



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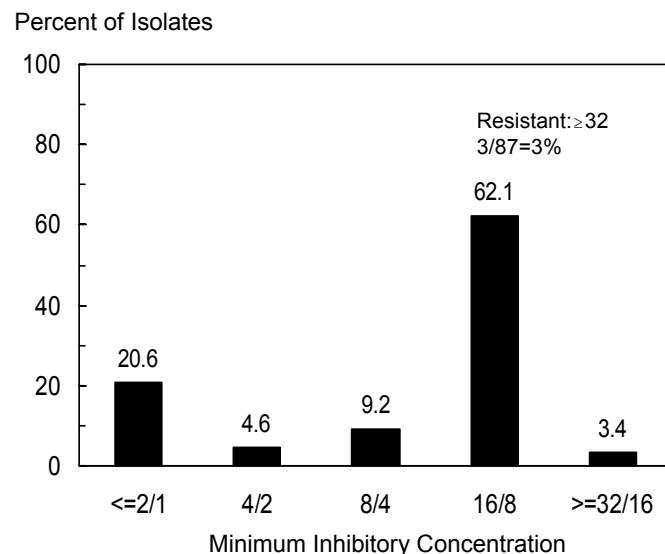
Figure 16a. MICs for amikacin among *Shigella flexneri* isolates, 1999-2001



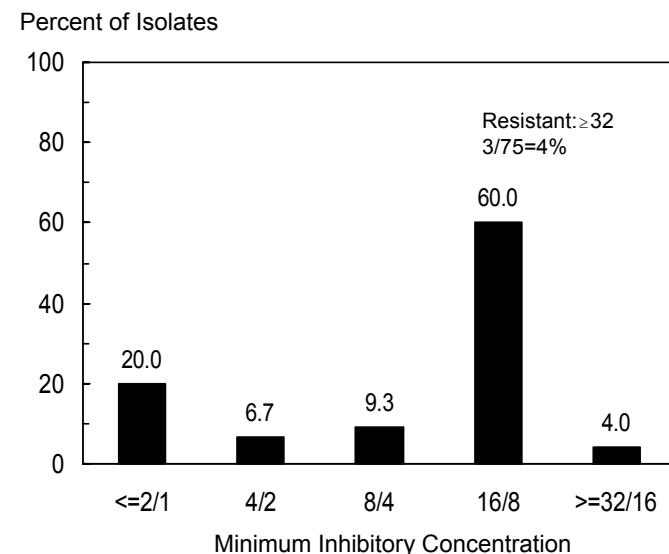
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Figure 16b. MICs for amoxicillin-clavulanic acid among *Shigella flexneri* isolates, 1999-2001

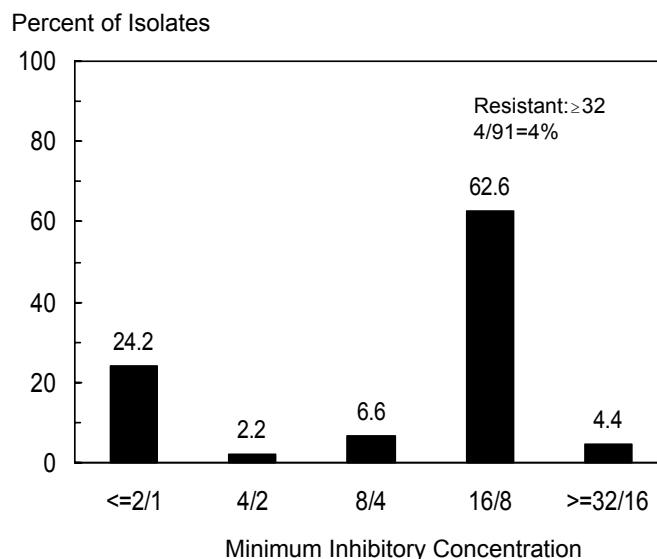
1999 (N=87)



2000 (N=75)



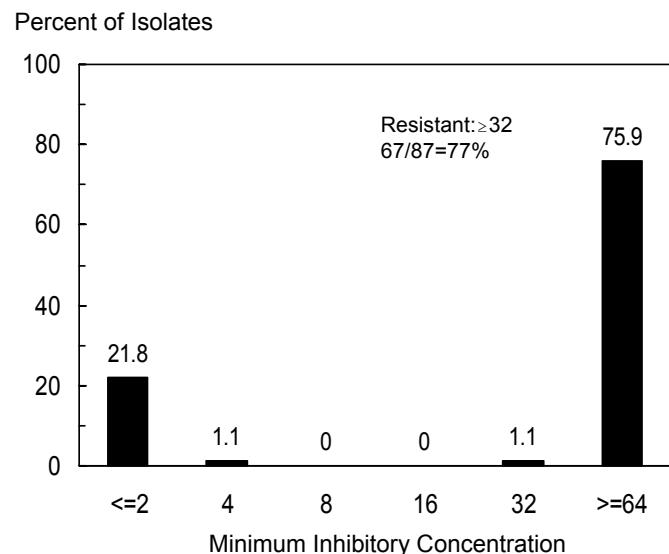
2001 (N=91)



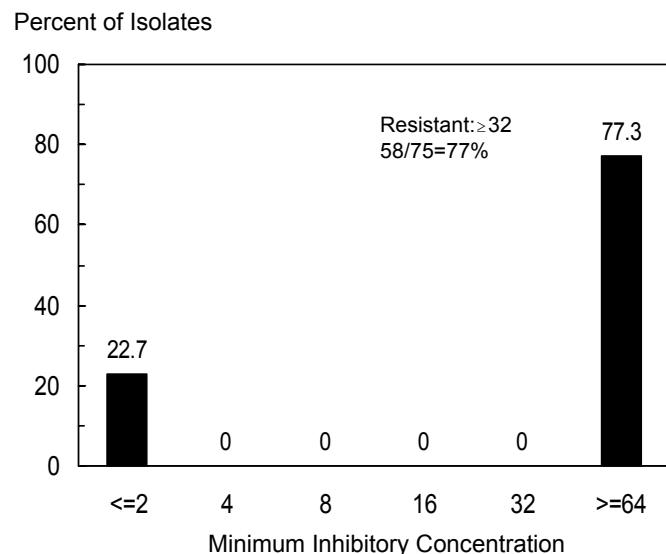
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Figure 16c. MICs for ampicillin among *Shigella flexneri* isolates, 1999-2001

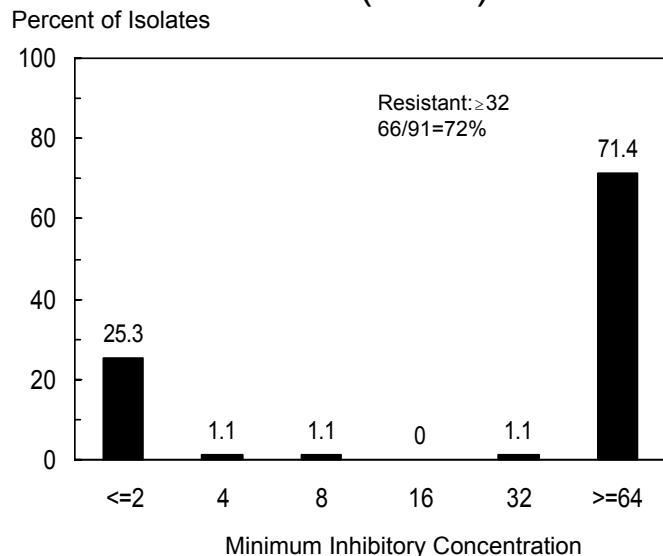
1999 (N=87)



2000 (N=75)



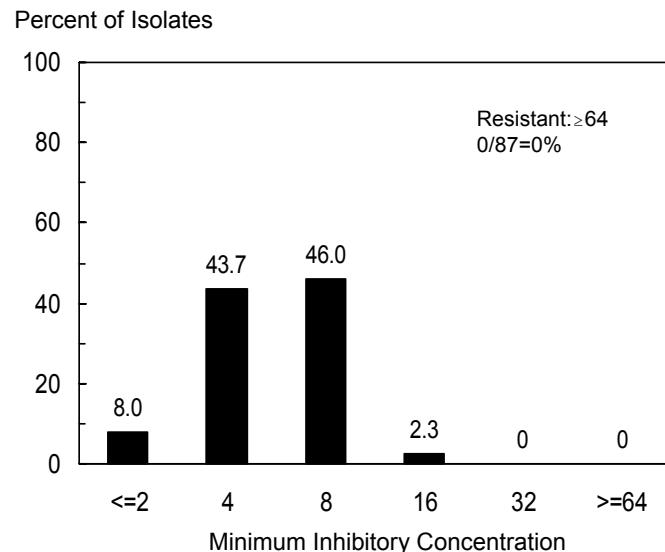
2001 (N=91)



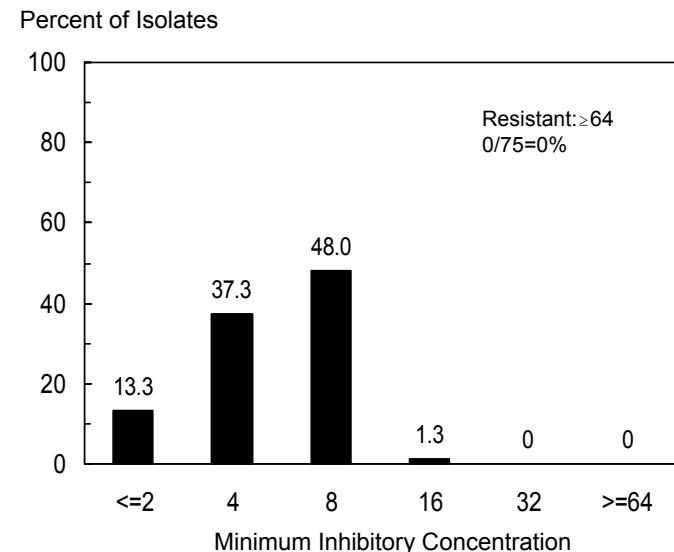
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Figure 16d. MICs for apramycin among *Shigella flexneri* isolates, 1999-2001

1999 (N=87)

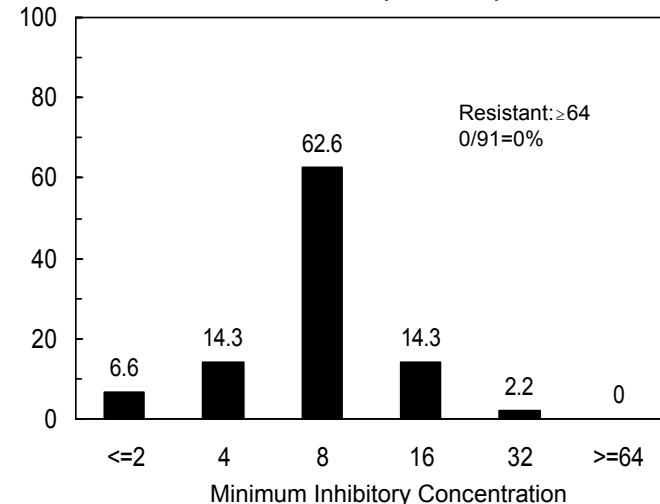


2000 (N=75)



Percent of Isolates

2001 (N=91)

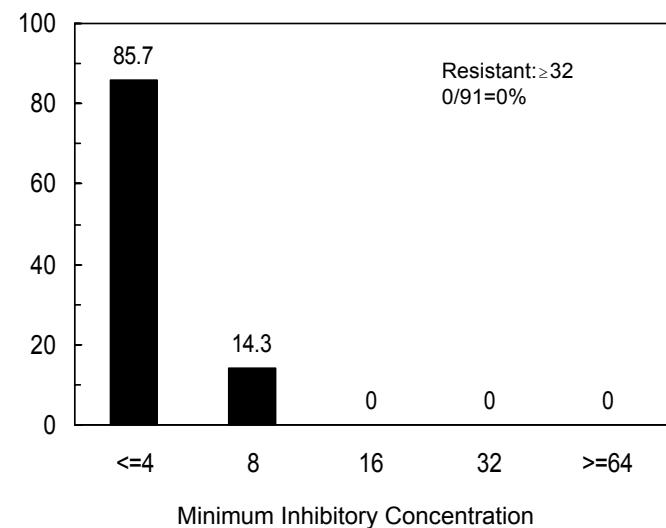


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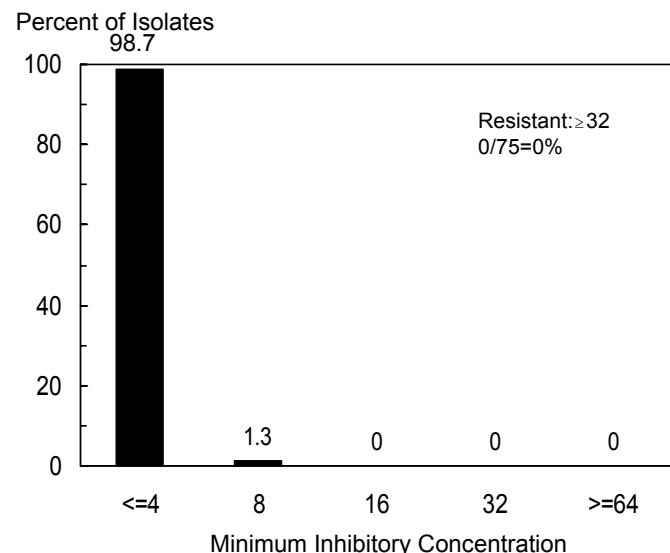
Figure 16e. MICs for cefoxitin among *Shigella flexneri* isolates, 1999-2001

Not Tested in 1999

2001 (N=91)

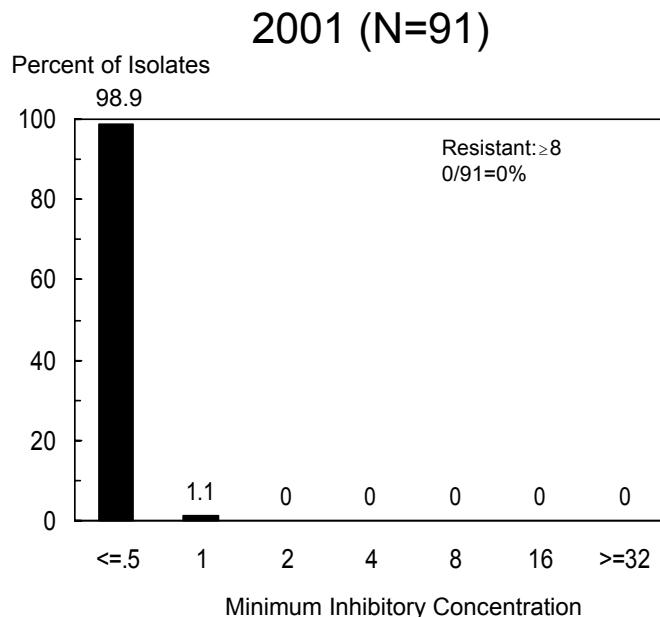
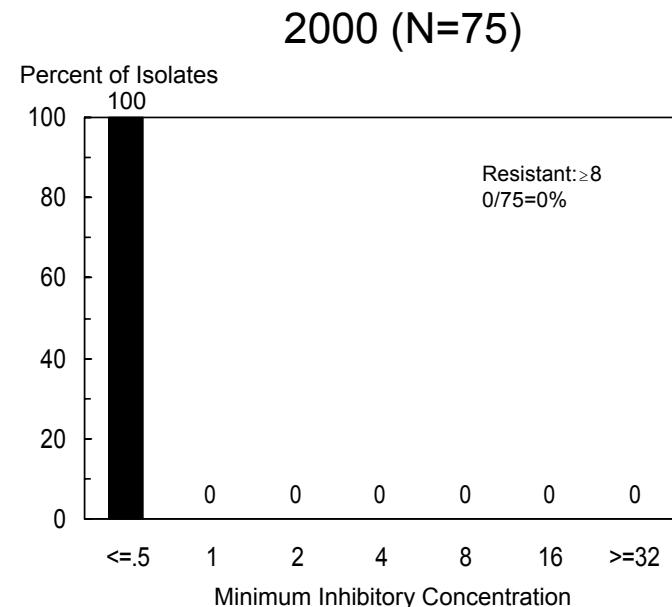
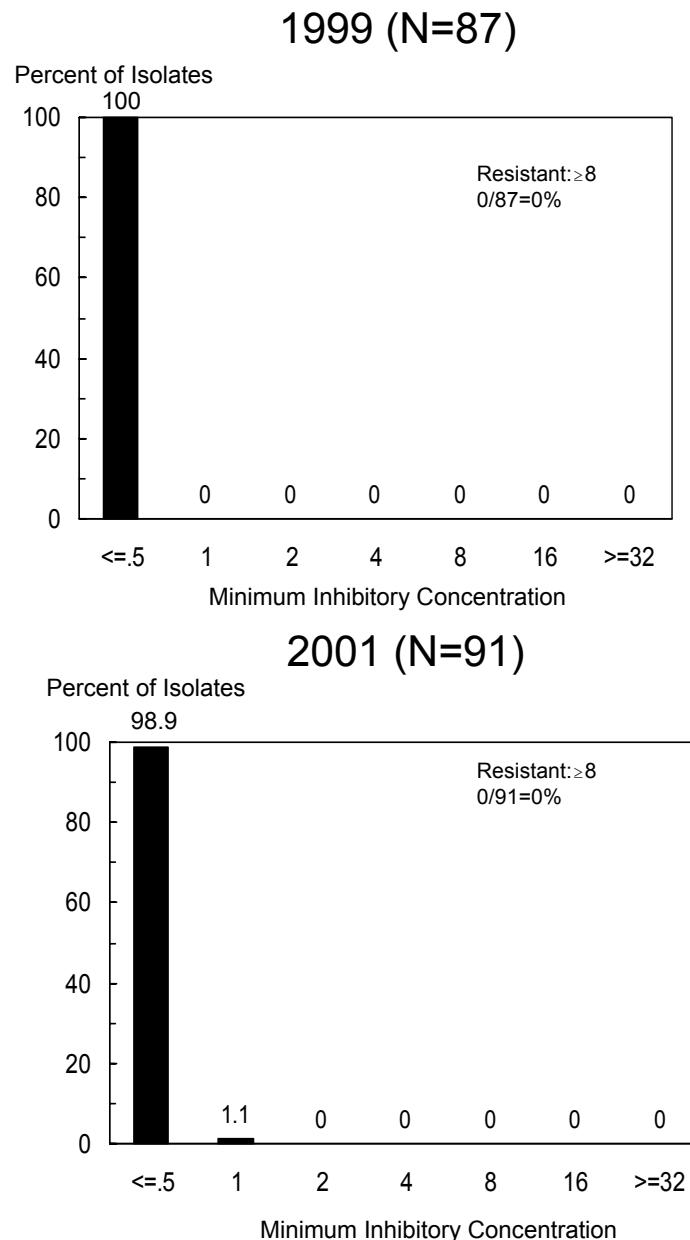


2000 (N=75)



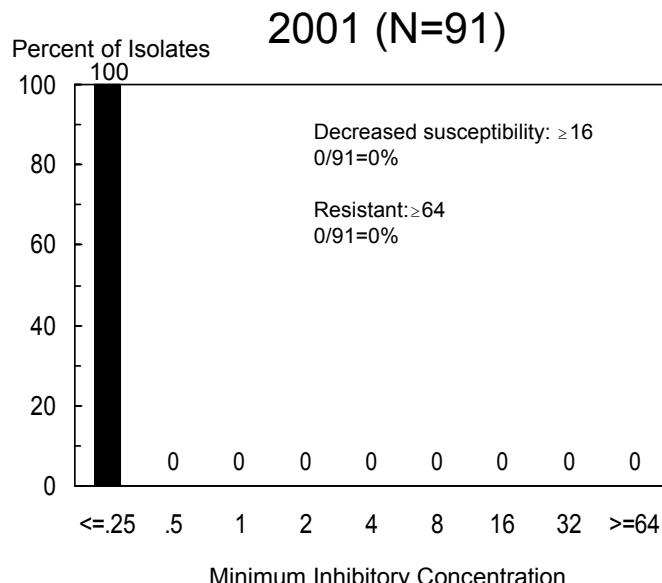
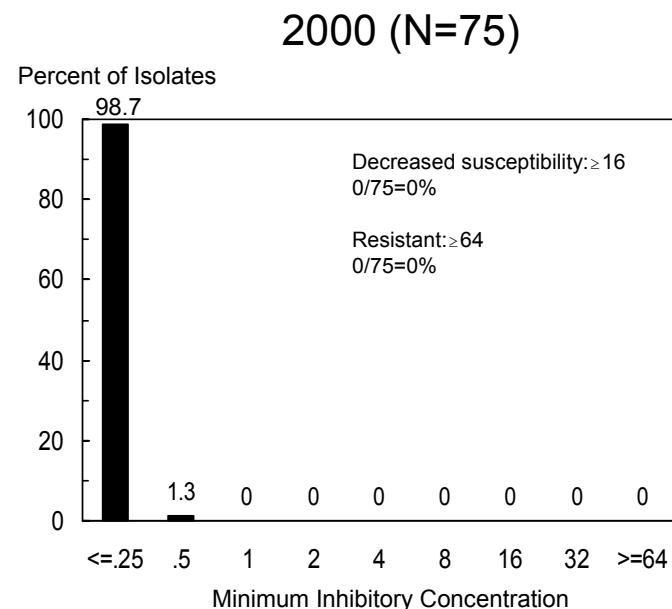
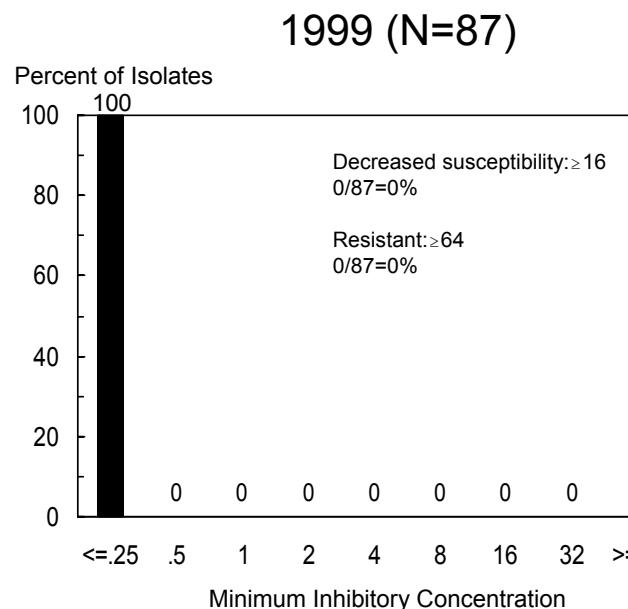
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Figure 16f. MICs for ceftiofur among *Shigella flexneri* isolates, 1999-2001



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Figure 16g. MICs for ceftriaxone* among *Shigella flexneri* isolates, 1999-2001

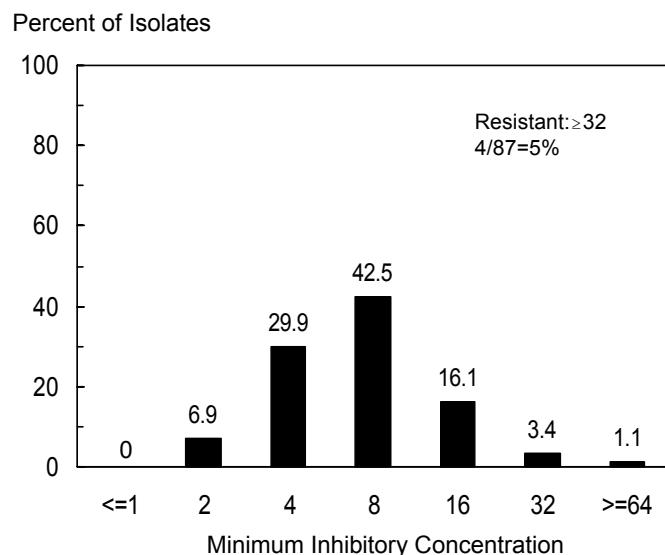


*Sensititre® results only

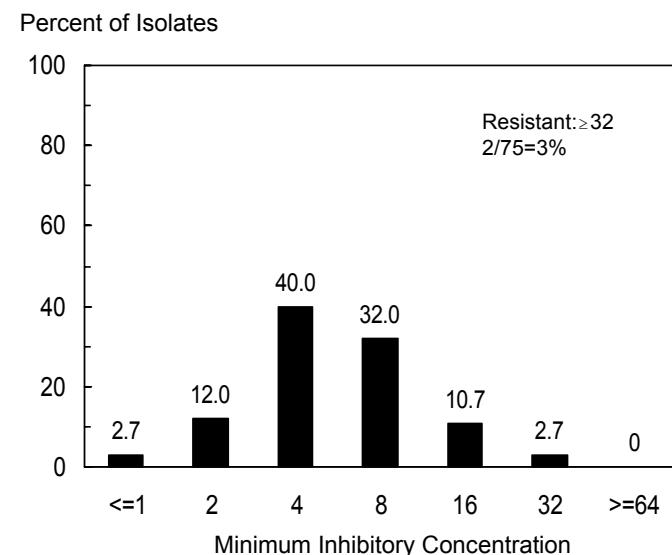
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Figure 16h. MICs for cephalothin among *Shigella flexneri* isolates, 1999-2001

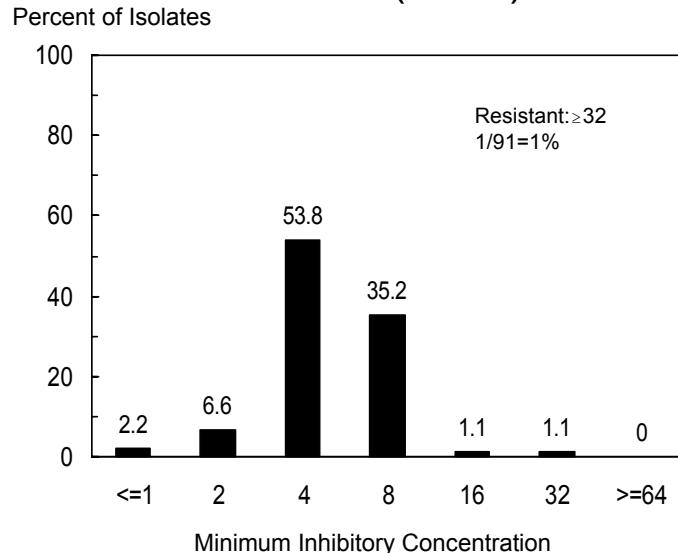
1999 (N=87)



2000 (N=75)



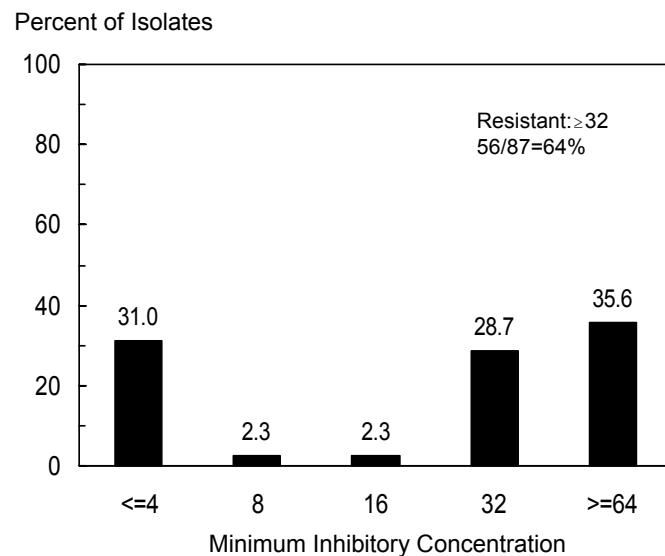
2001 (N=91)



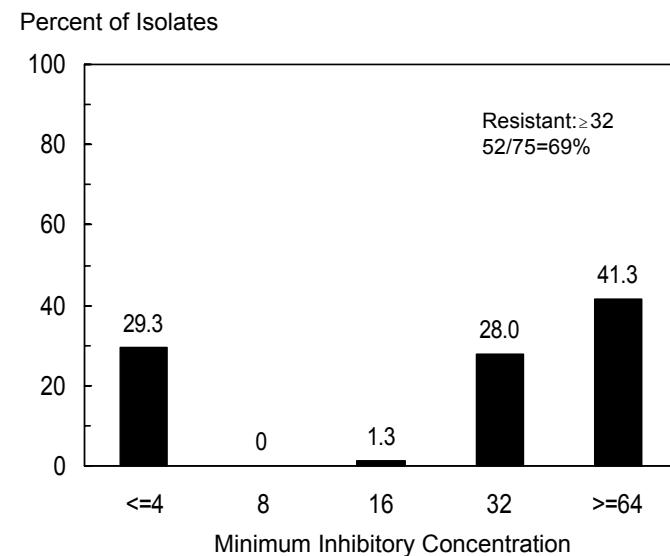
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Figure 16i. MICs for chloramphenicol among *Shigella flexneri* isolates, 1999-2001

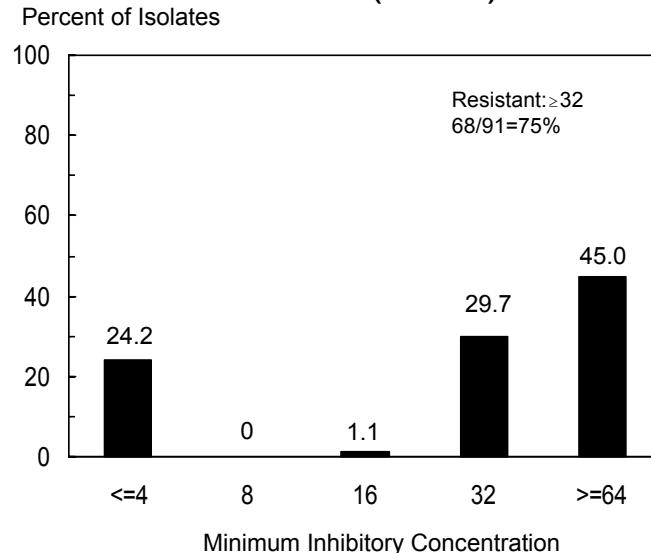
1999 (N=87)



2000 (N=75)



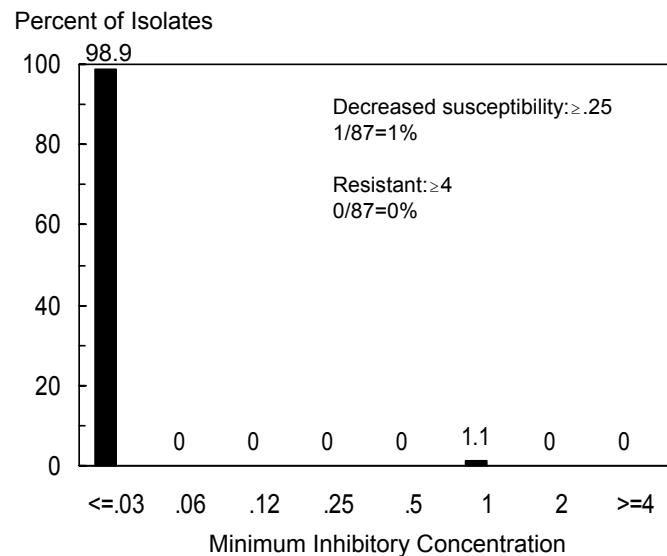
2001 (N=91)



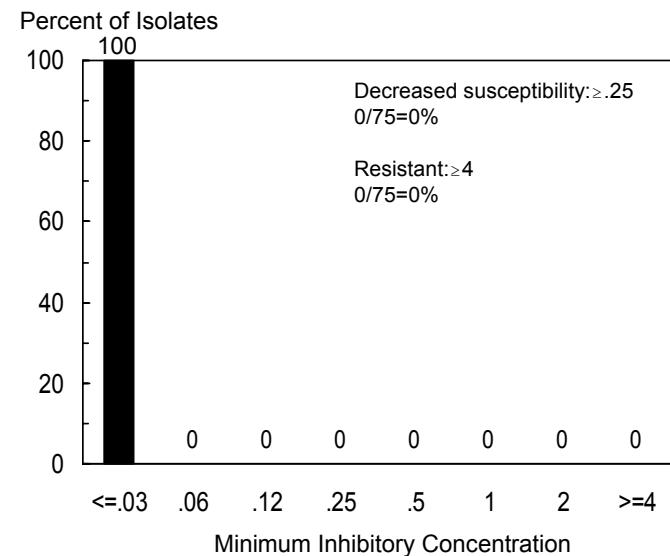
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Figure 16j. MICs for ciprofloxacin among *Shigella flexneri* isolates, 1999-2001

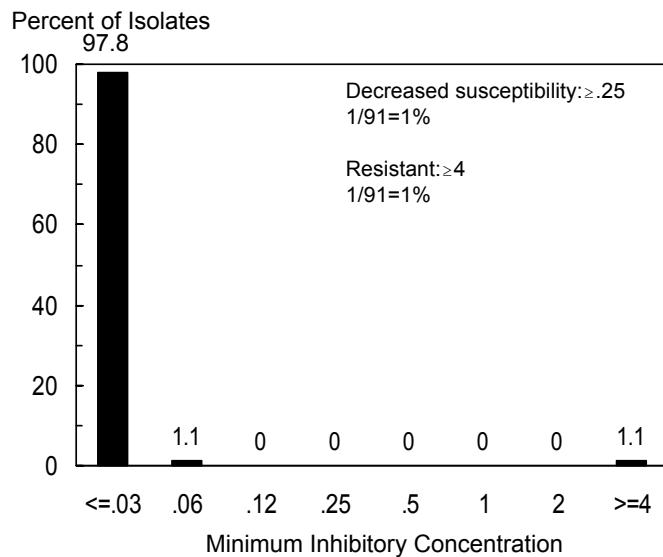
1999 (N=87)



2000 (N=75)

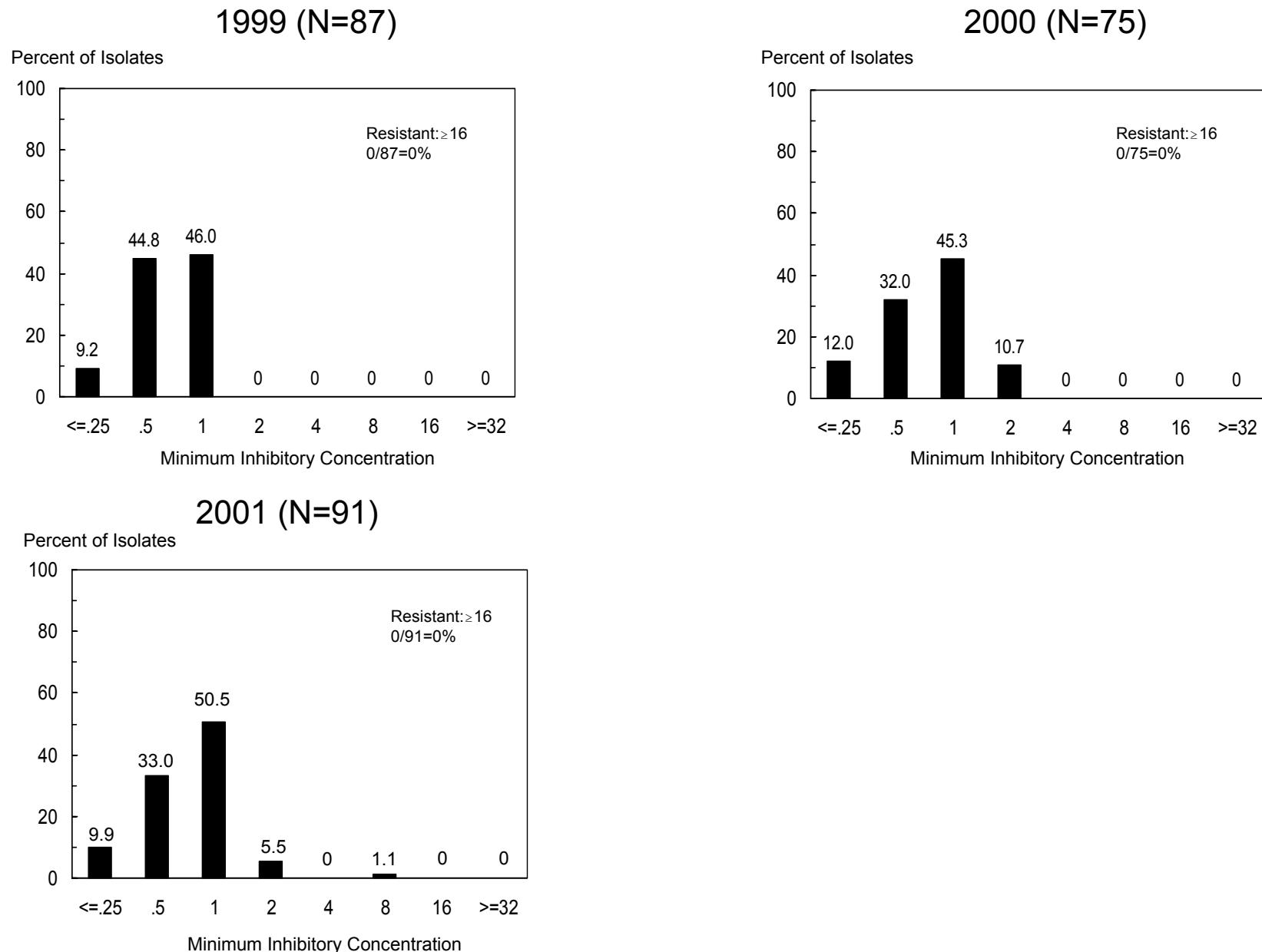


2001 (N=91)



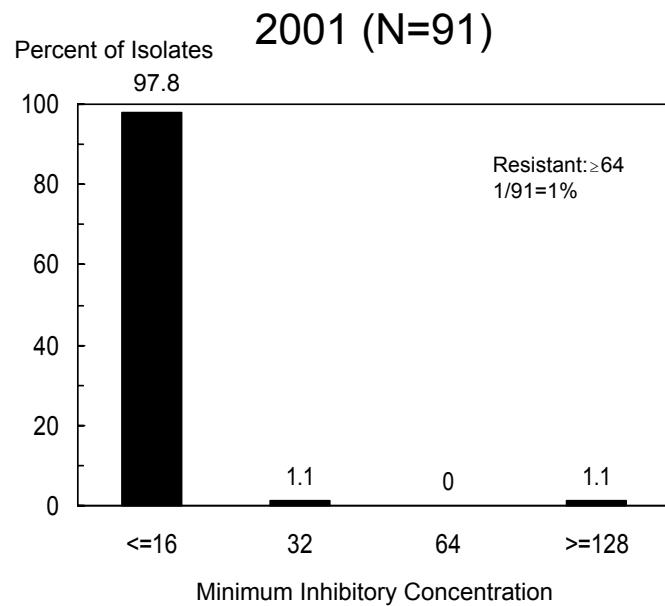
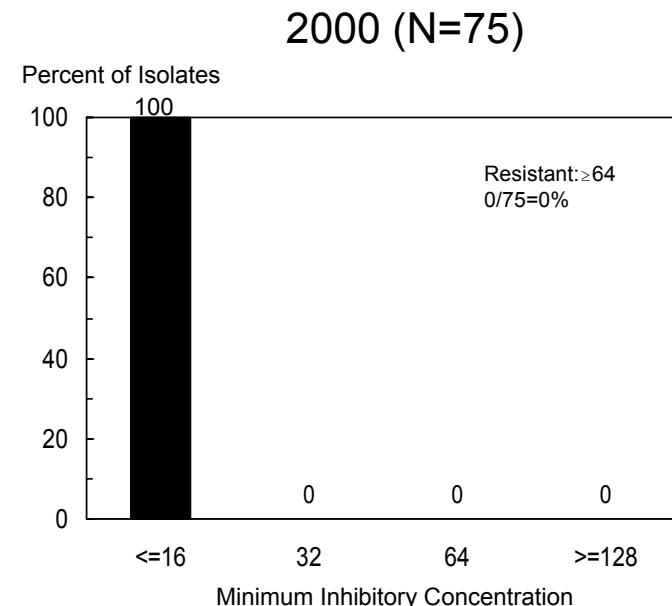
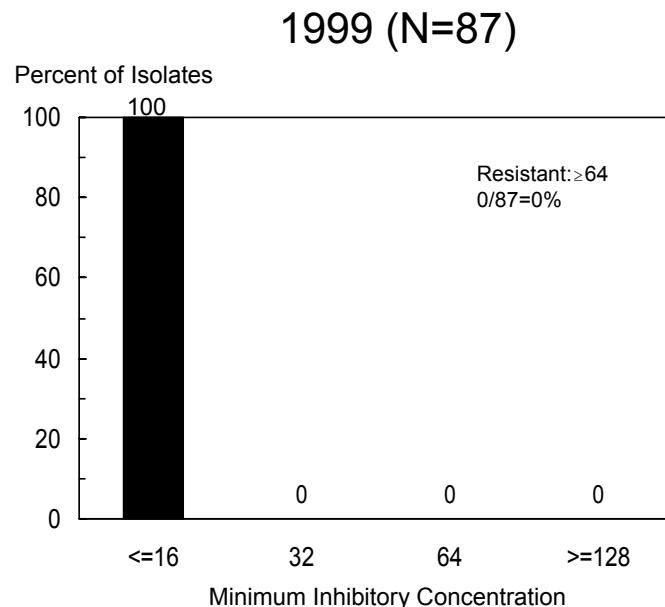
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Figure 16k. MICs for gentamicin among *Shigella flexneri* isolates, 1999-2001



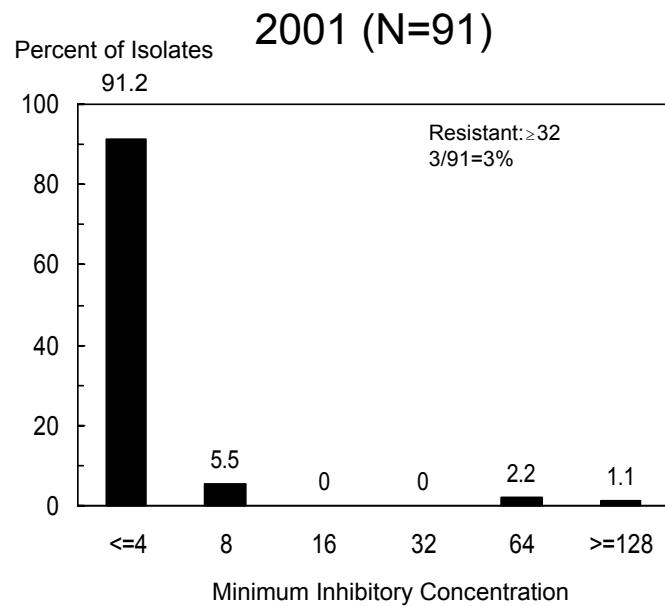
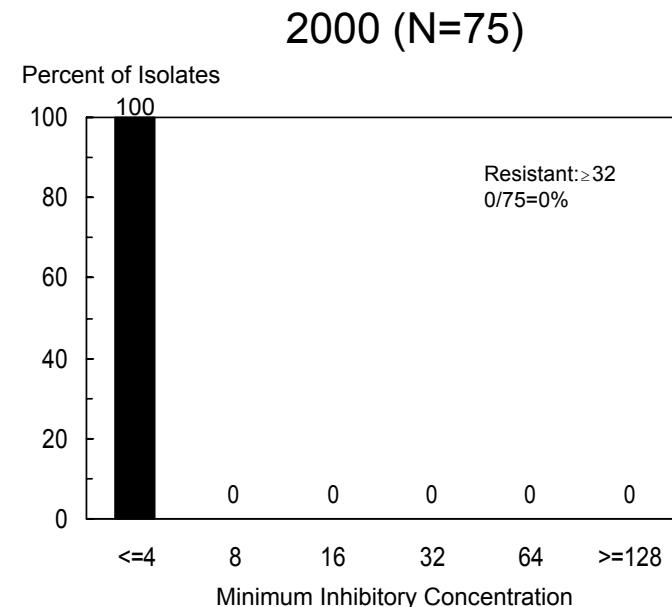
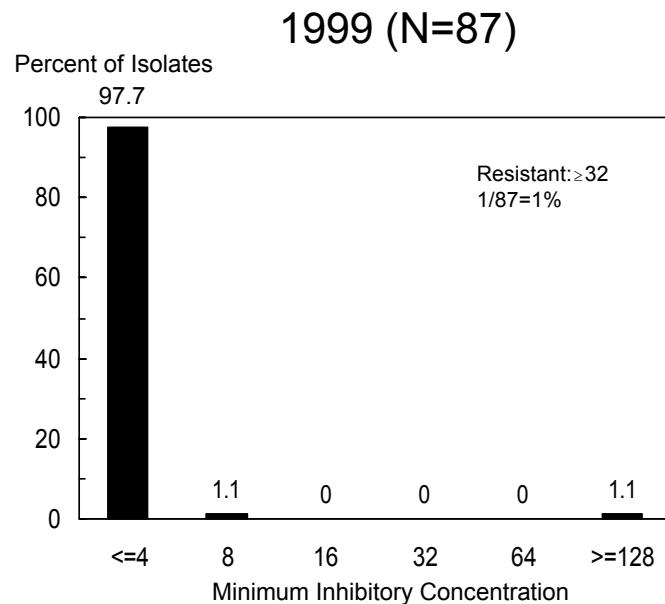
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Figure 16I. MICs for kanamycin among *Shigella flexneri* isolates, 1999-2001



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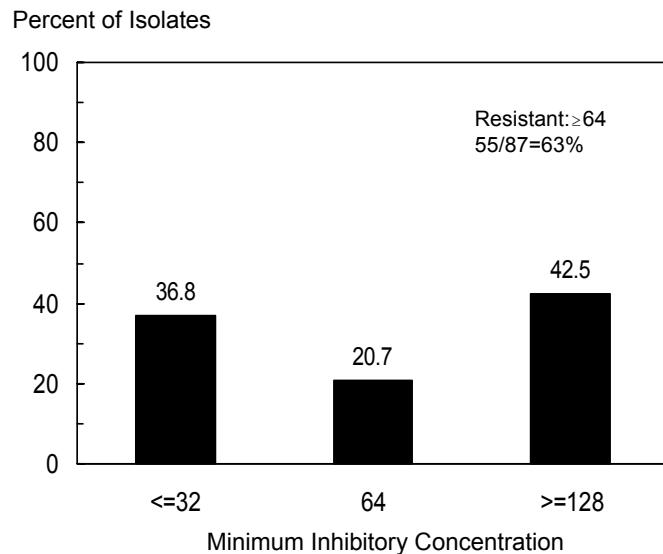
Figure 16m. MICs for nalidixic acid among *Shigella flexneri* isolates, 1999-2001



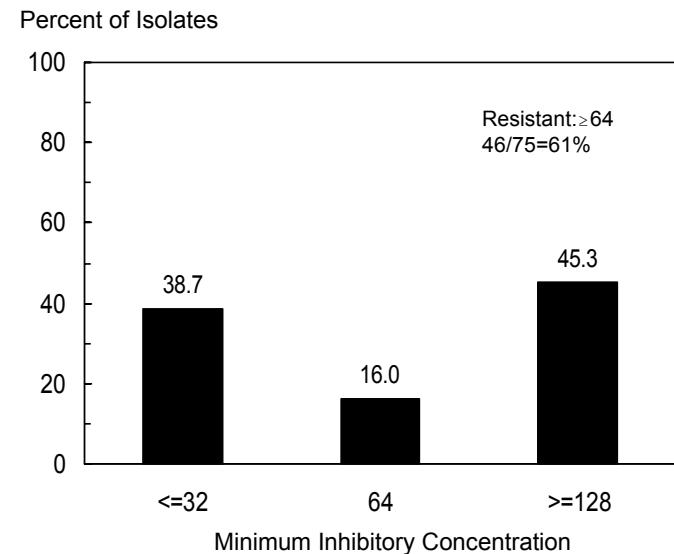
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Figure 16n. MICs for streptomycin among *Shigella flexneri* isolates, 1999-2001

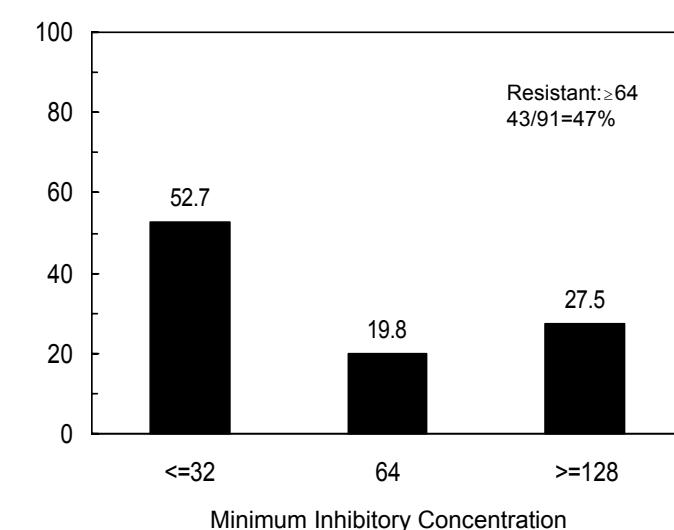
1999 (N=87)



2000 (N=75)



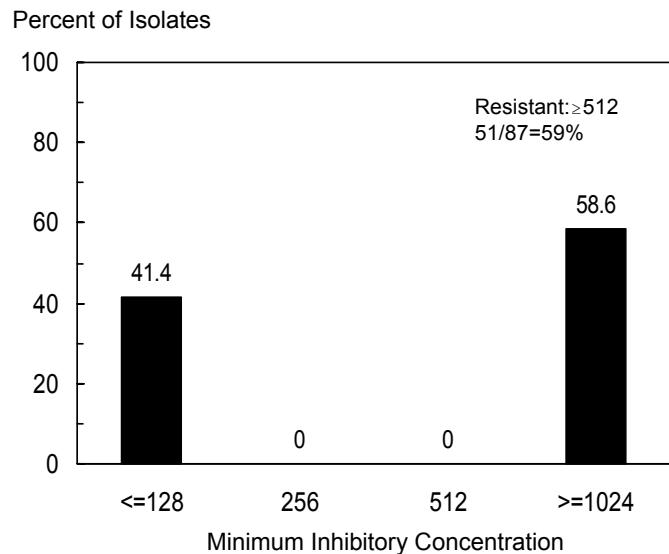
2001 (N=91)



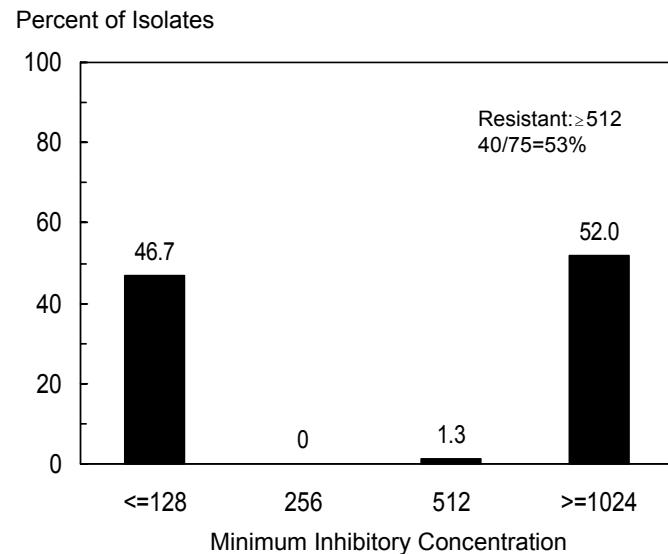
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Figure 16o. MICs for sulfamethoxazole among *Shigella flexneri* isolates, 1999-2001

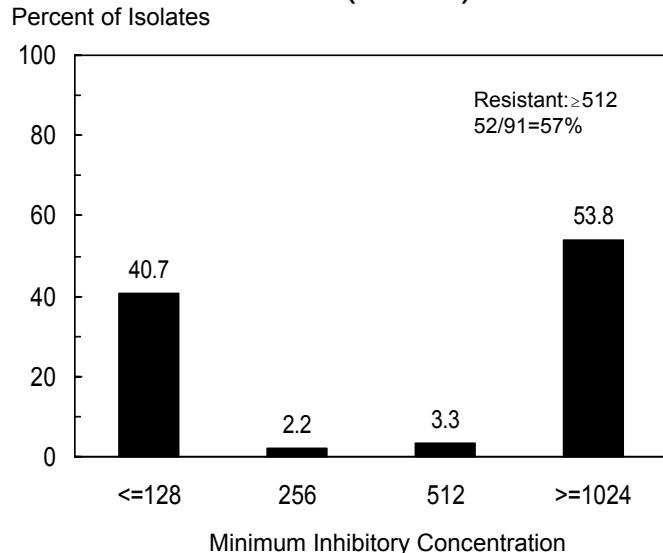
1999 (N=87)



2000 (N=75)



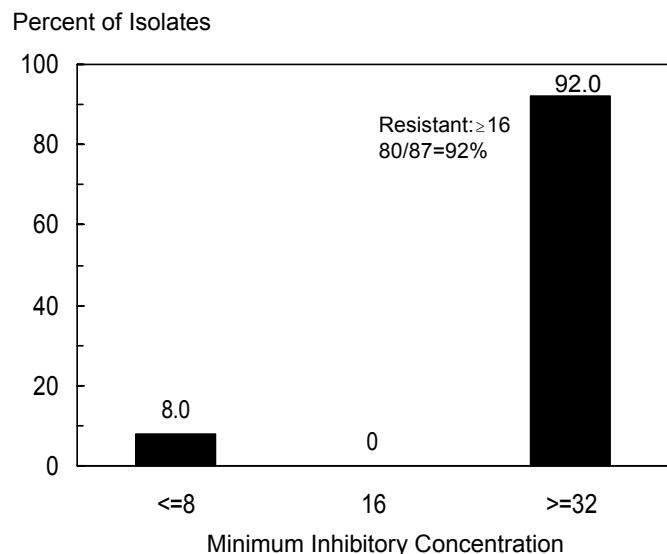
2001 (N=91)



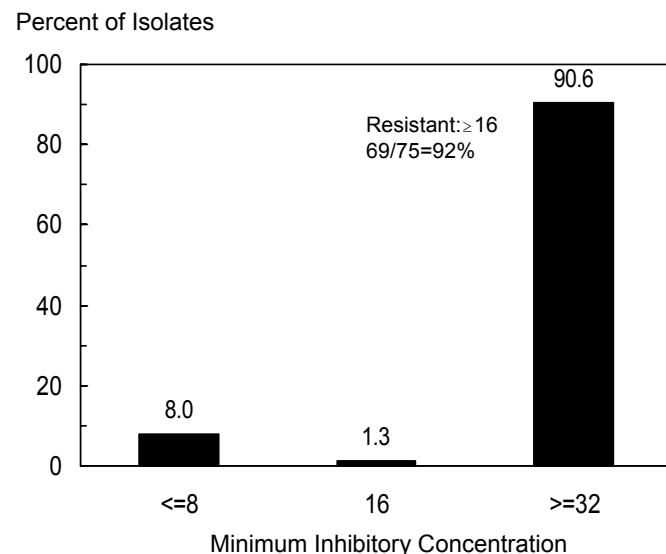
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Figure 16p. MICs for tetracycline among *Shigella flexneri* isolates, 1999-2001

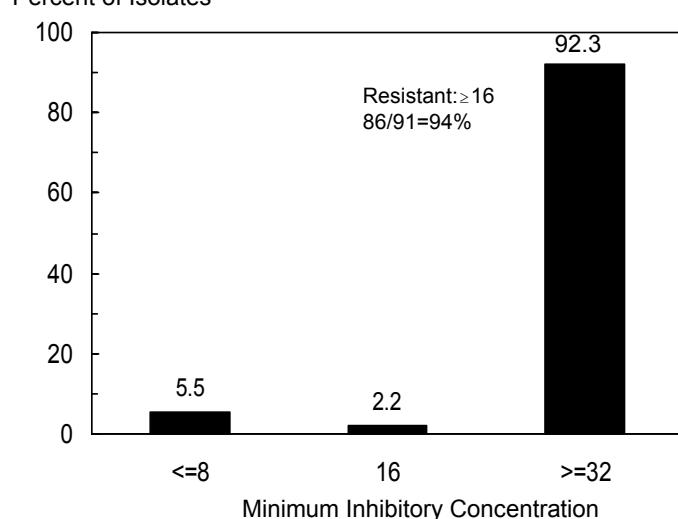
1999 (N=87)



2000 (N=75)



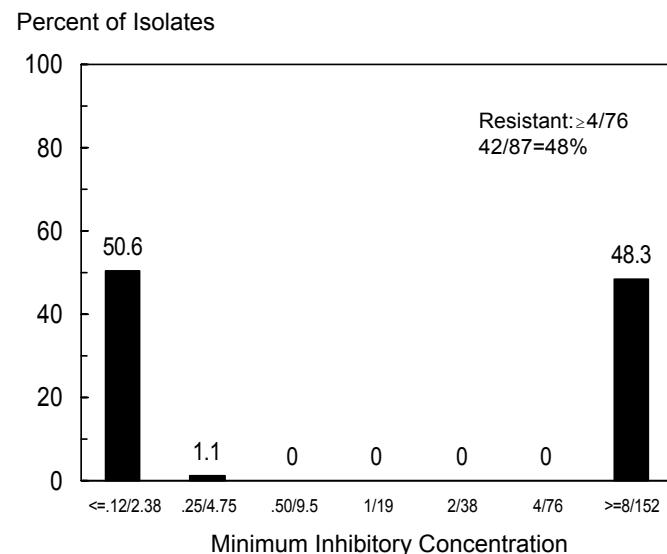
2001 (N=91)



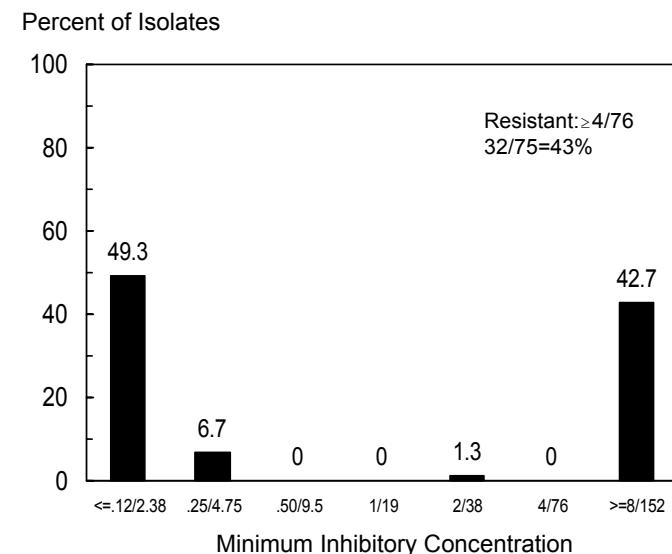
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Figure 16q. MICs for trimethoprim-sulfamethoxazole among *Shigella flexneri* isolates, 1999-2001

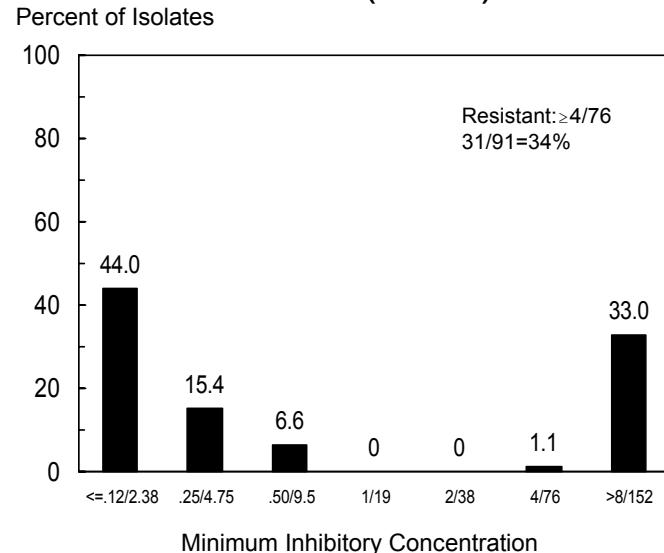
1999 (N=87)



2000 (N=75)

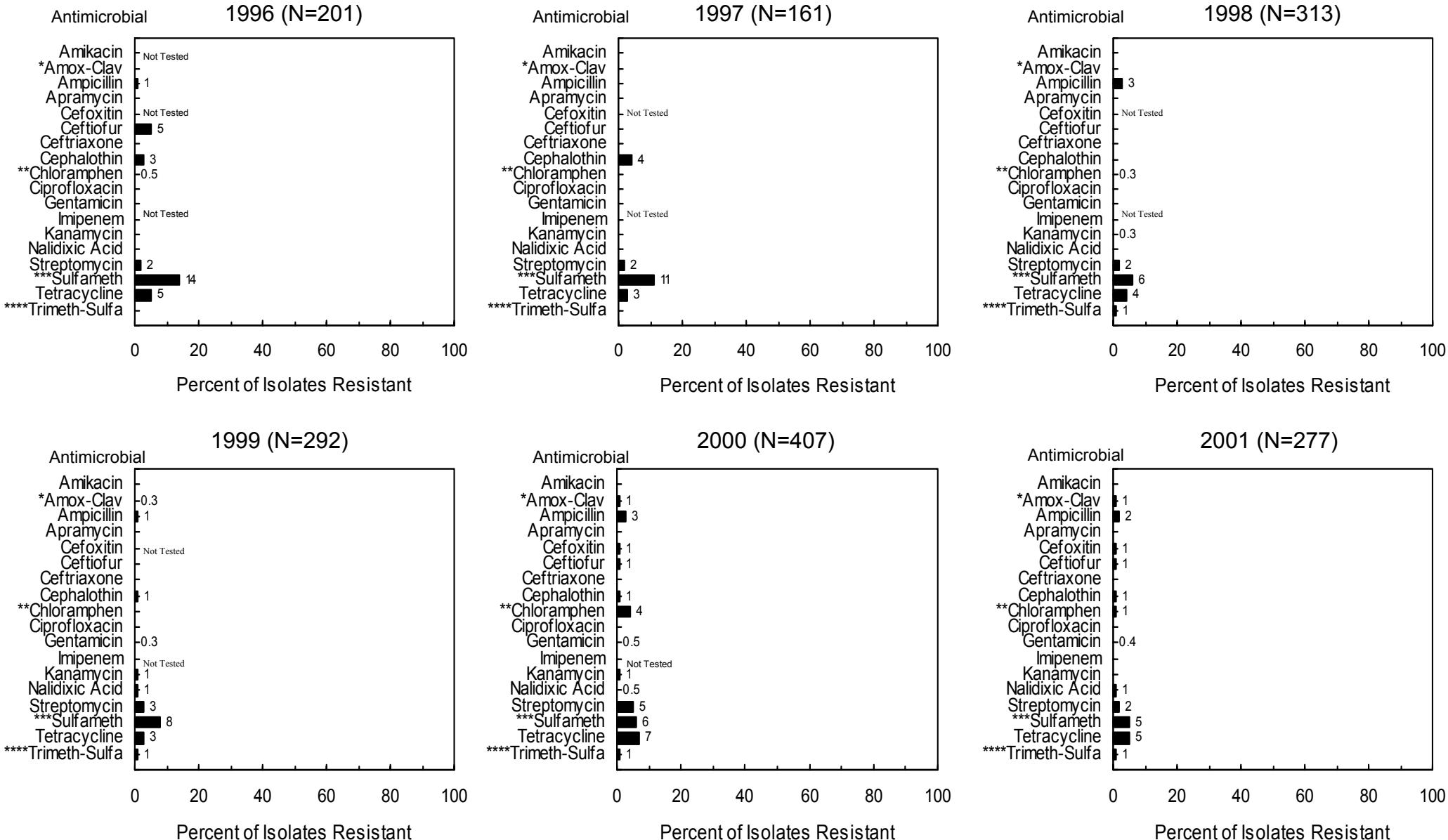


2001 (N=91)



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Figure 17. Resistance among *E. coli* O157 isolates, 1996-2001

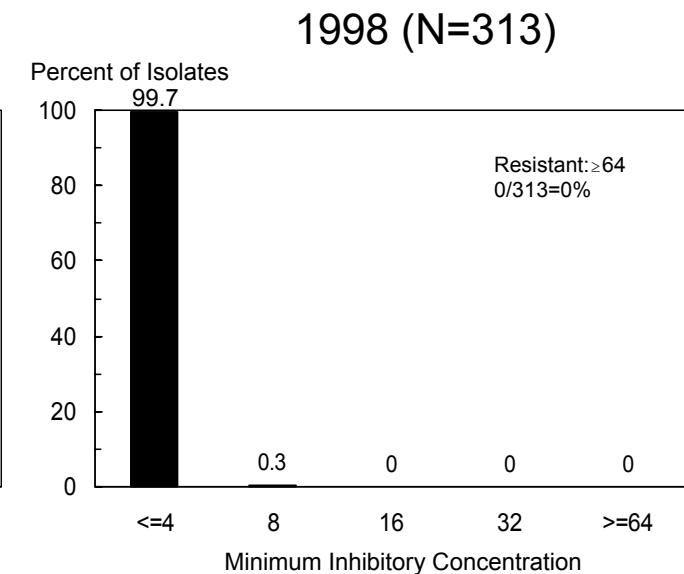
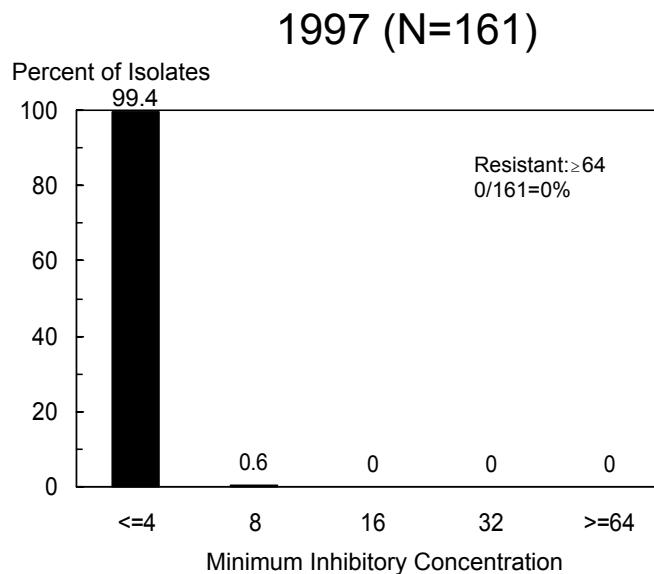


*Amox-Clav=Amoxicillin-Clavulanic Acid **Chloramphen=Chloramphenicol ***Sulfameth=Sulfamethoxazole ****Trimeth-Sulfa=Trimethoprim-Sulfamethoxazole

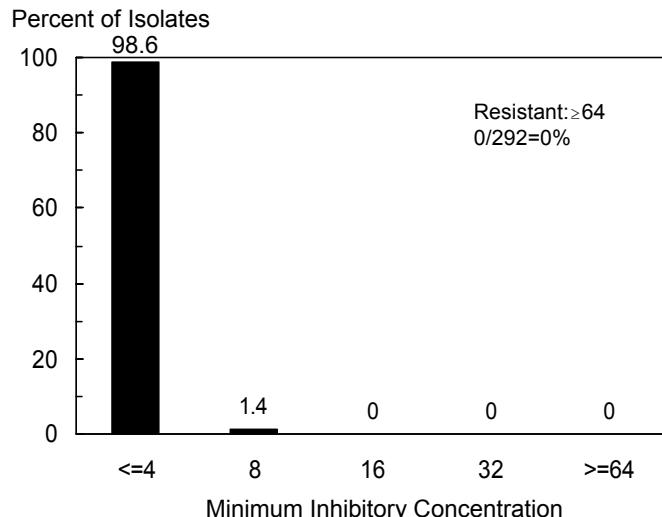
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Figure 18a. MICs for amikacin among *E. coli* O157 isolates, 1996-2001

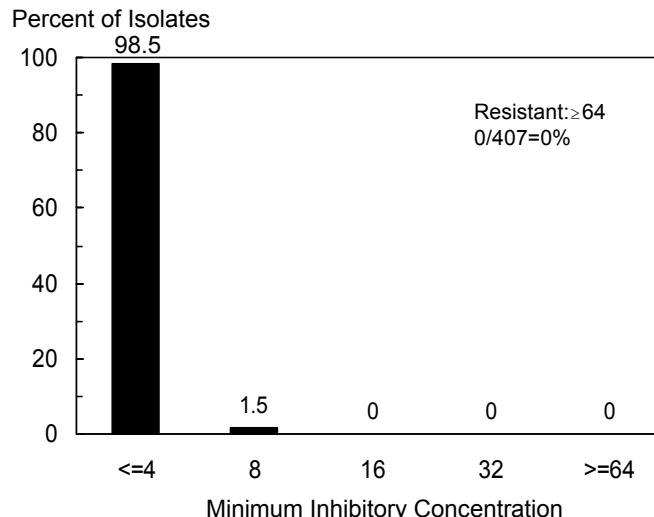
Not Tested in 1996



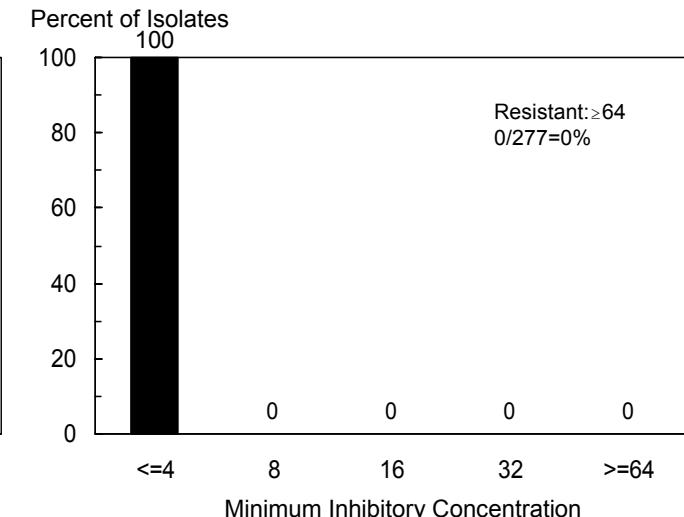
1999 (N=292)



2000 (N=407)

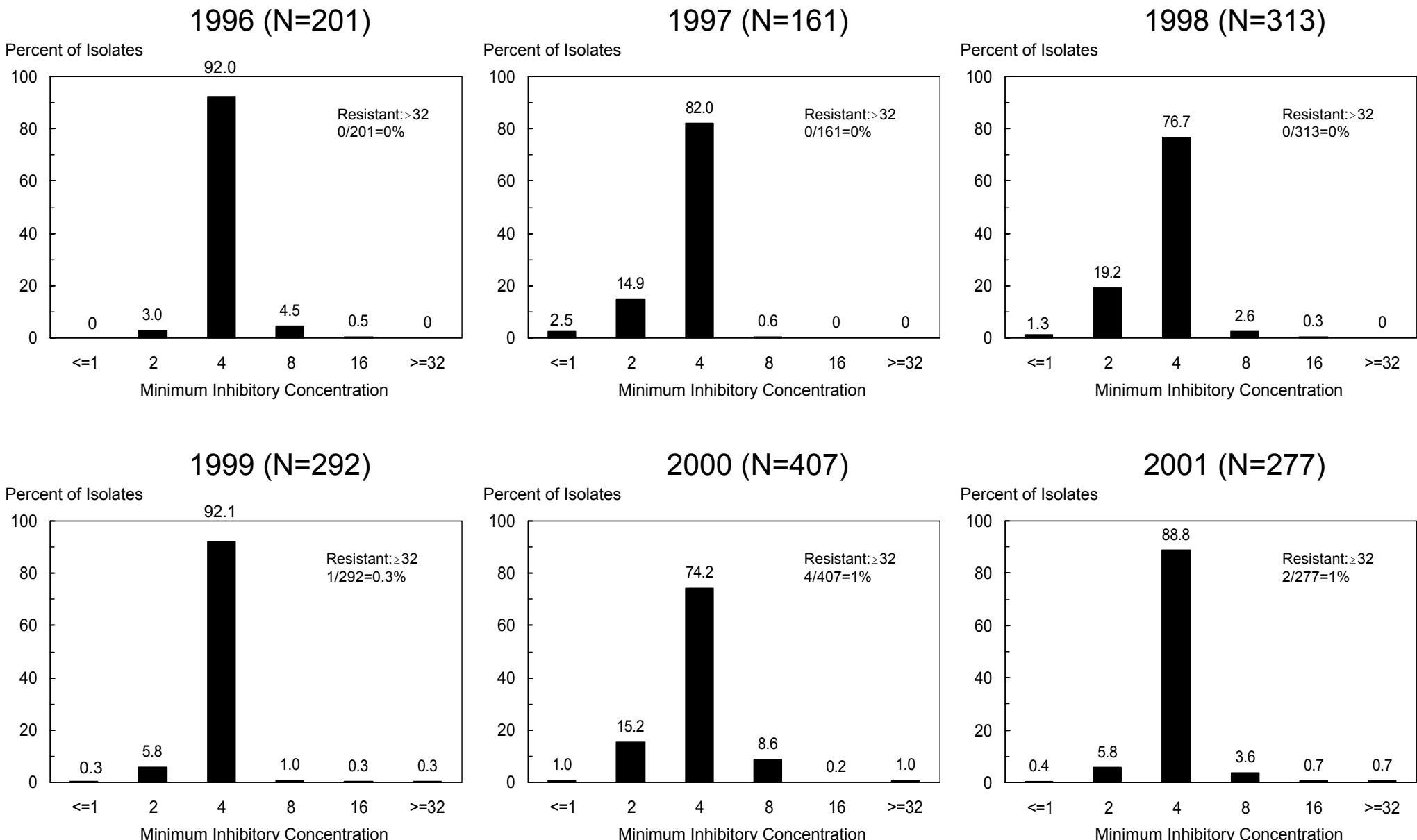


2001 (N=277)



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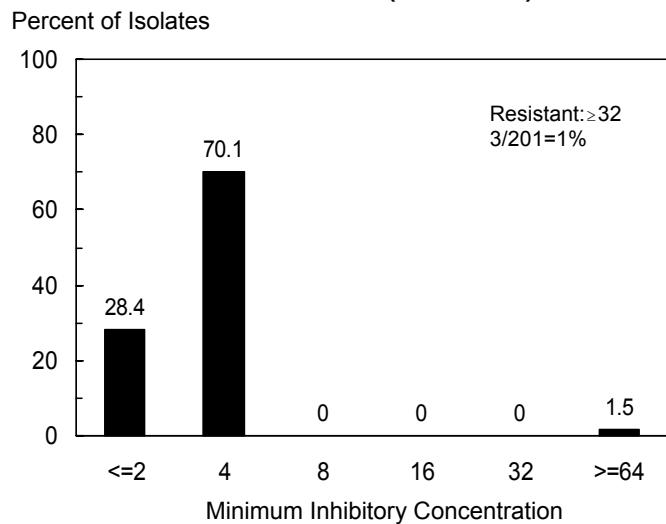
Figure 18b. MICs for amoxicillin-clavulanic acid among *E. coli* O157 isolates, 1996-2001



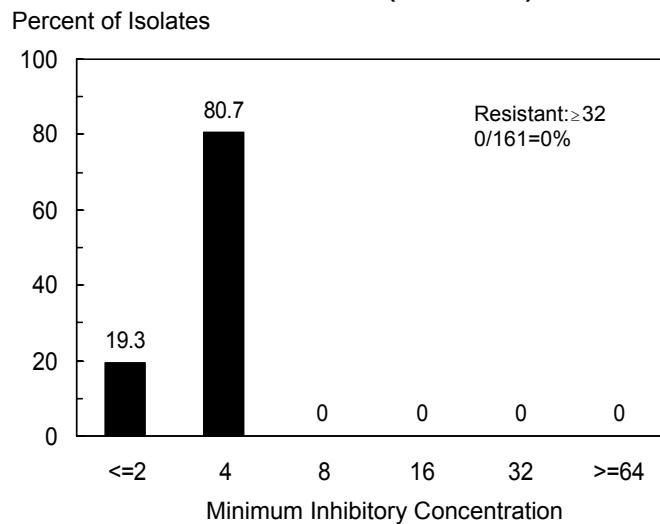
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Figure 18c. MICs for ampicillin among *E. coli* O157 isolates, 1996-2001

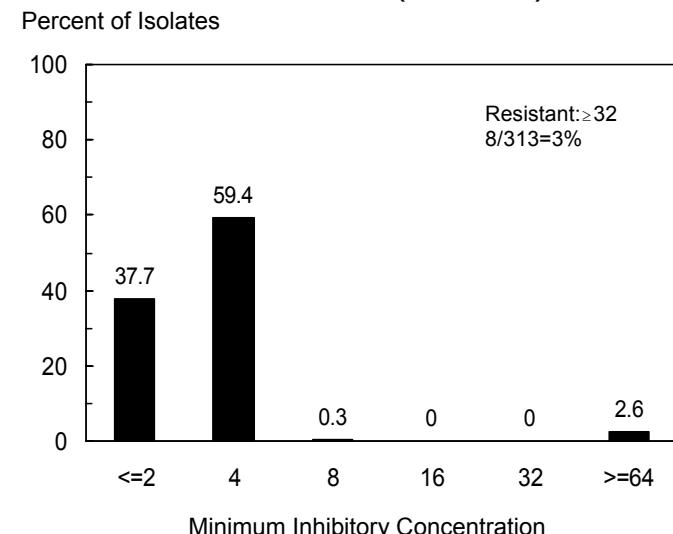
1996 (N=201)



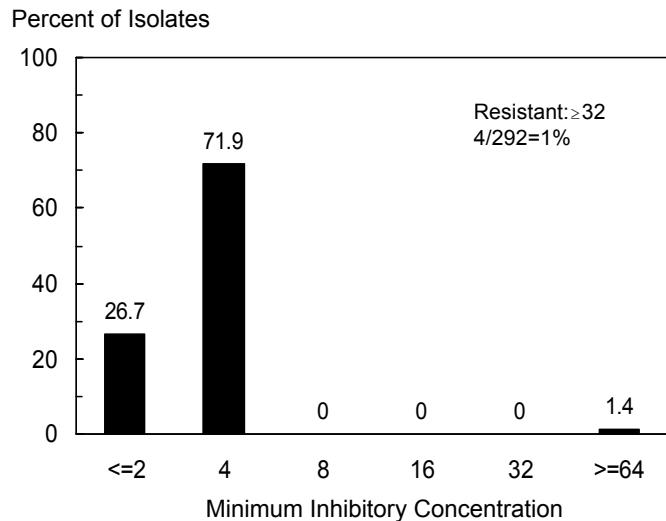
1997 (N=161)



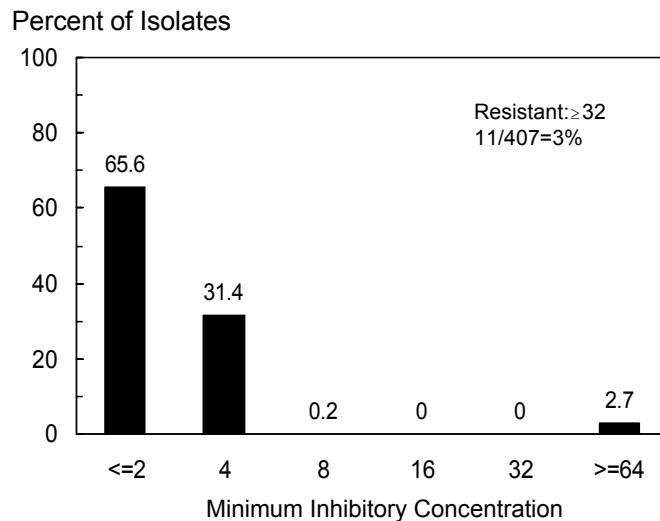
1998 (N=313)



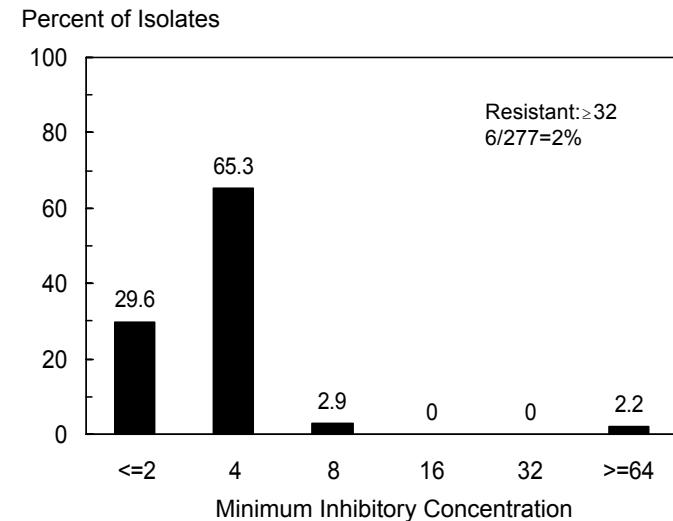
1999 (N=292)



2000 (N=407)



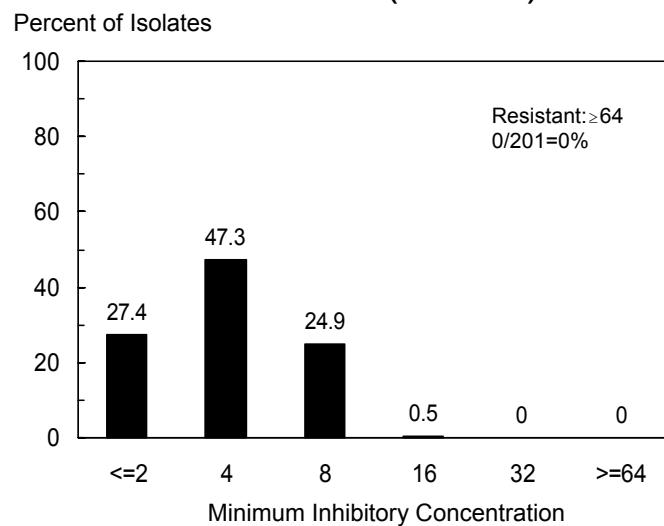
2001 (N=277)



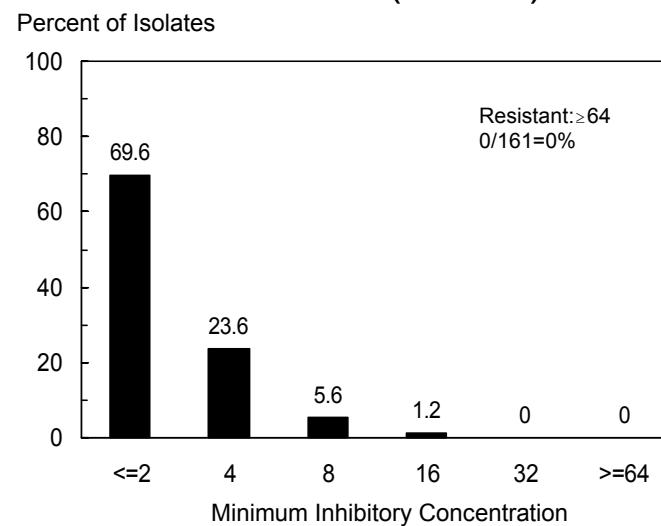
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Figure 18d. MICs for apramycin among *E. coli* O157 isolates, 1996-2001

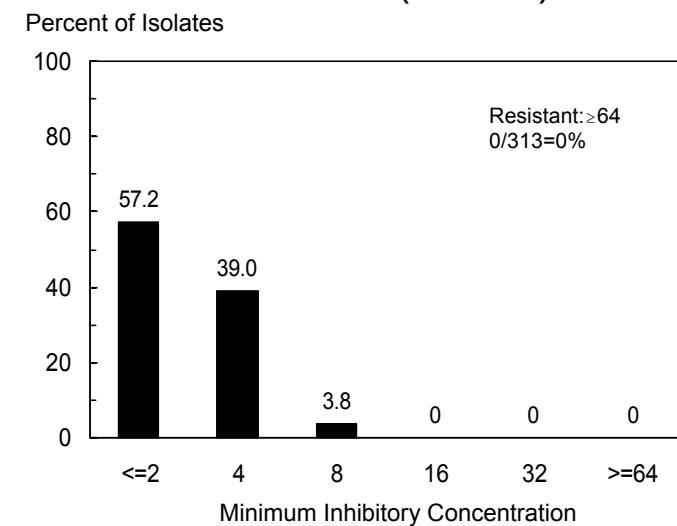
1996 (N=201)



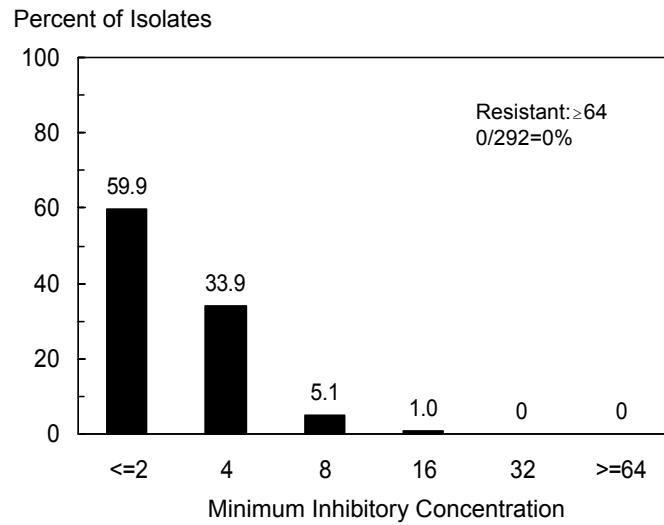
1997 (N=161)



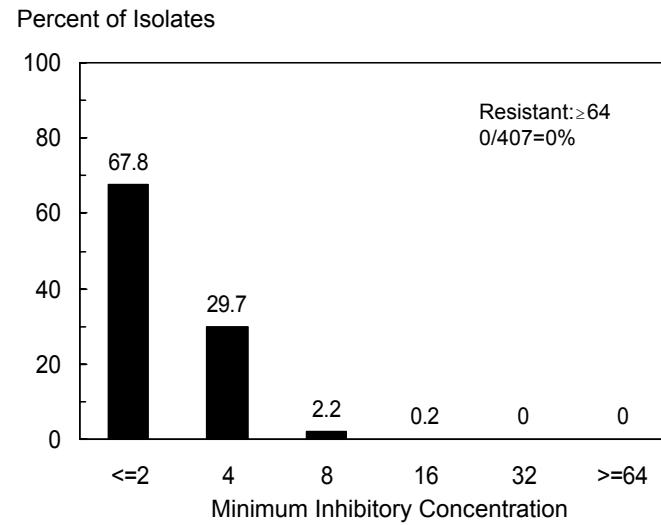
1998 (N=313)



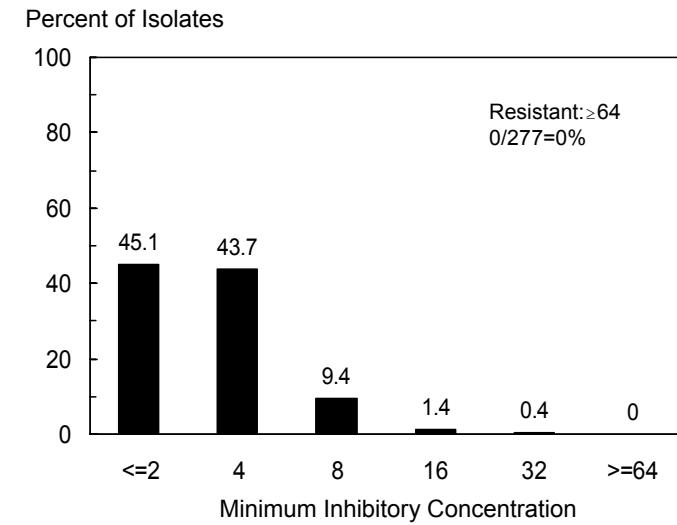
1999 (N=292)



2000 (N=407)



2001 (N=277)

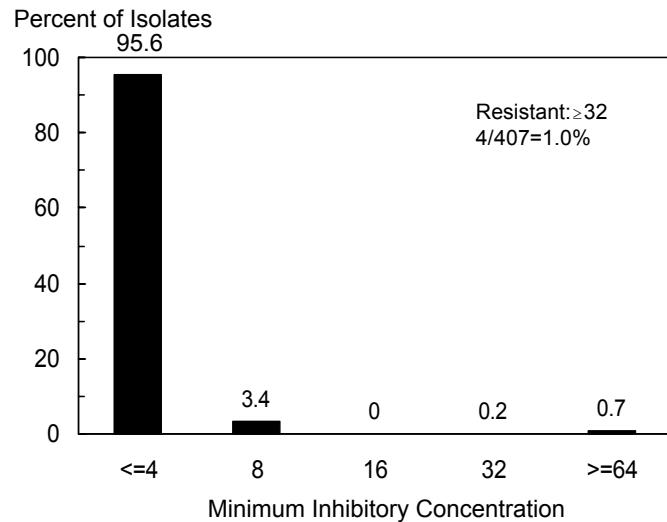


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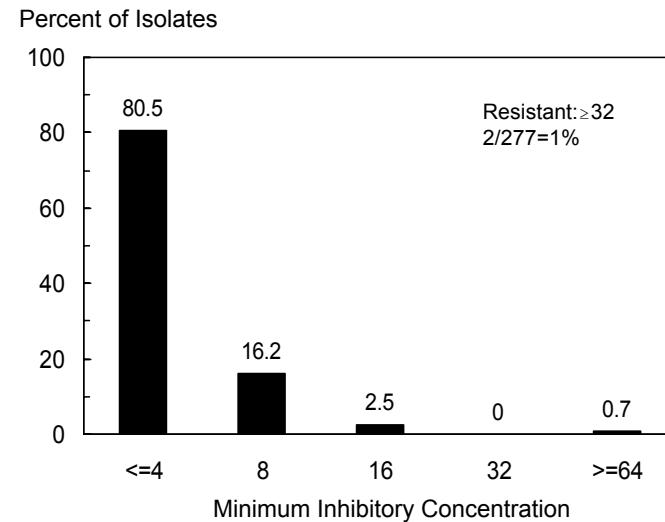
Figure 18e. MICs for cefoxitin among *E. coli* O157 isolates, 1996-2001

Not Tested in 1996-1999

2000 (N=407)

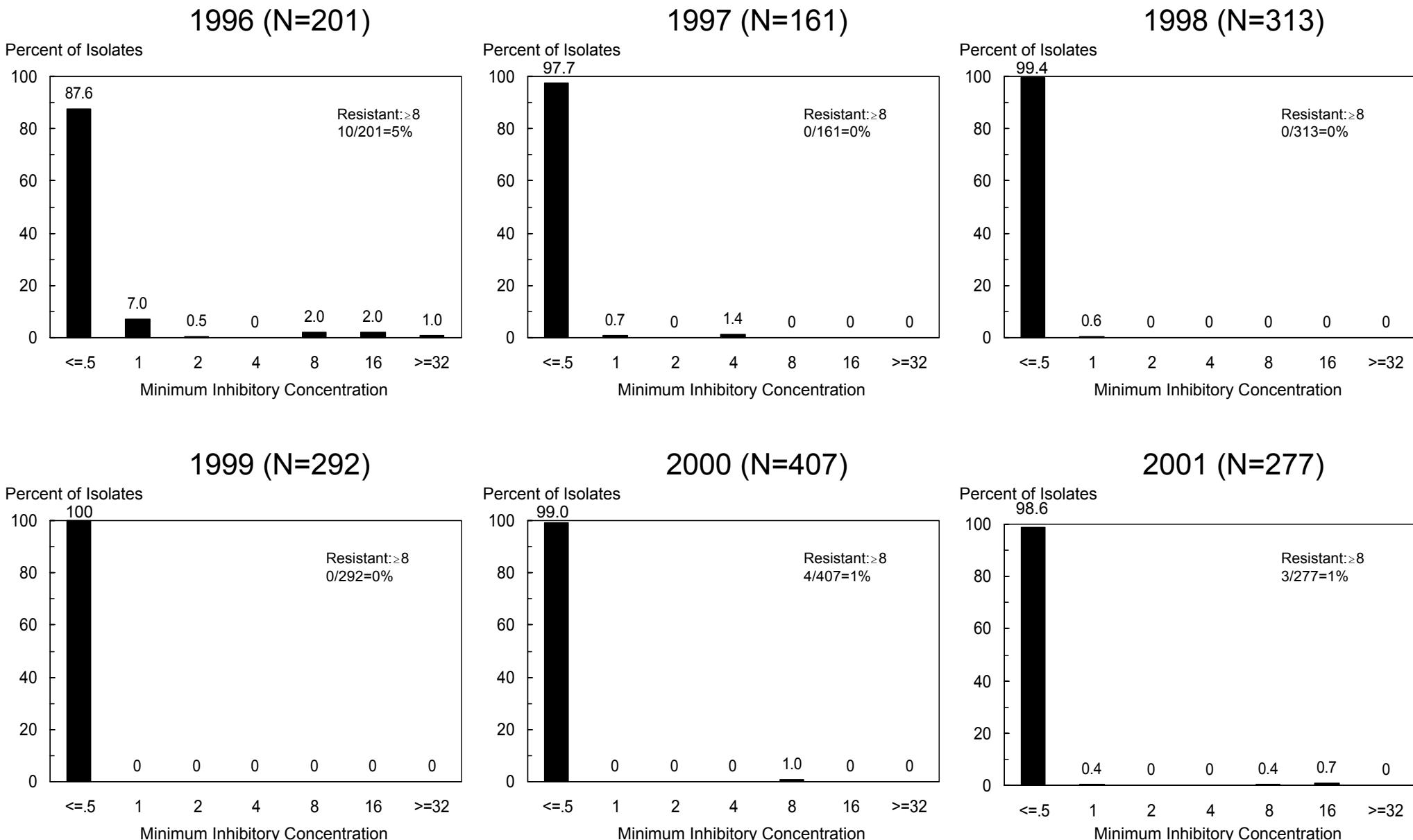


2001 (N=277)



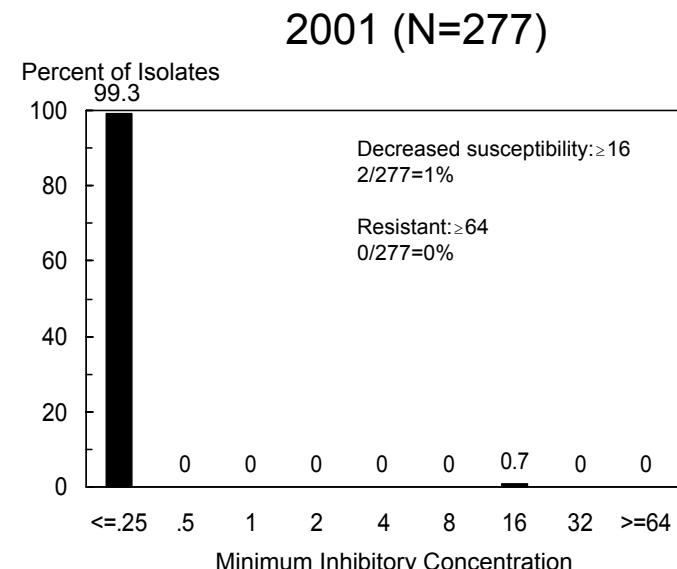
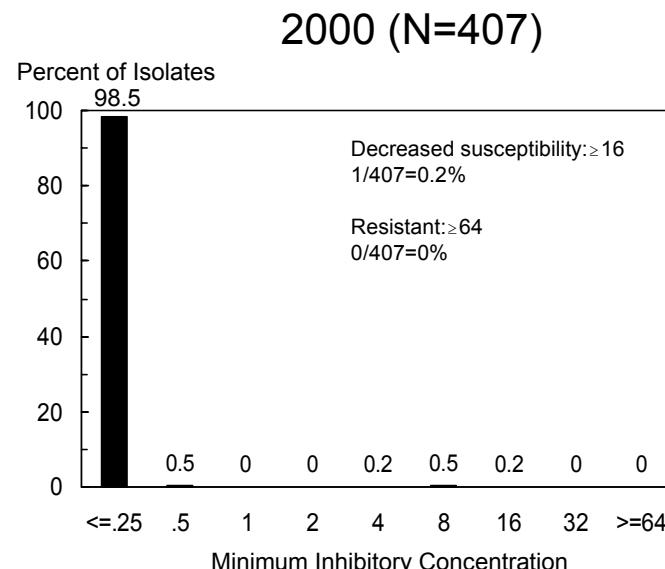
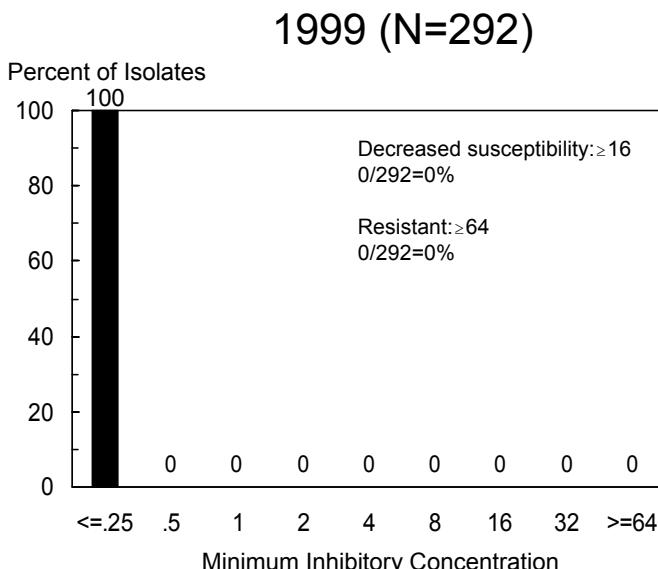
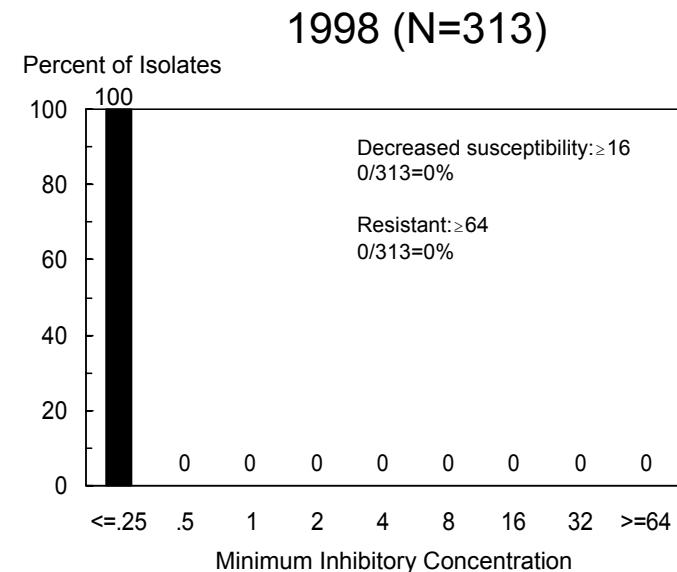
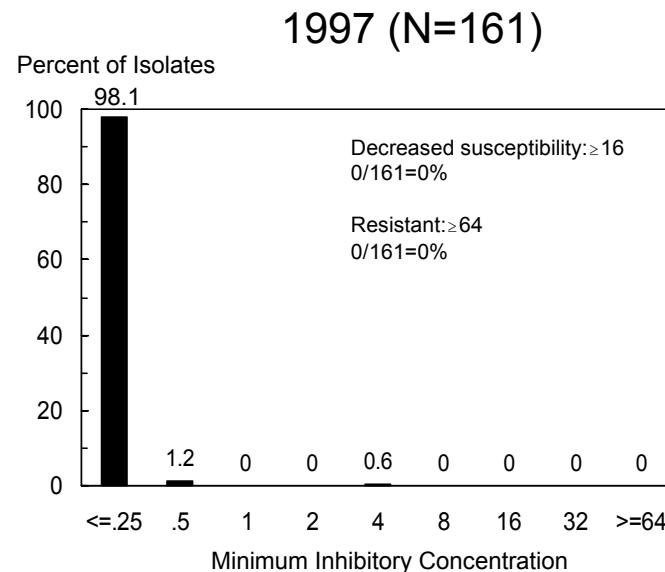
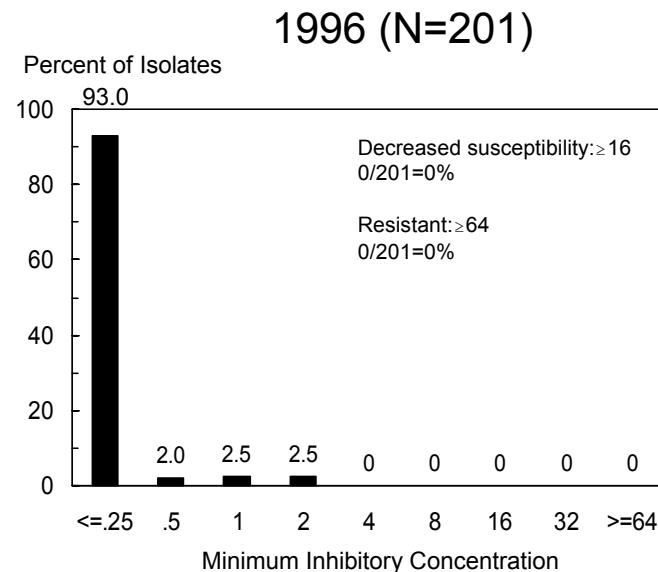
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Figure 18f. MICs for ceftiofur among *E. coli* O157 isolates, 1996-2001



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Figure 18g. MICs for ceftriaxone* among *E. coli* O157 isolates, 1996-2001

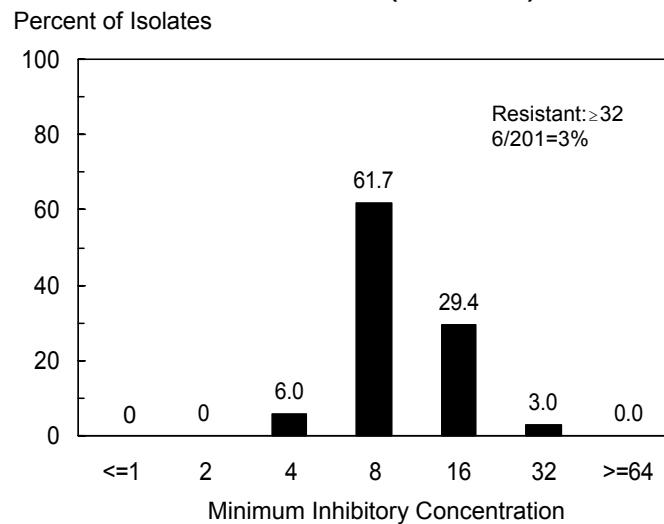


*Sensititre® results only

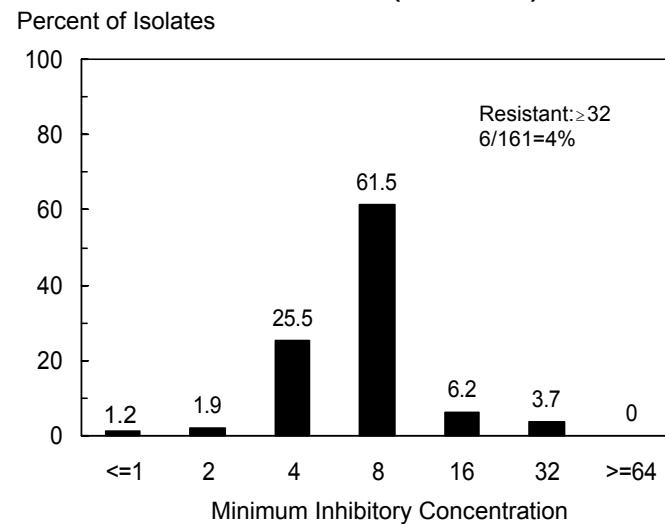
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Figure 18h. MICs for cephalothin among *E. coli* O157 isolates, 1996-2001

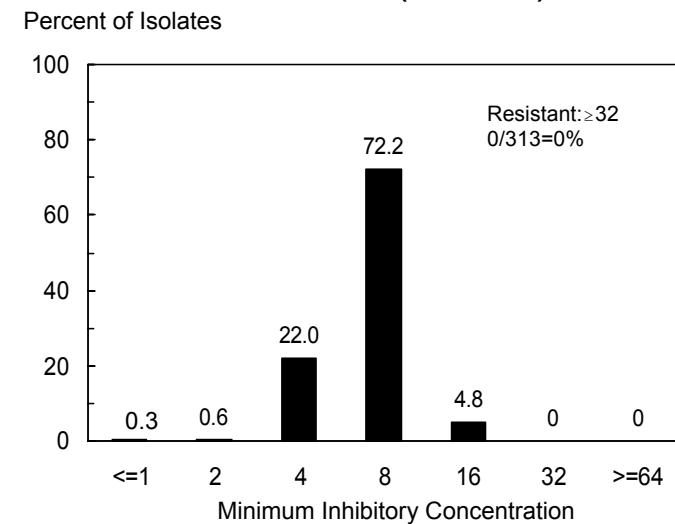
1996 (N=201)



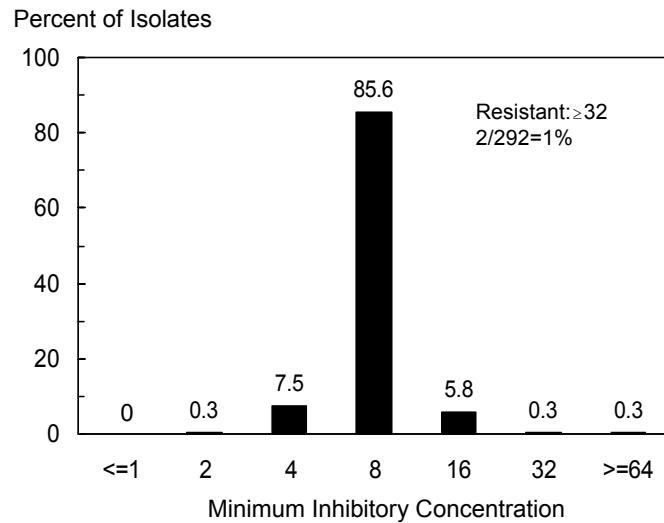
1997 (N=161)



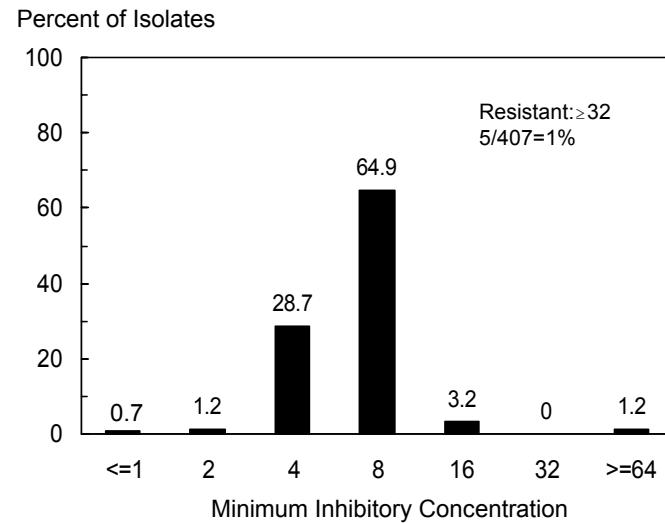
1998 (N=313)



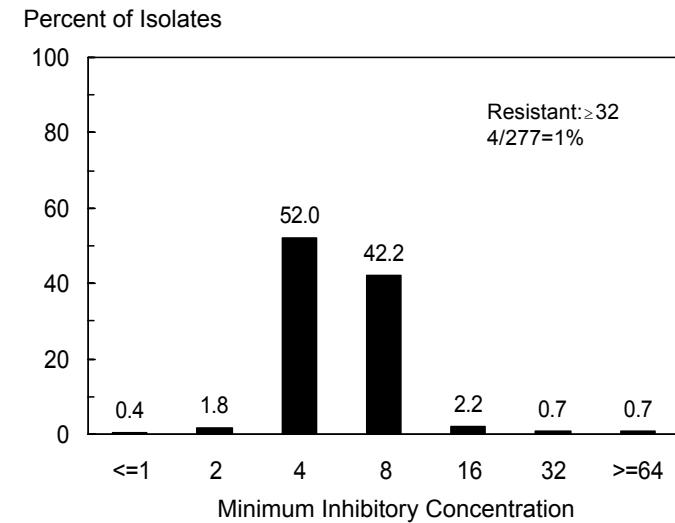
1999 (N=292)



2000 (N=407)

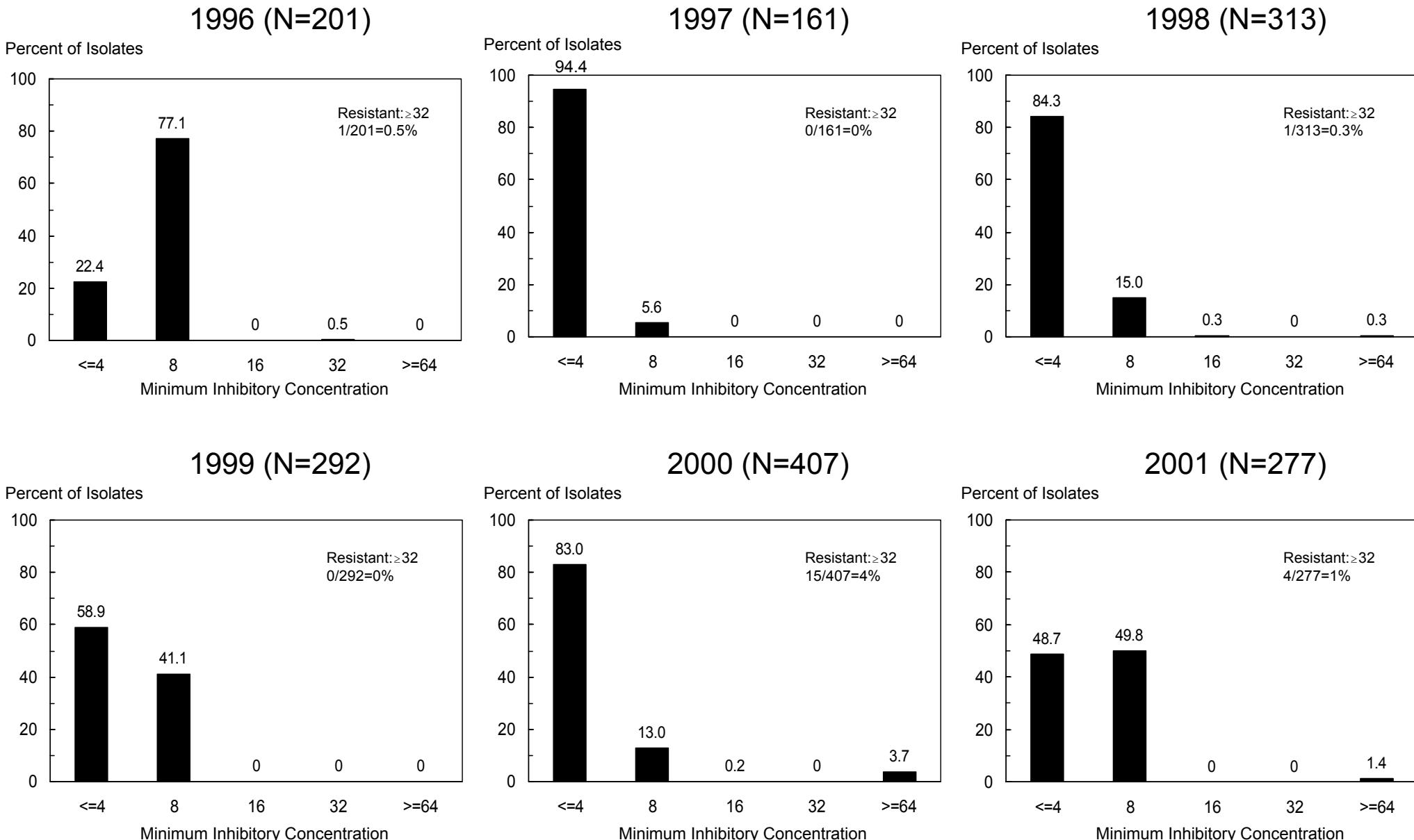


2001 (N=277)



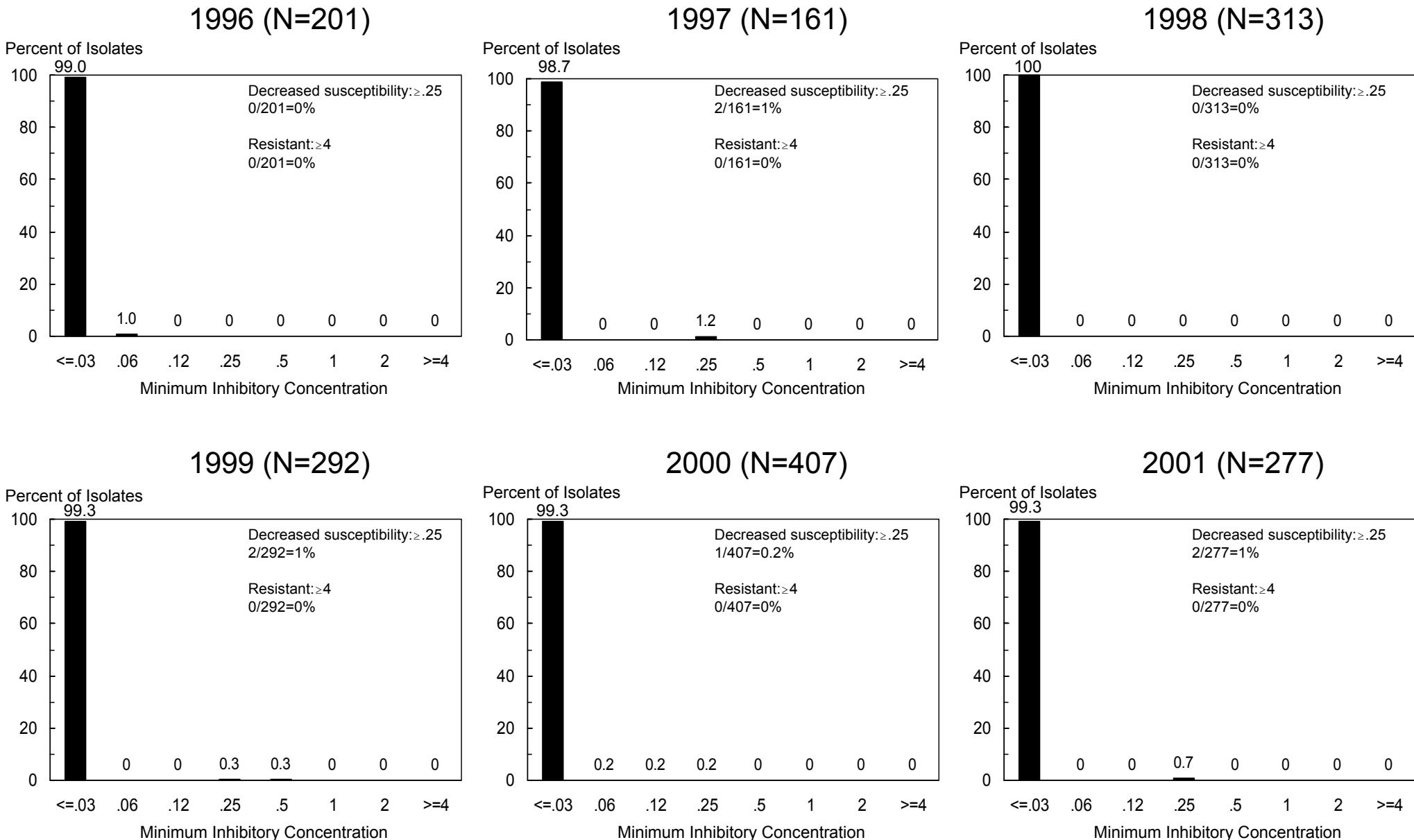
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Figure 18i. MICs for chloramphenicol among *E. coli* O157 isolates, 1996-2001



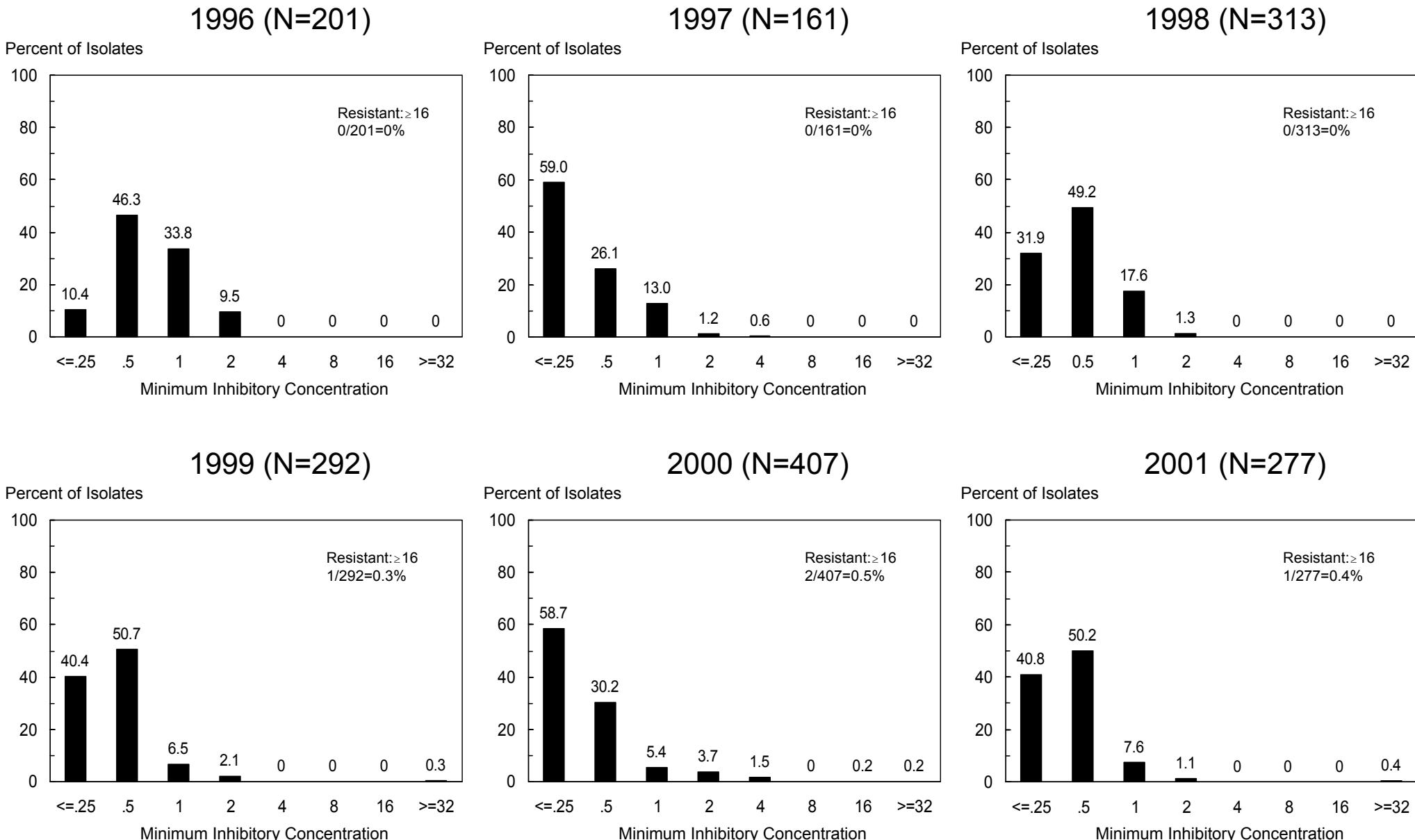
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Figure 18j. MICs for ciprofloxacin among *E. coli* O157 isolates, 1996-2001



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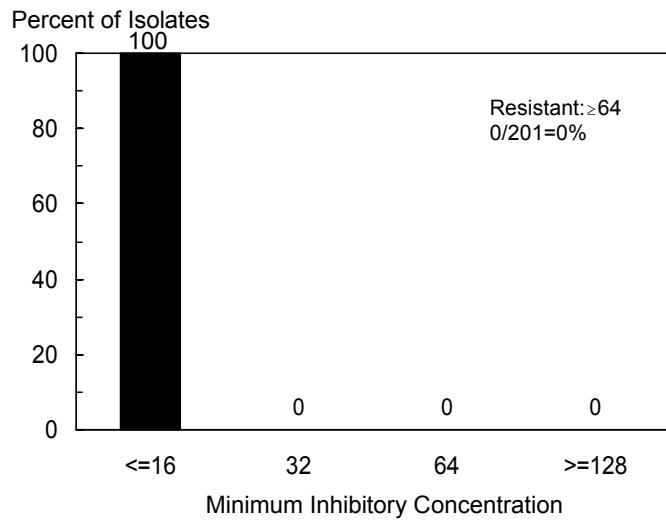
Figure 18k. MICs for gentamicin among *E. coli* O157 isolates, 1996-2001



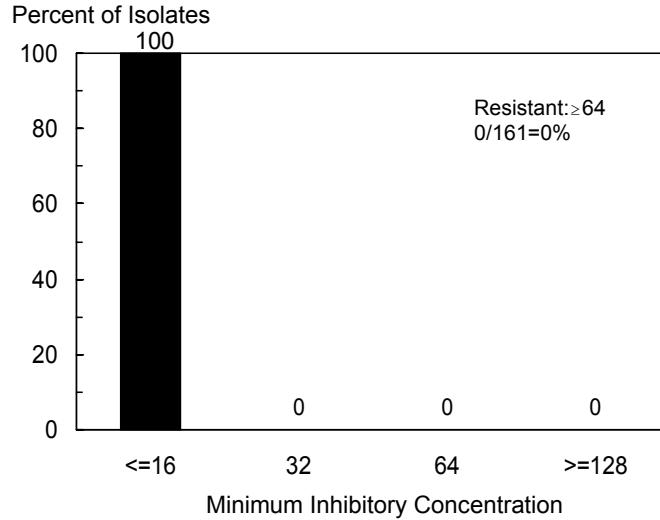
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Figure 18I. MICs for kanamycin among *E. coli* O157 isolates, 1996-2001

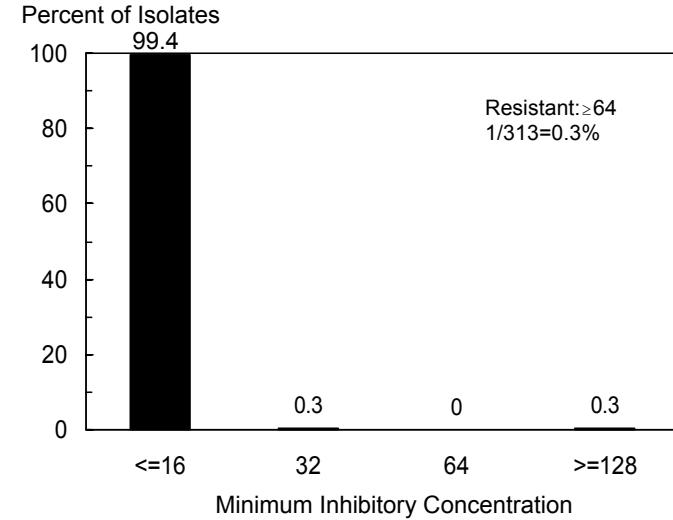
1996 (N=201)



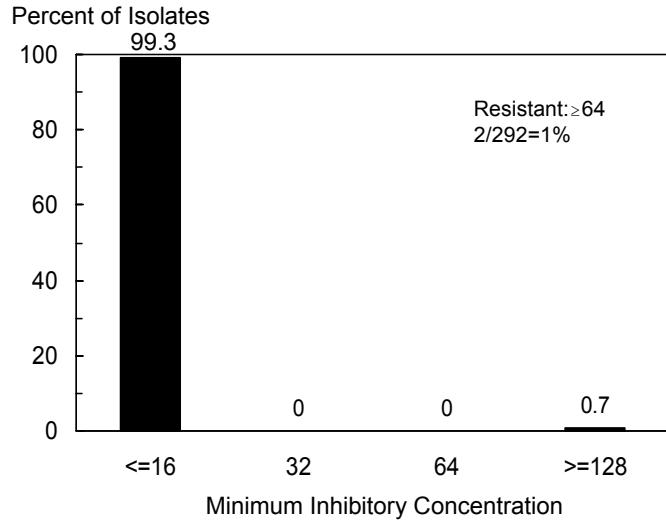
1997 (N=161)



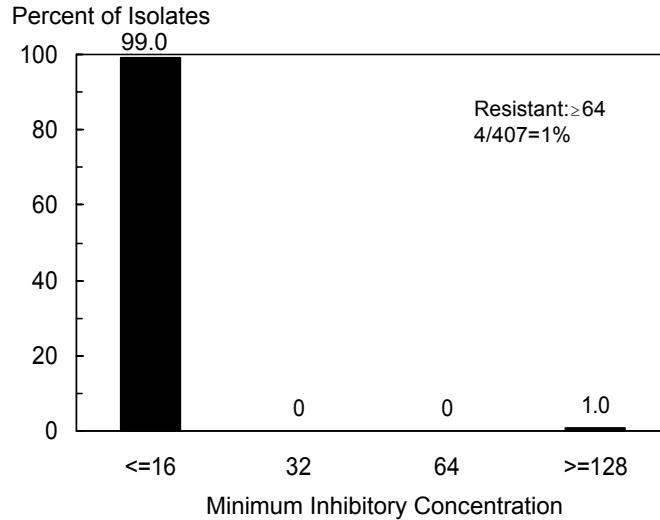
1998 (N=313)



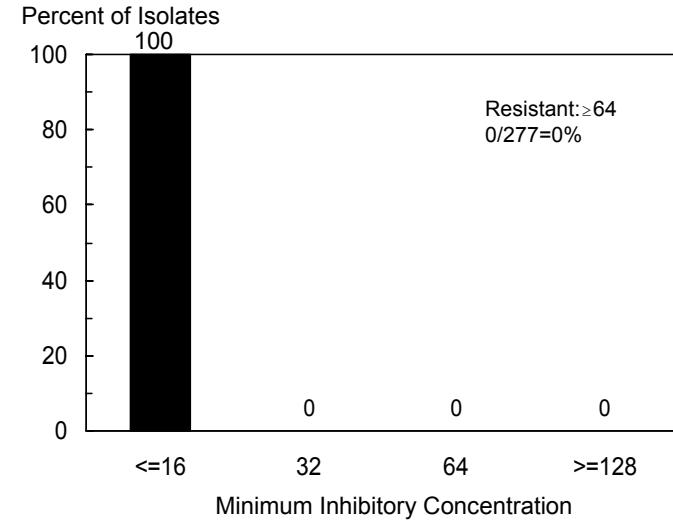
1999 (N=292)



2000 (N=407)

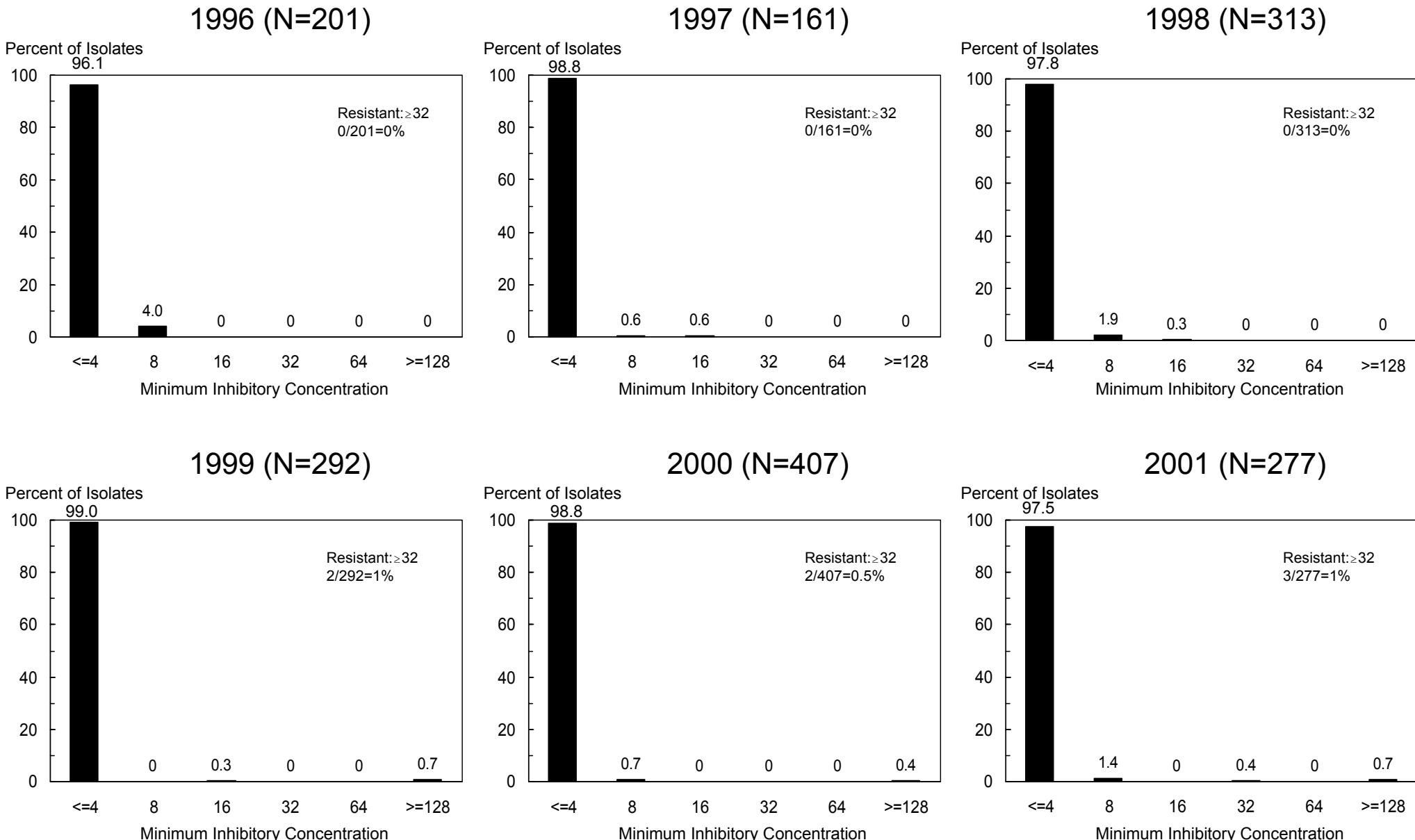


2001 (N=277)



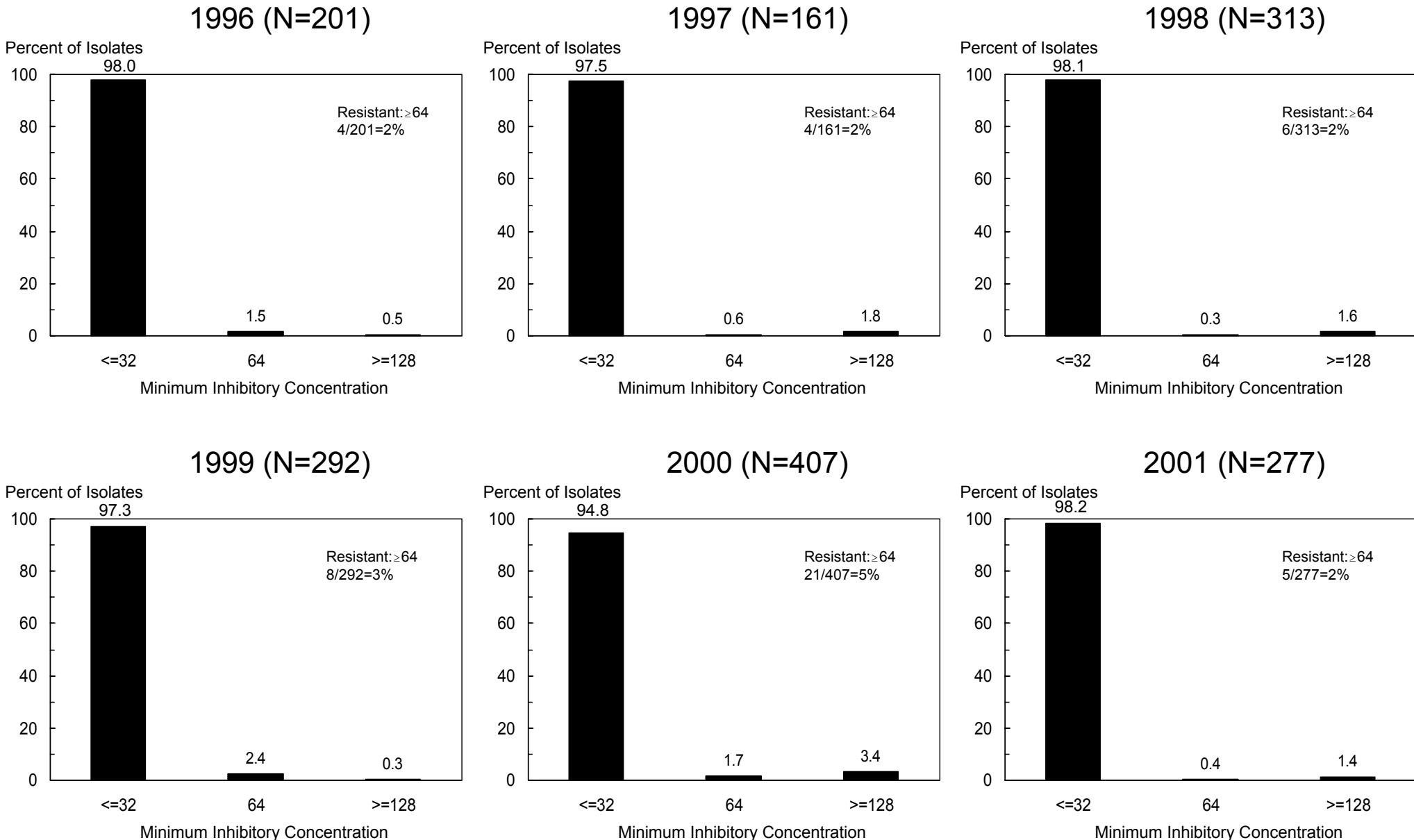
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Figure 18m. MICs for nalidixic acid among *E. coli* O157 isolates, 1996-2001



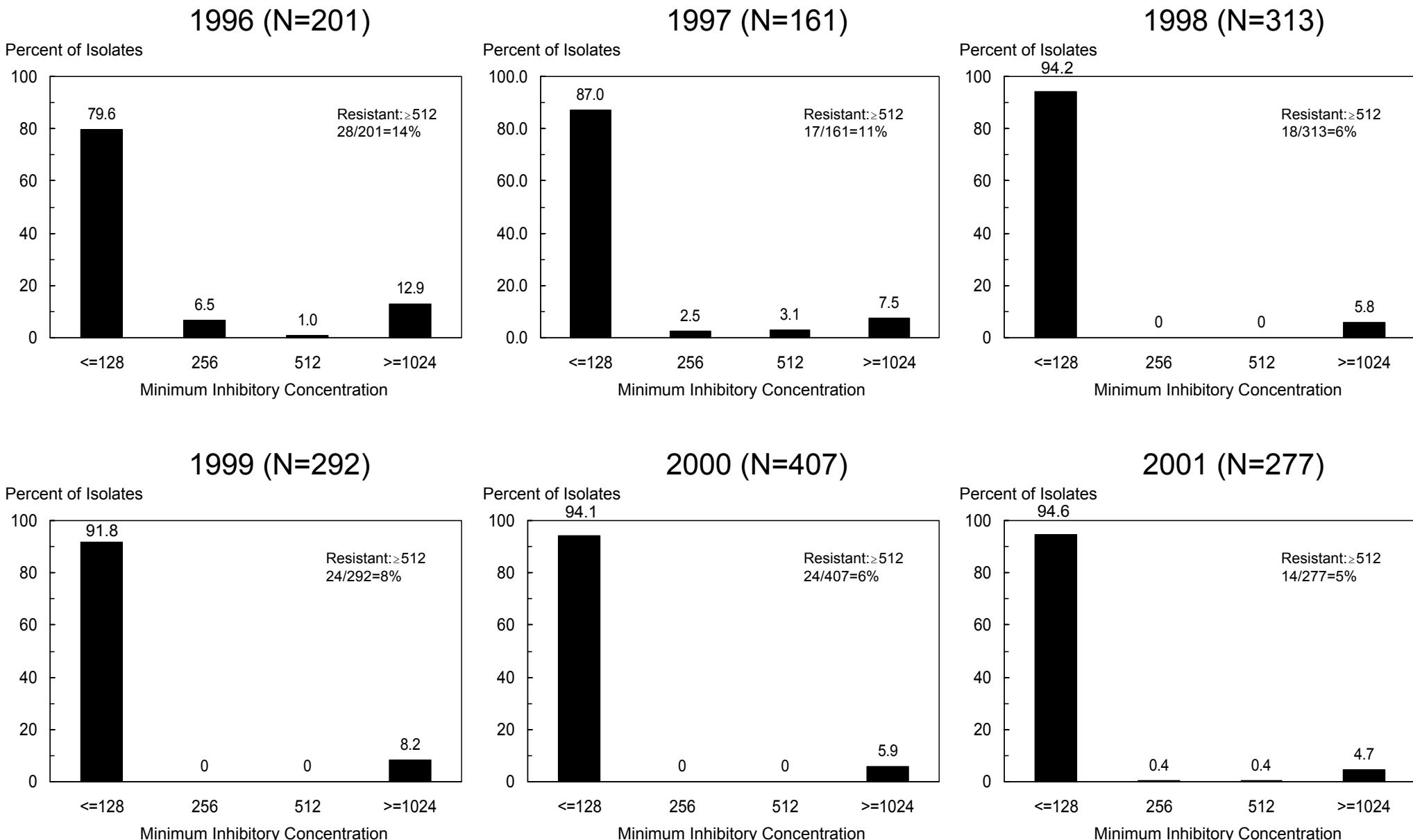
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Figure 18n. MICs for streptomycin among *E. coli* O157 isolates, 1996-2001



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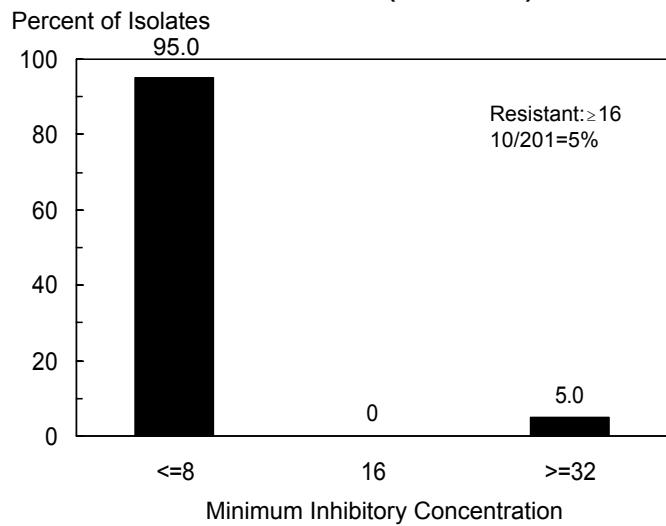
Figure 18o. MICs for sulfamethoxazole among *E. coli* O157 isolates, 1996-2001



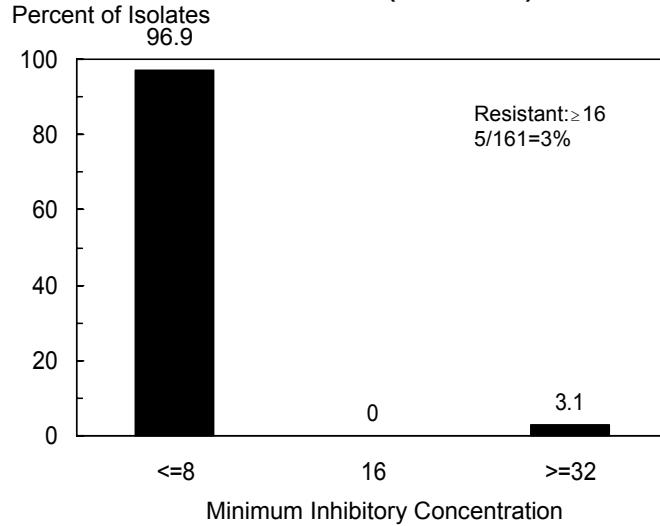
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Figure 18p. MICs for tetracycline among *E. coli* O157 isolates, 1996-2001

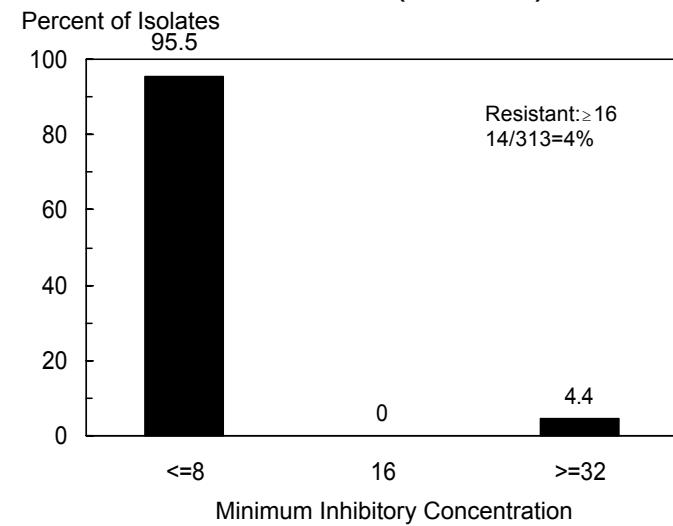
1996 (N=201)



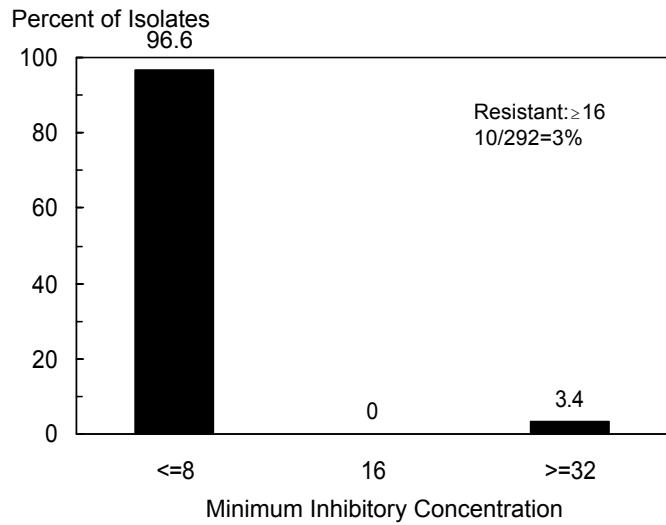
1997 (N=161)



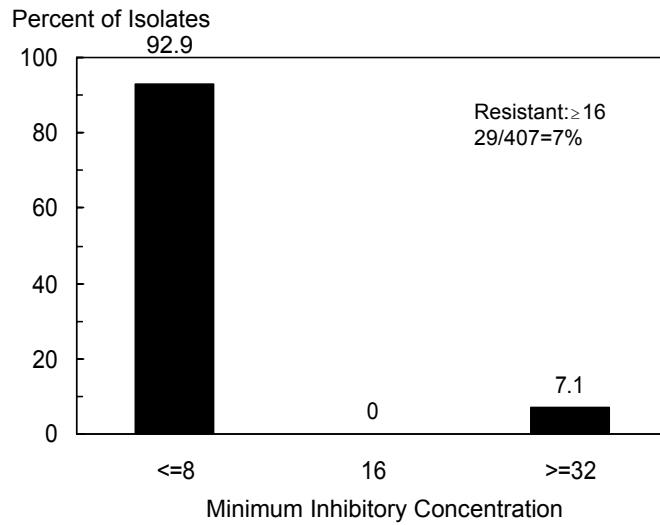
1998 (N=313)



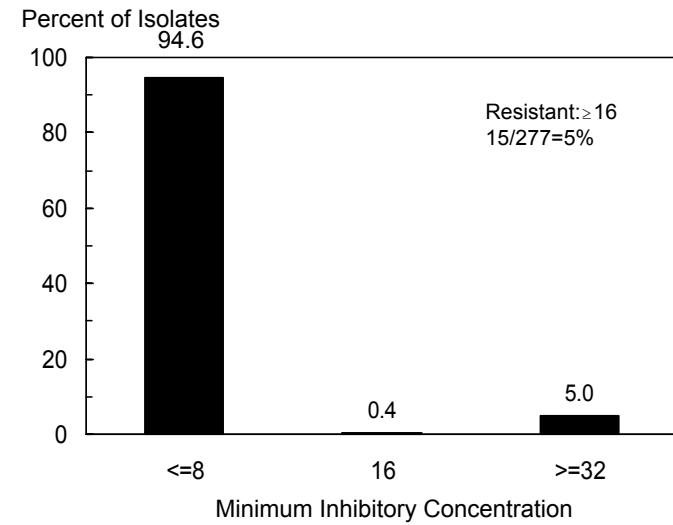
1999 (N=292)



2000 (N=407)



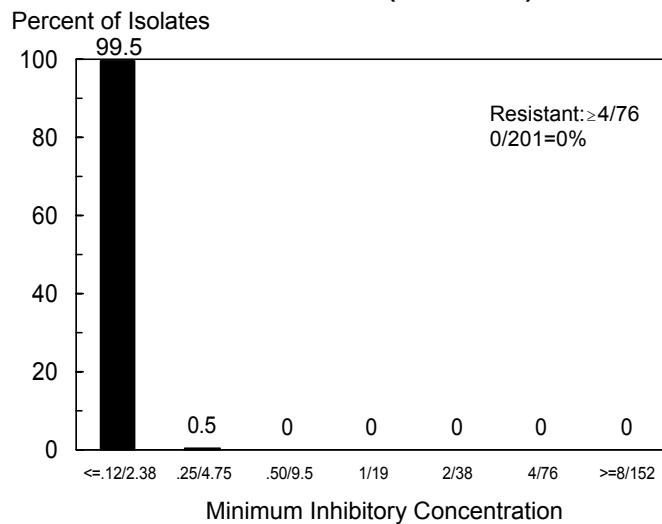
2001 (N=277)



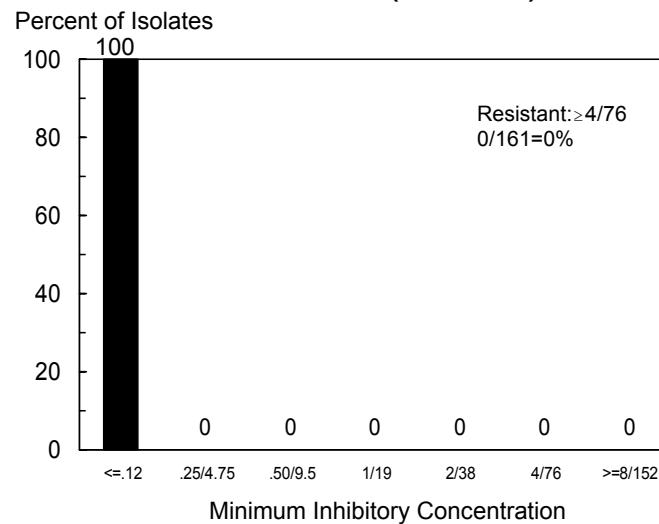
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Figure 18q. MICs for trimethoprim-sulfamethoxazole among *E. coli* O157 isolates, 1996-2001

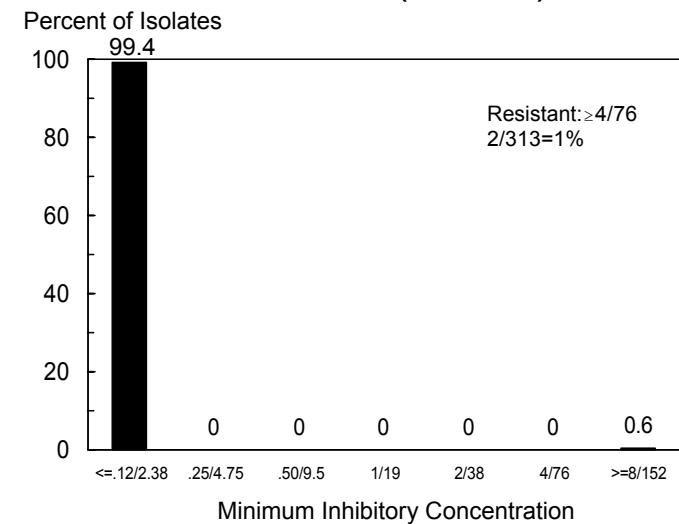
1996 (N=201)



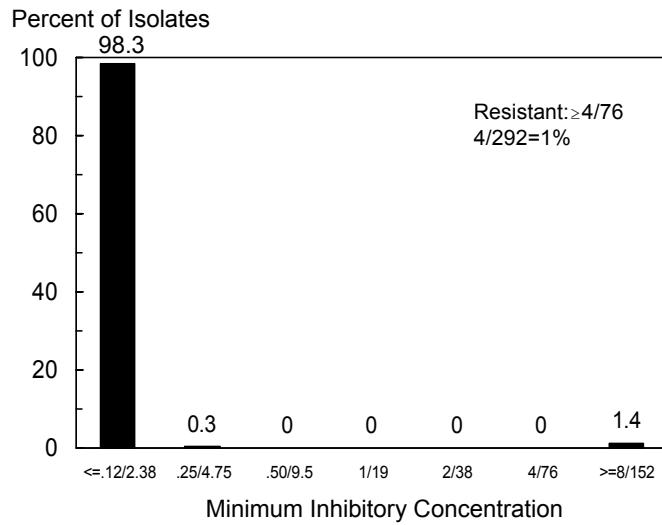
1997 (N=161)



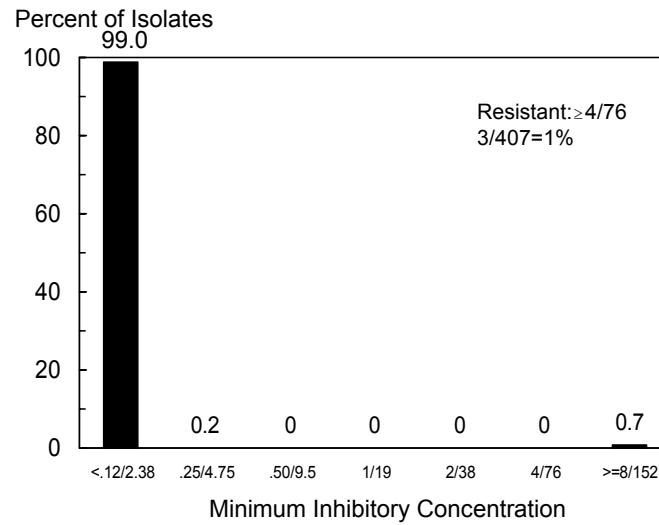
1998 (N=313)



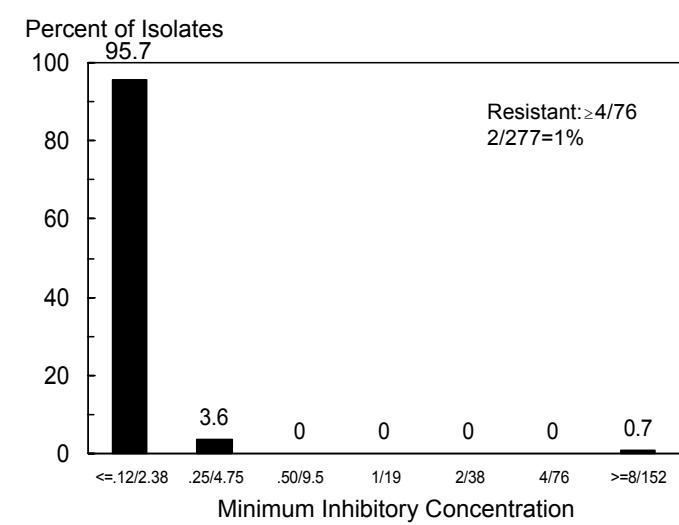
1999 (N=292)



2000 (N=407)

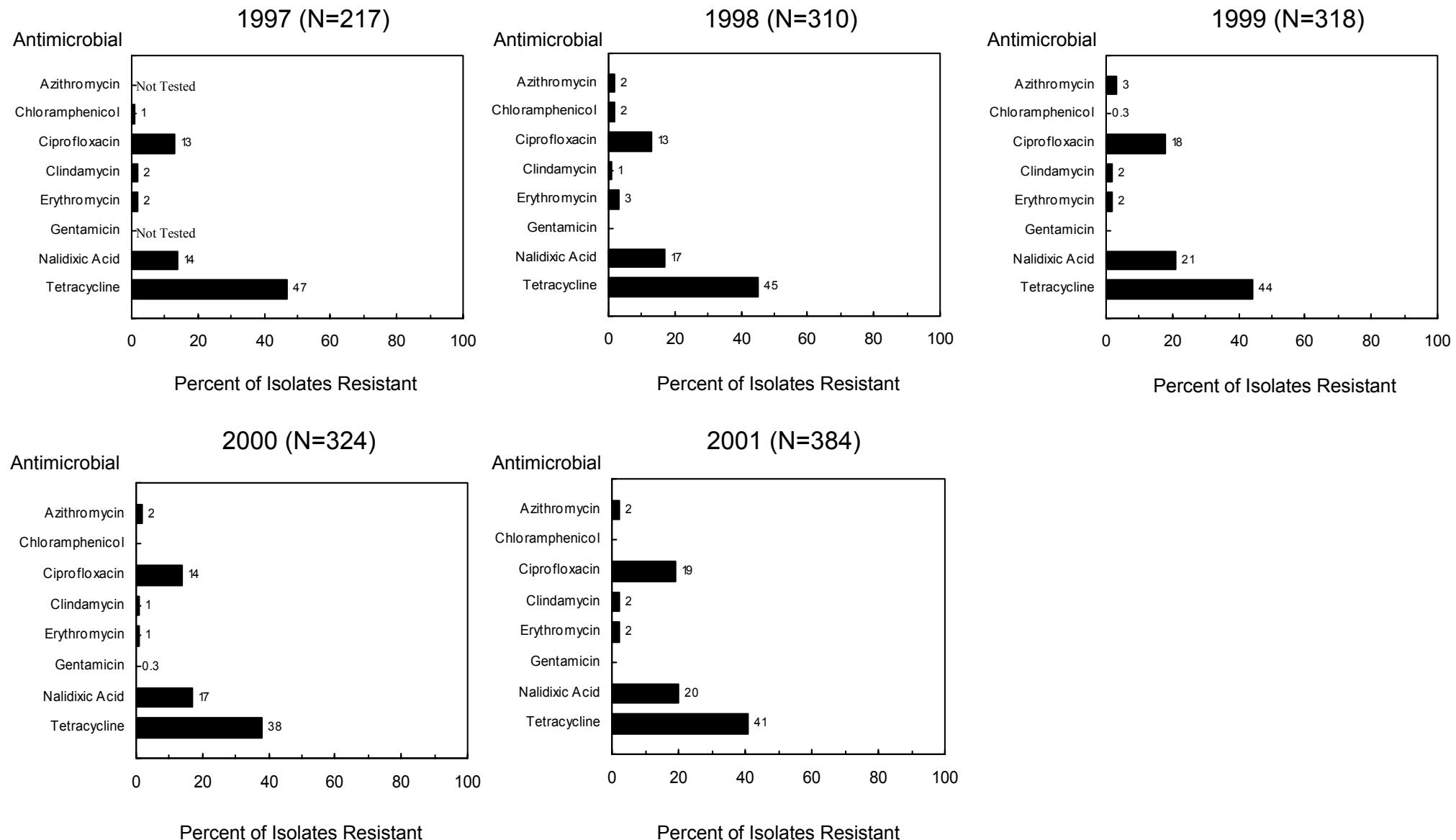


2001 (N=277)



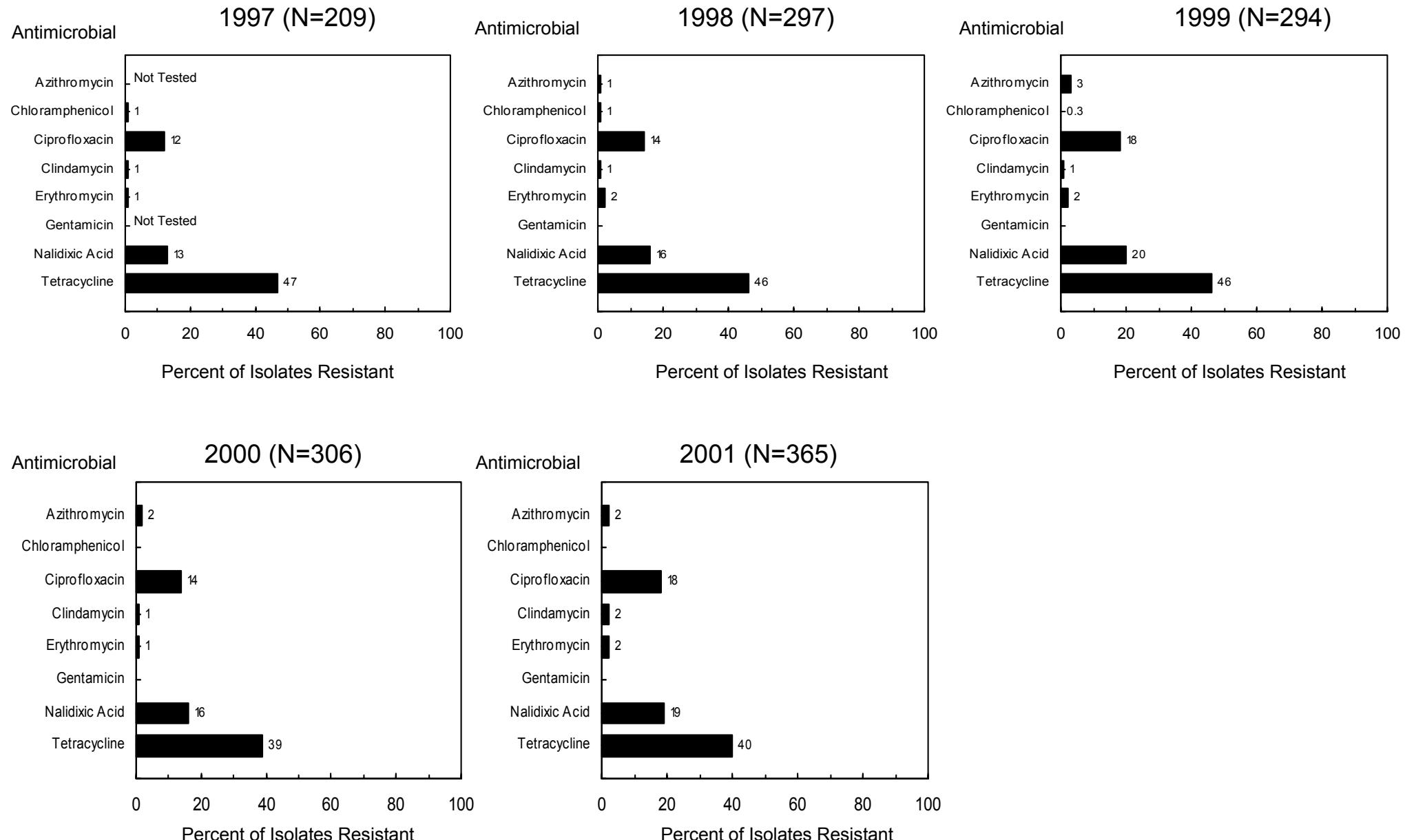
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Figure 19. Resistance among *Campylobacter* isolates, 1997-2001



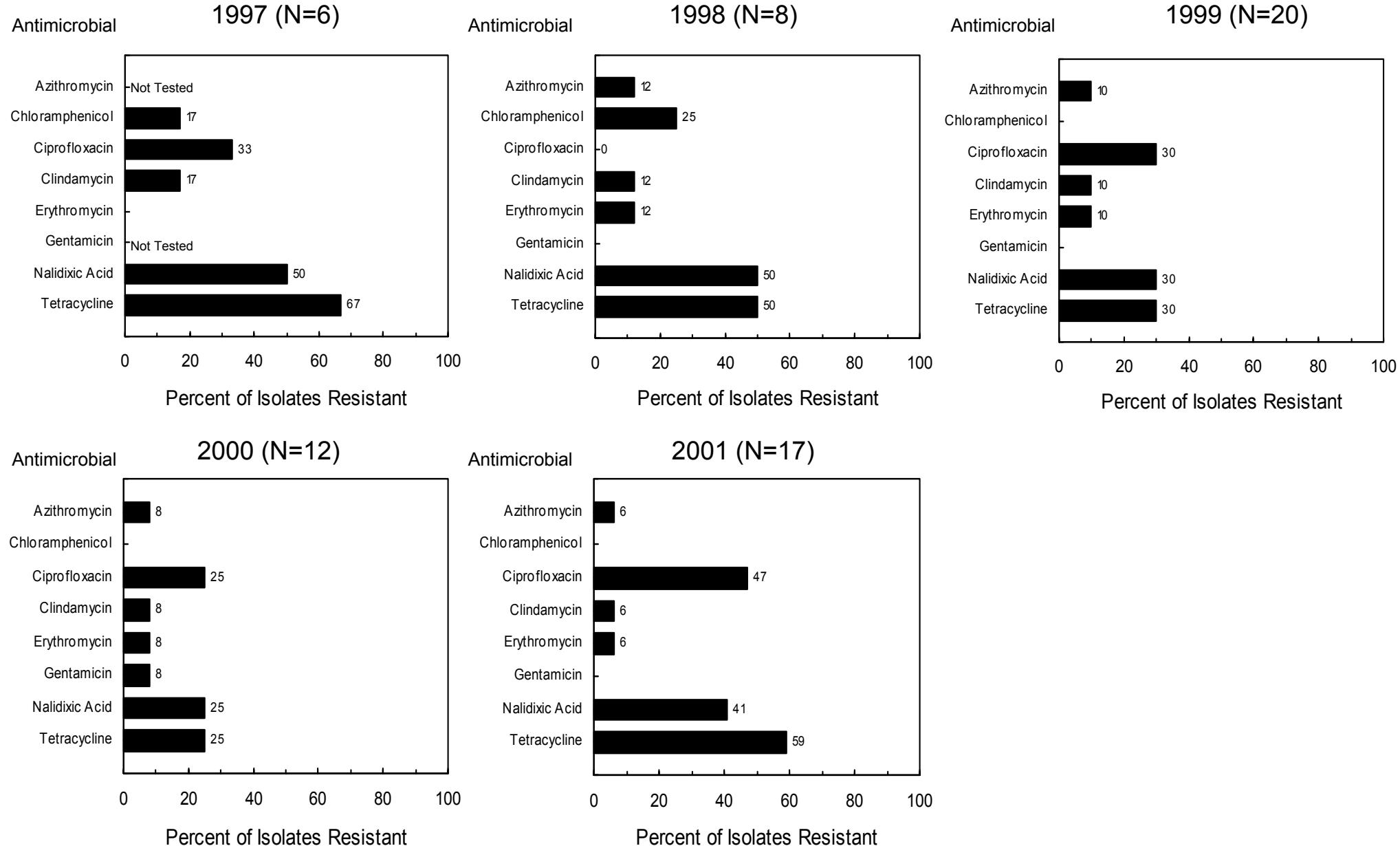
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Figure 20a. Resistance among *Campylobacter jejuni* isolates, 1997-2001



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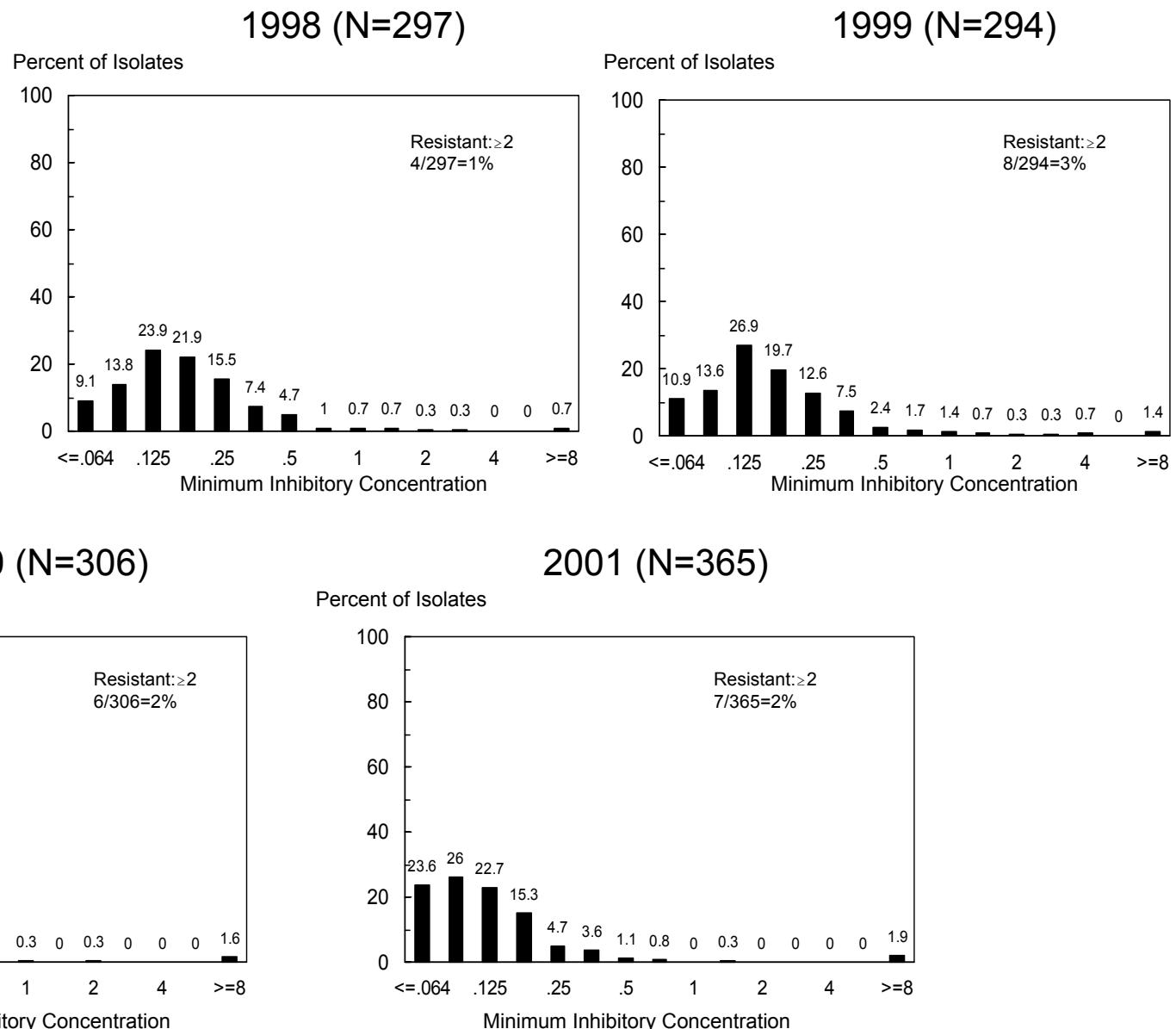
Figure 20b. Resistance among *Campylobacter coli* isolates, 1997-2001



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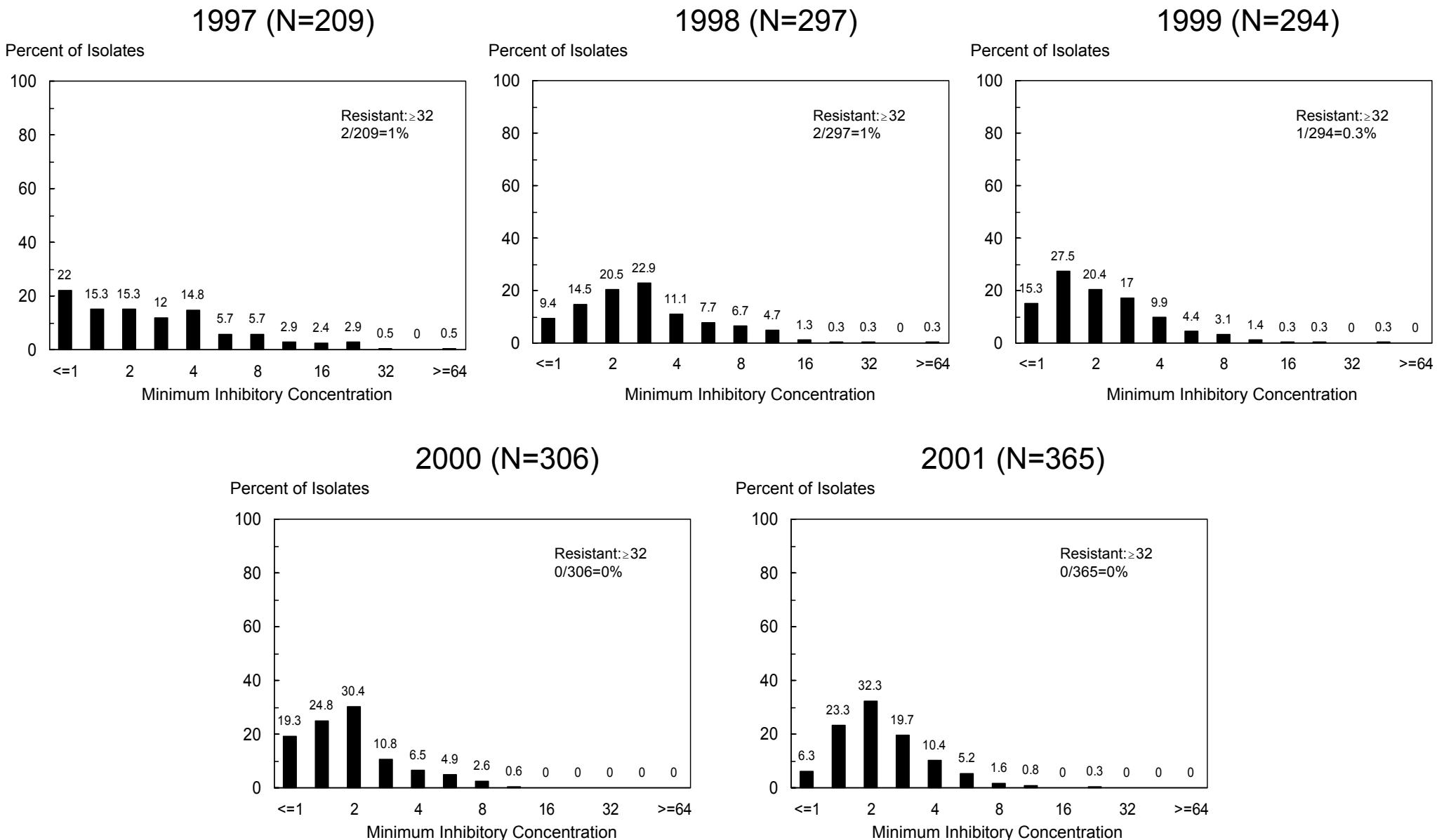
Figure 21a. MICs for azithromycin among *Campylobacter jejuni* isolates, 1997-2001

Not Tested in 1997



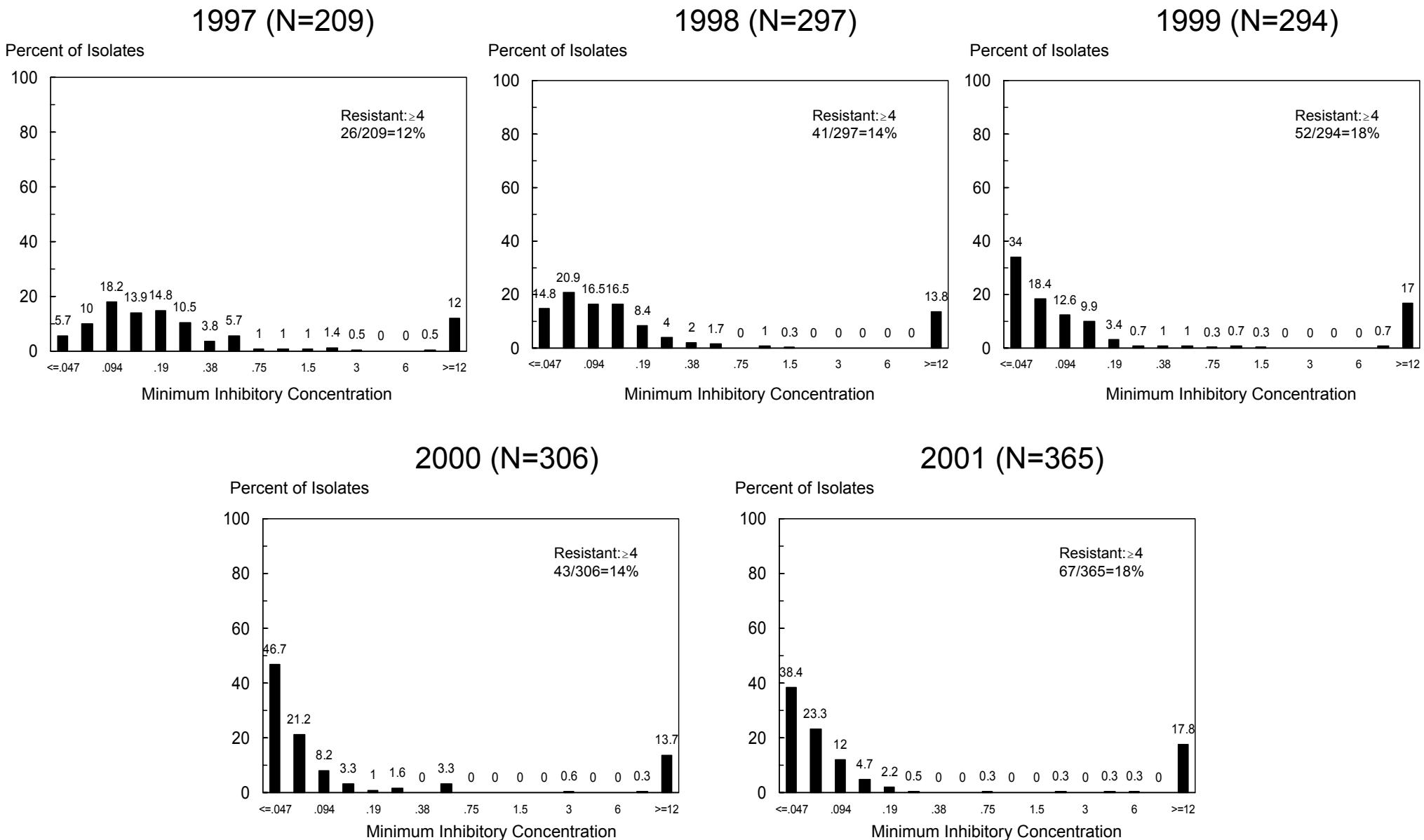
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Figure 21b. MICs for chloramphenicol among *Campylobacter jejuni* isolates, 1997-2001



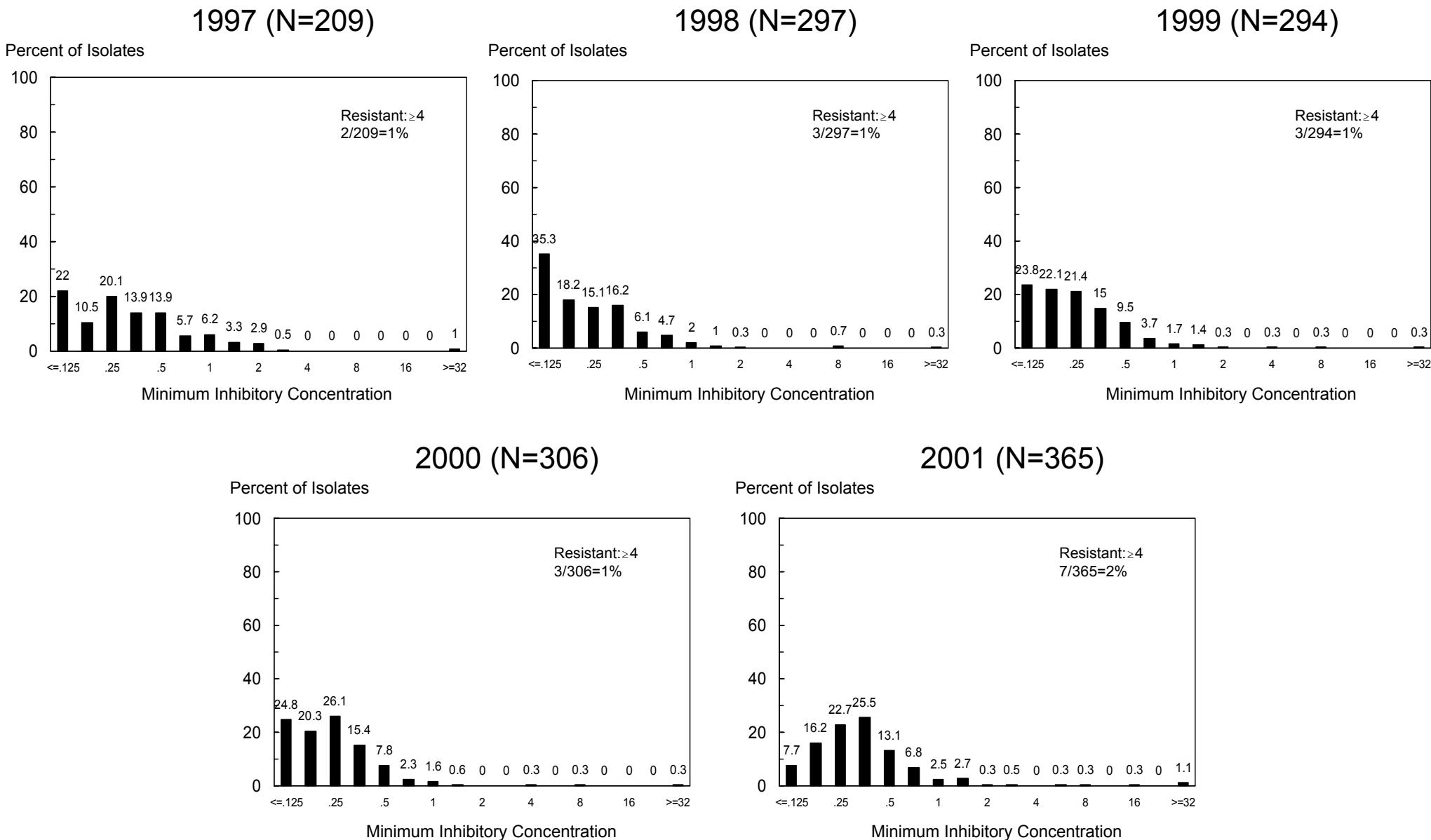
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Figure 21c. MICs for ciprofloxacin among *Campylobacter jejuni* isolates, 1997-2001



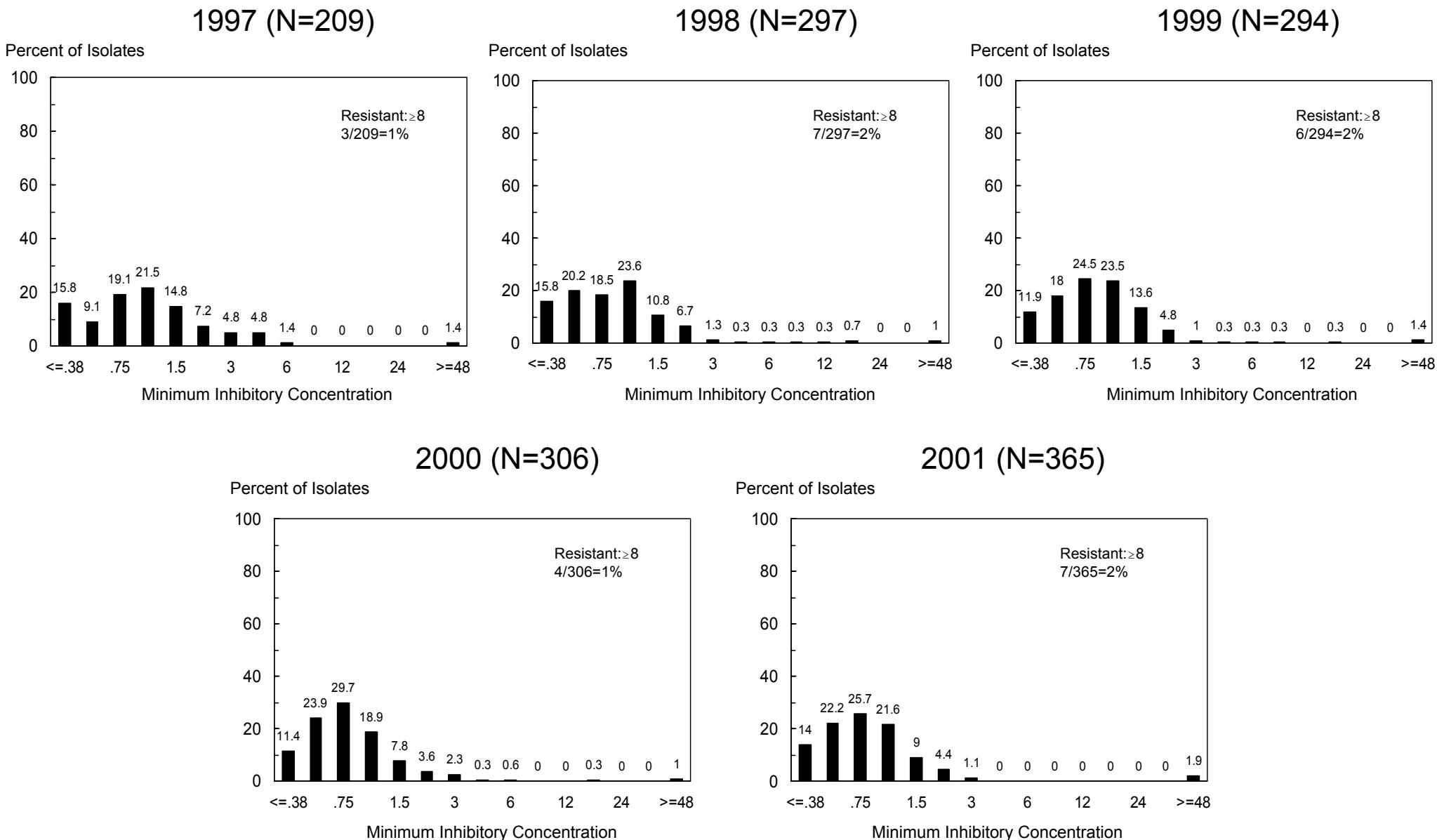
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Figure 21d. MICs for clindamycin among *Campylobacter jejuni* isolates, 1997-2001



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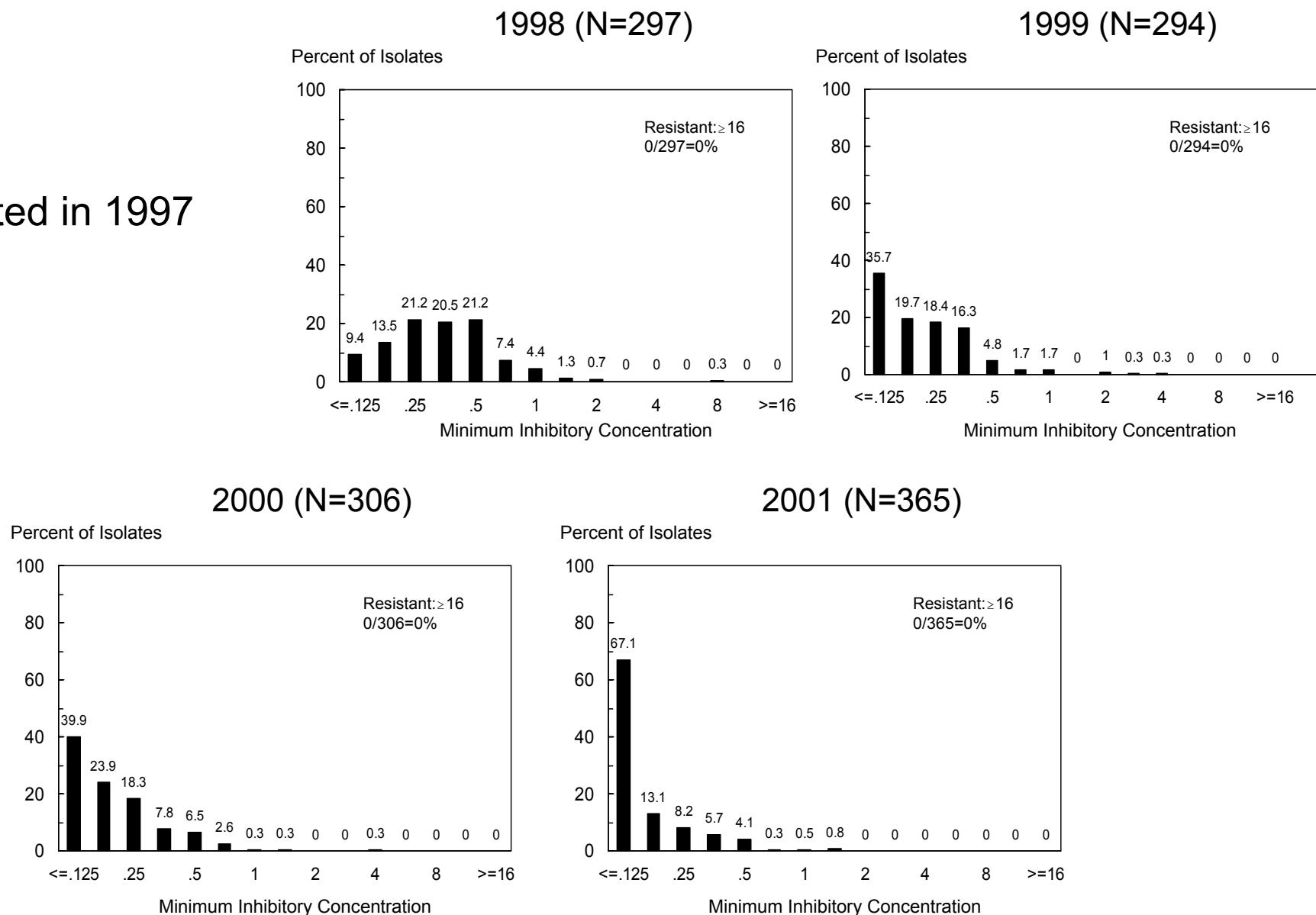
Figure 21e. MICs for erythromycin among *Campylobacter jejuni* isolates, 1997-2001



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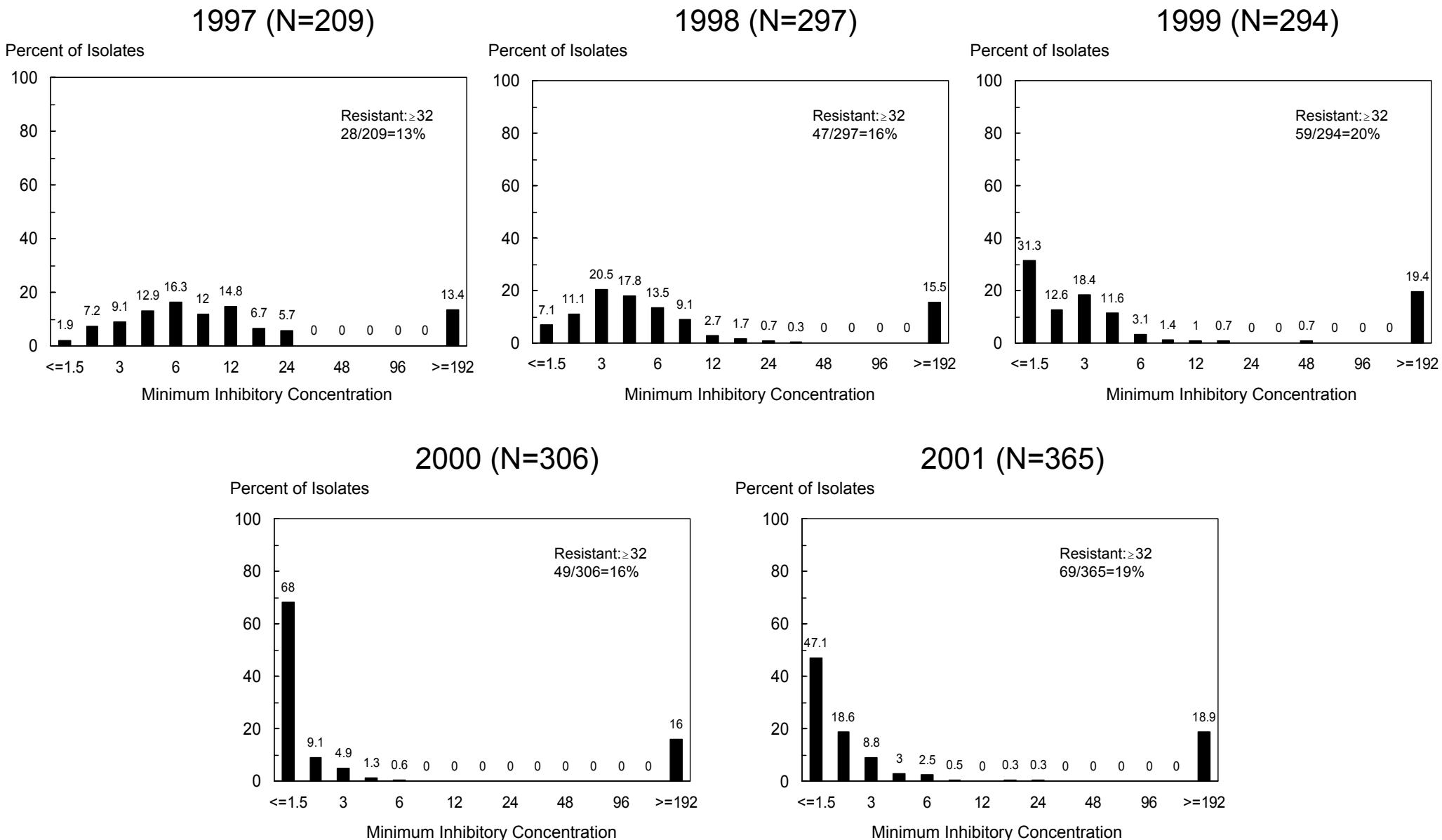
Figure 21f. MICs for gentamicin among *Campylobacter jejuni* isolates, 1997-2001

Not Tested in 1997



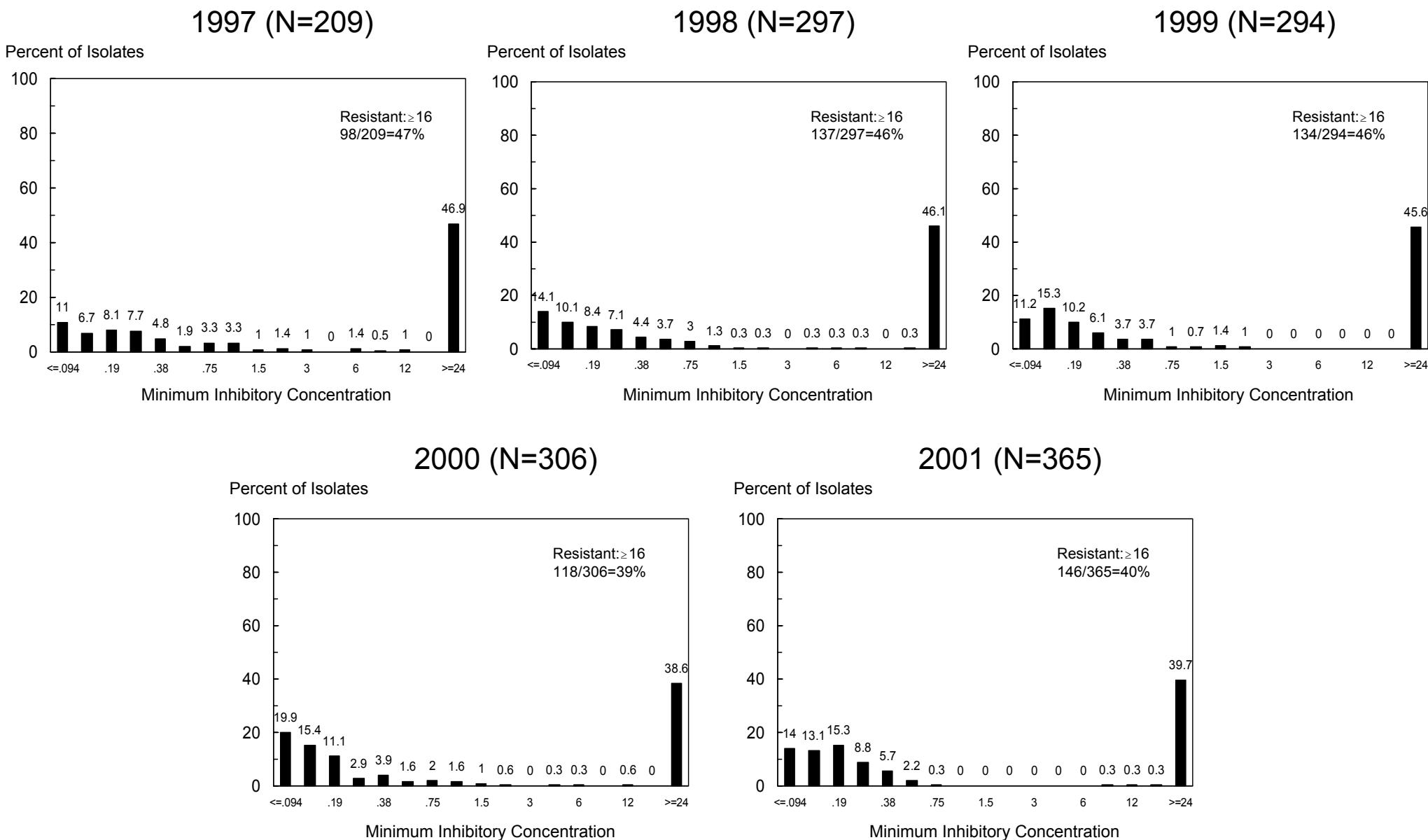
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Figure 21g. MICs for nalidixic acid among *Campylobacter jejuni* isolates, 1997-2001



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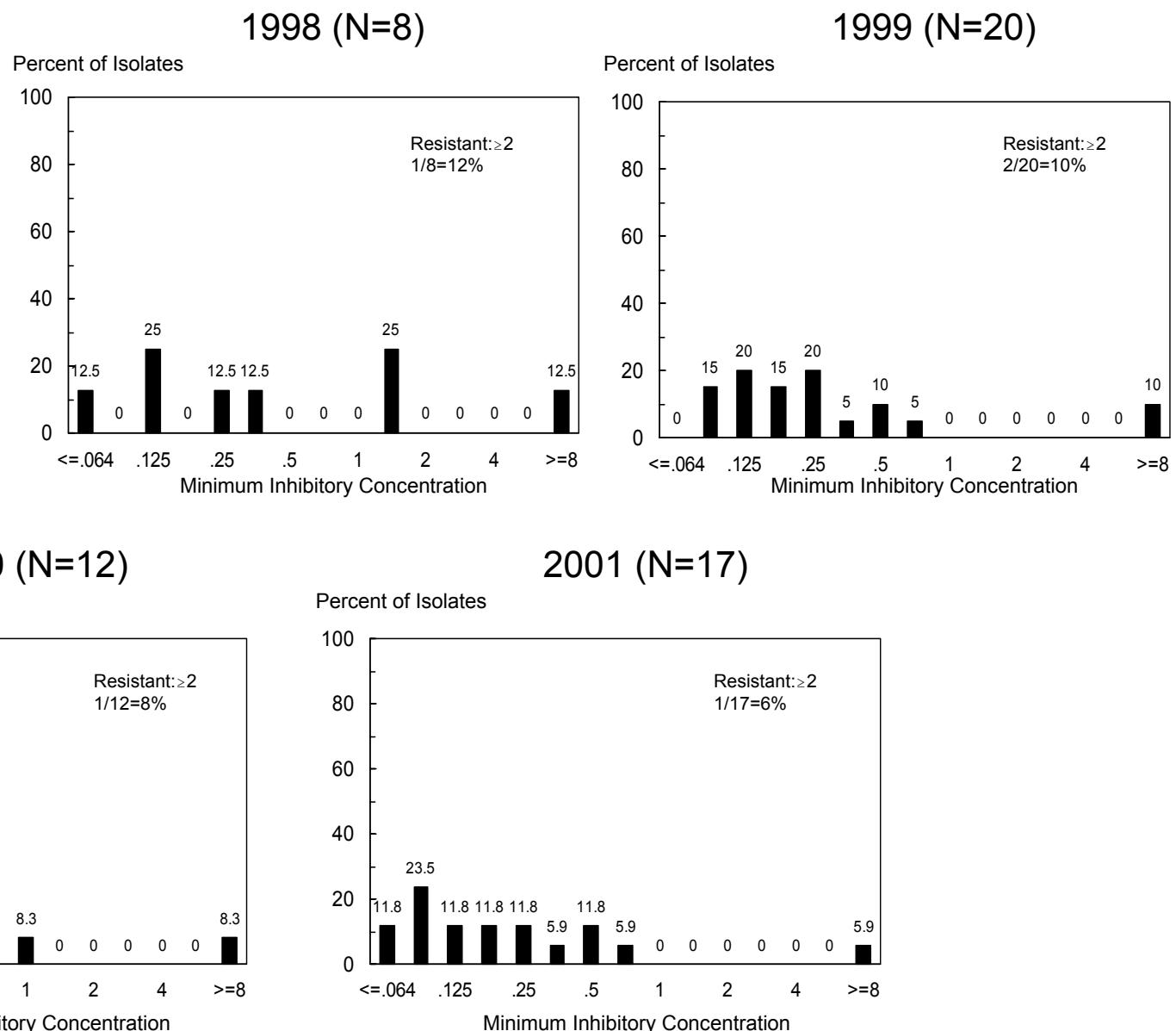
Figure 21h. MICs for tetracycline among *Campylobacter jejuni* isolates, 1997-2001



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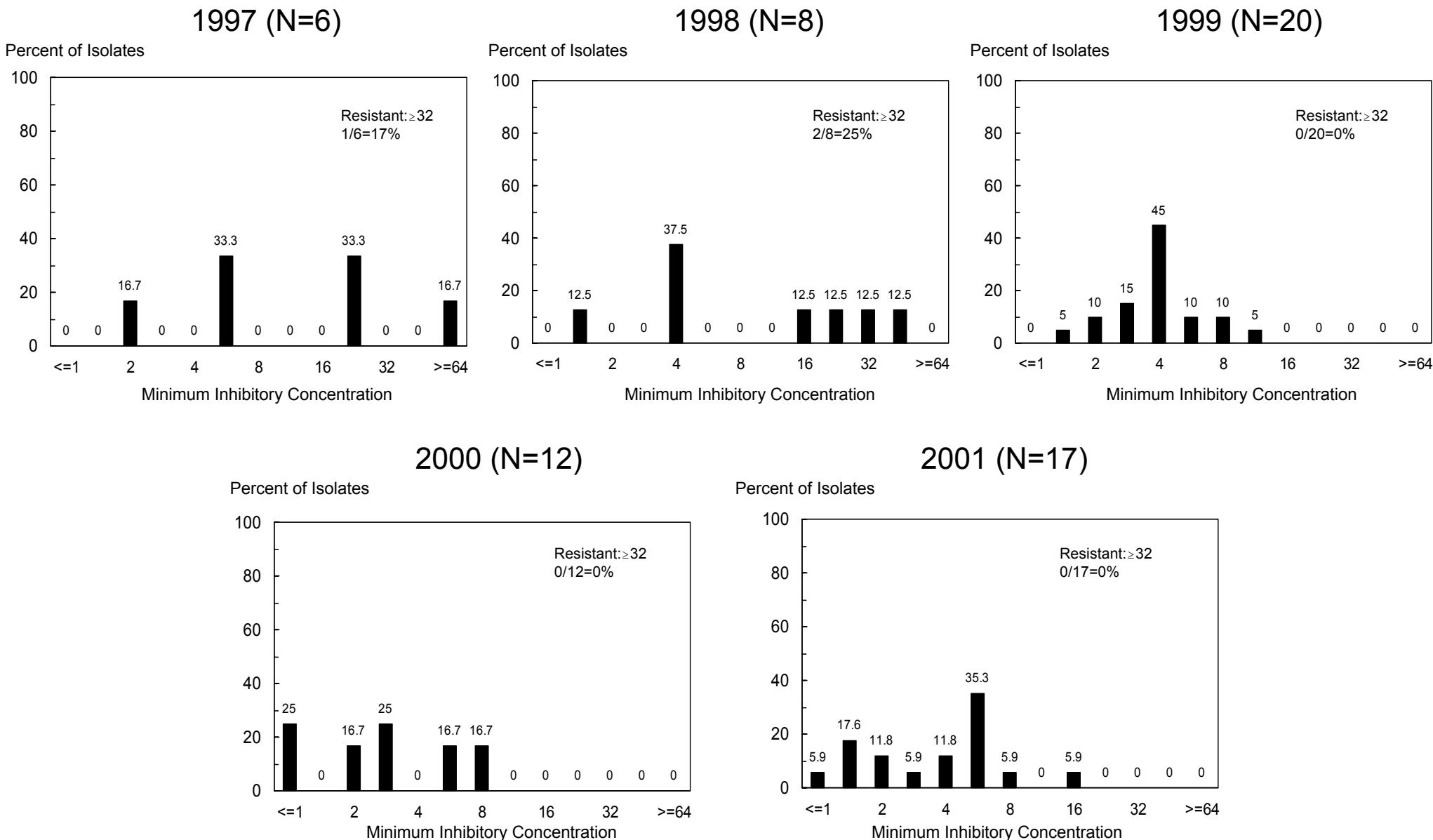
Figure 22a. MICs for azithromycin among *Campylobacter coli* isolates, 1997-2001

Not Tested in 1997



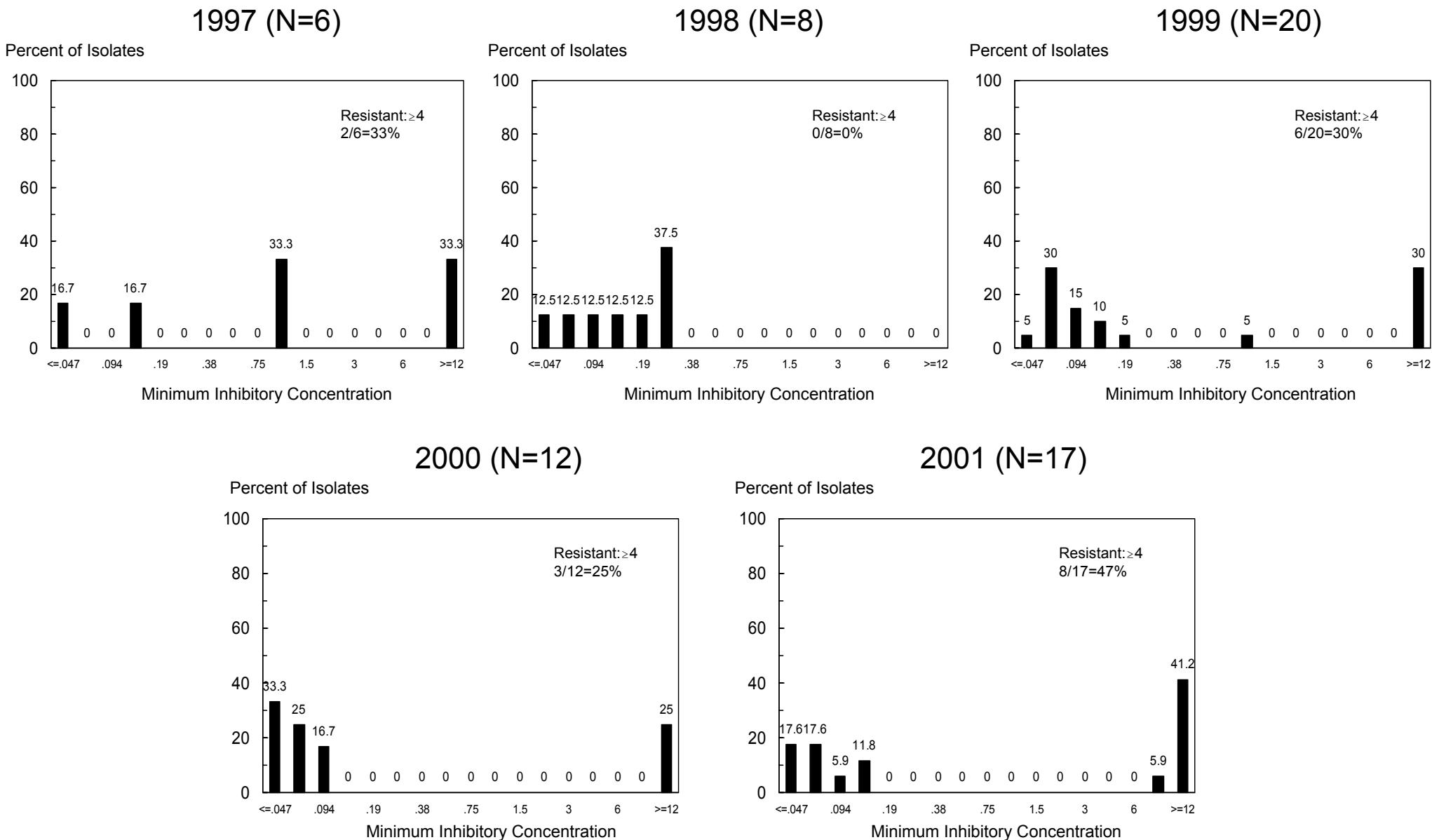
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Figure 22b. MICs for chloramphenicol among *Campylobacter coli* isolates, 1997-2001



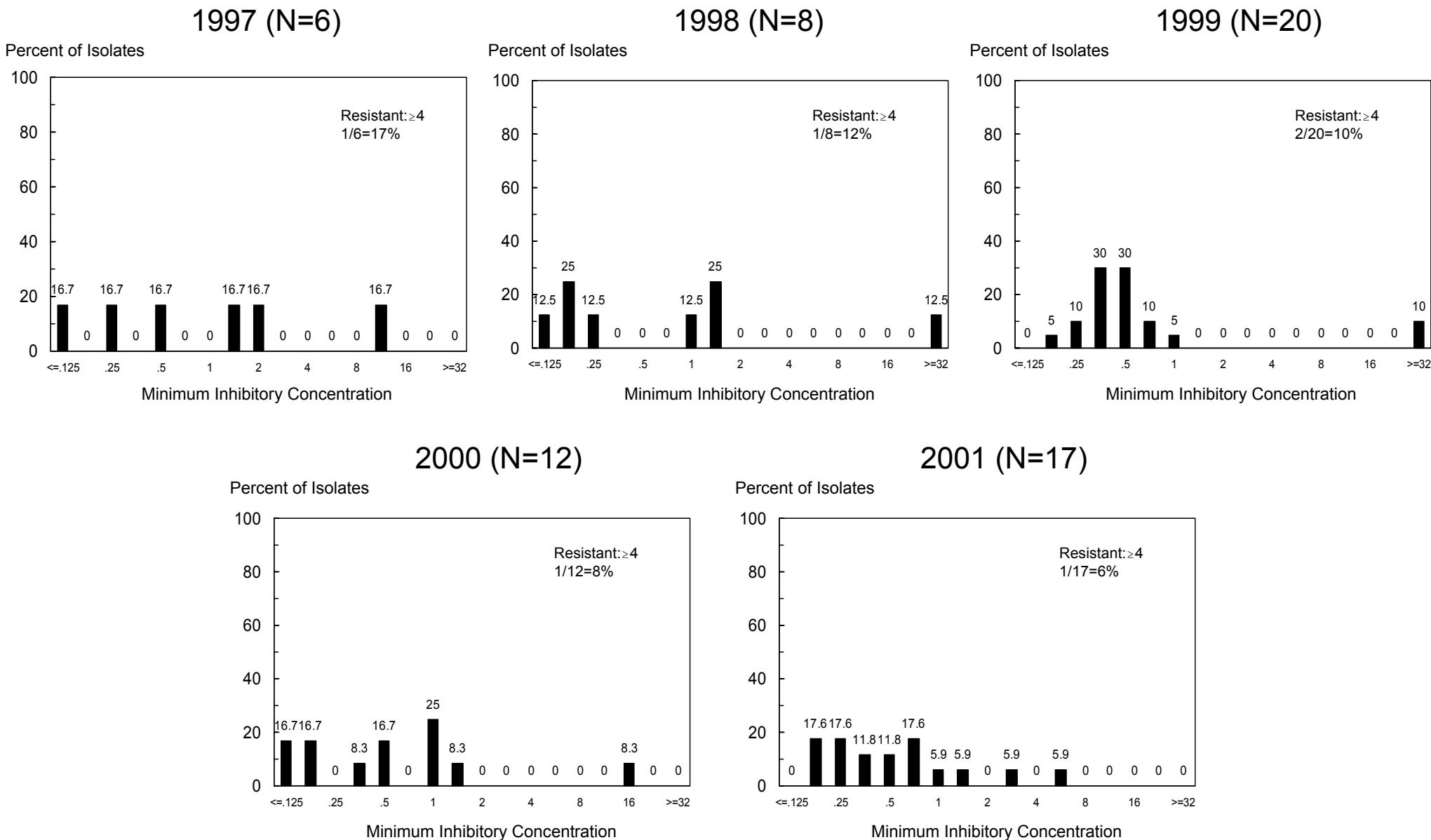
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Figure 22c. MICs for ciprofloxacin among *Campylobacter coli* isolates, 1997-2001



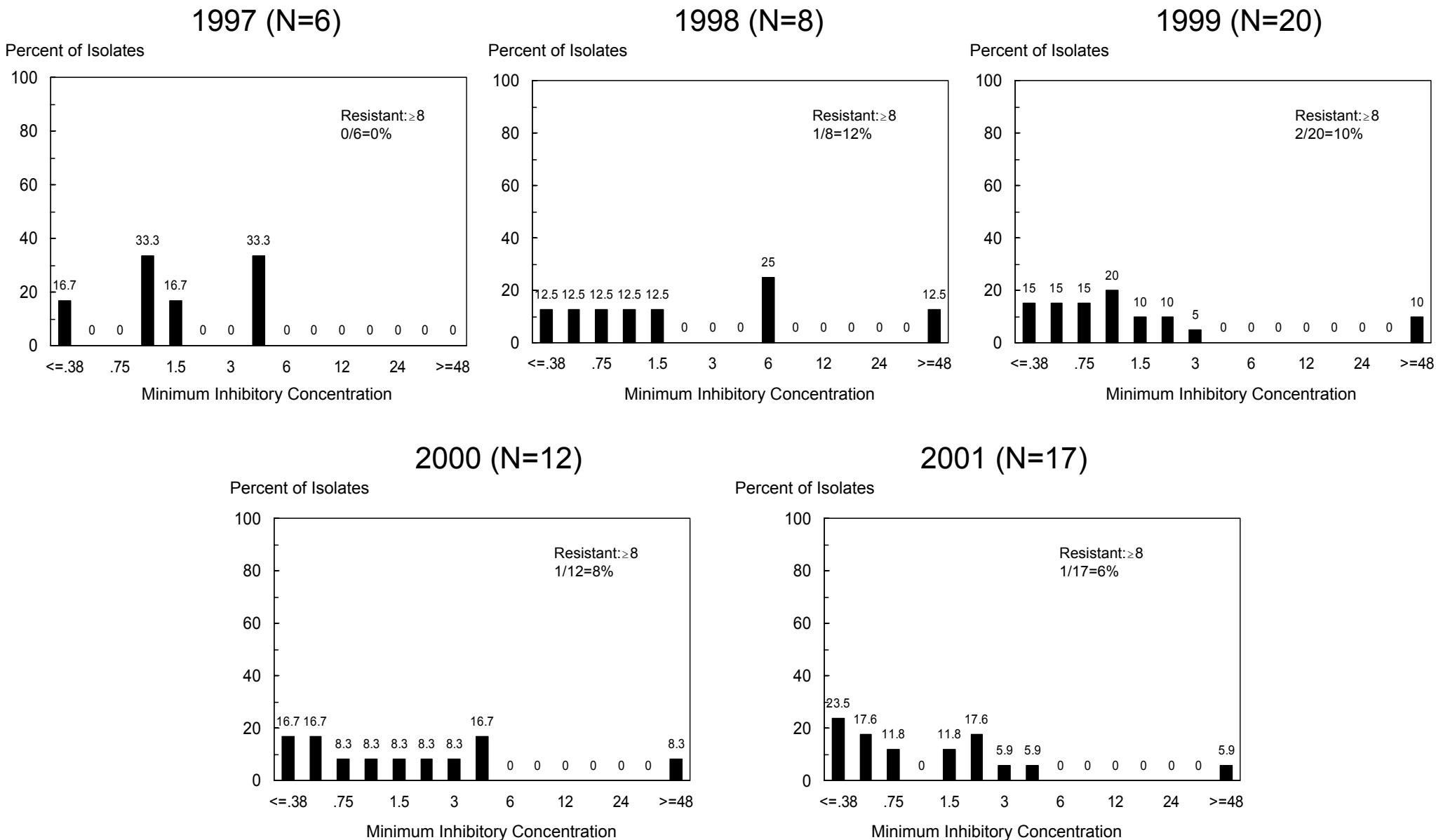
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Figure 22d. MICs for clindamycin among *Campylobacter coli* isolates, 1997-2001



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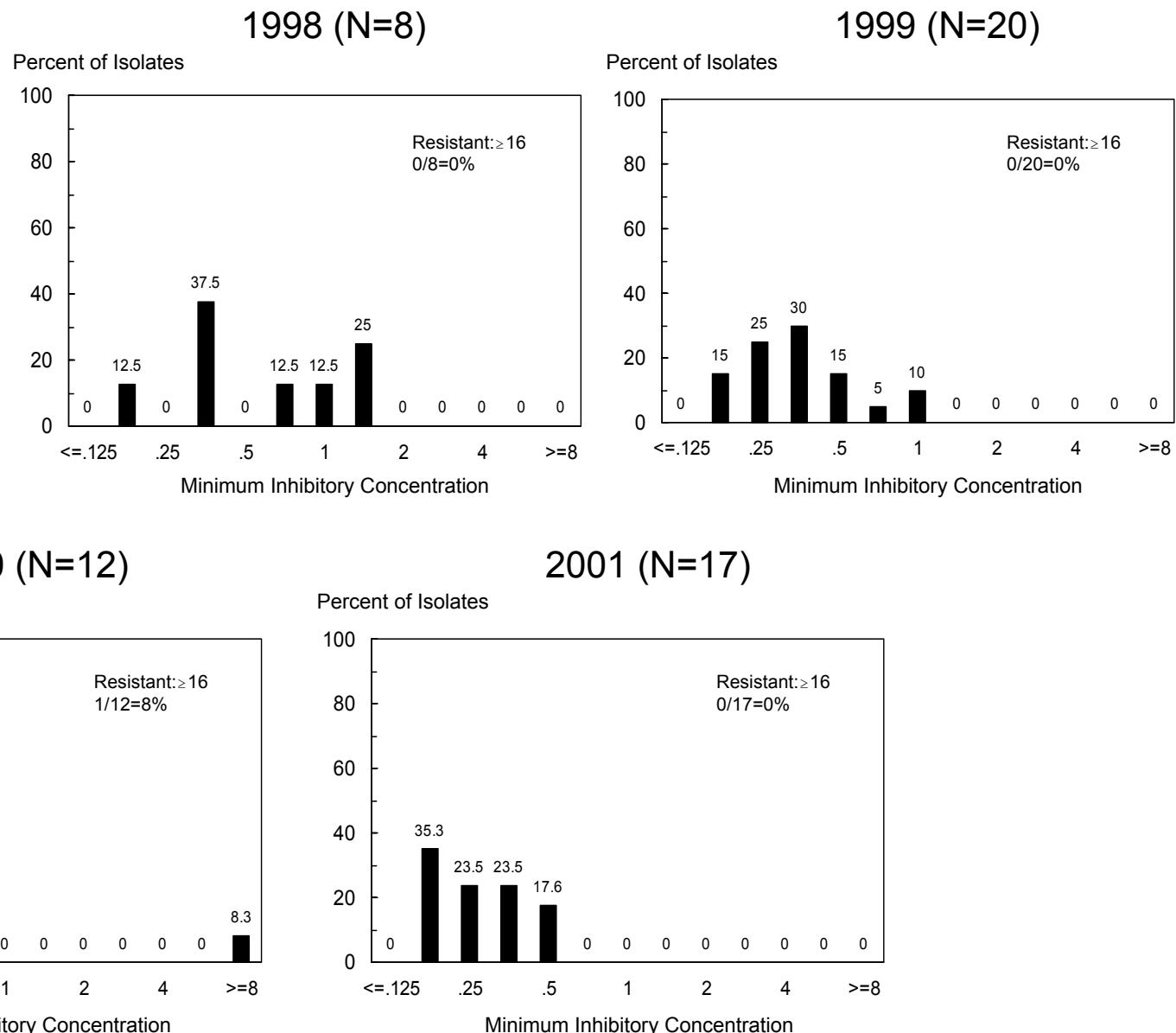
Figure 22e. MICs for erythromycin among *Campylobacter coli* isolates, 1997-2001



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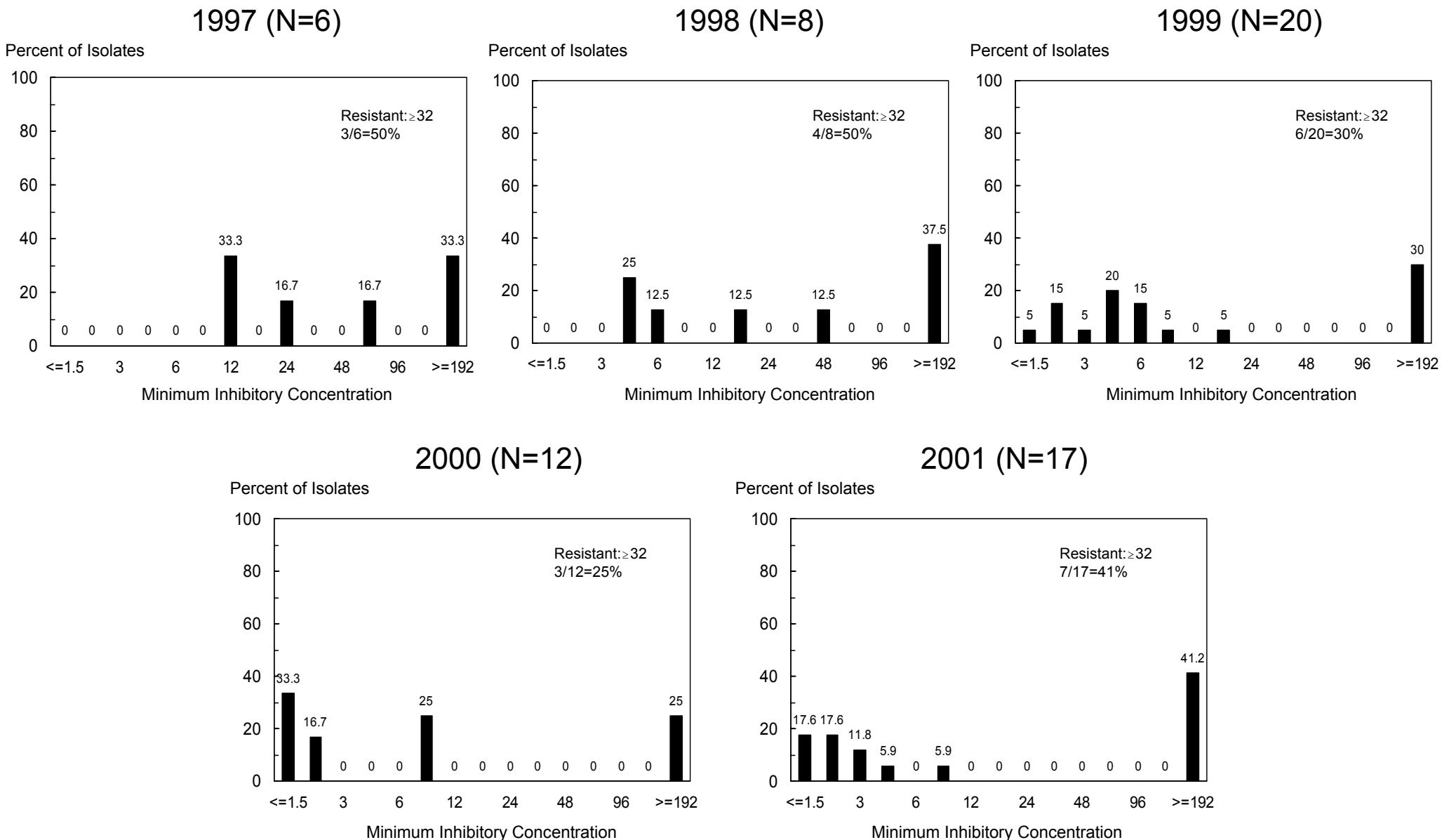
Figure 22f. MICs for gentamicin among *Campylobacter coli* isolates, 1997-2001

Not Tested in 1997



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Figure 22g. MICs for nalidixic acid among *Campylobacter coli* isolates, 1997-2001



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Figure 22h. MICs for tetracycline among *Campylobacter coli* isolates, 1997-2001

