

Shiga Toxin–Associated Hemolytic Uremic Syndrome in Adults, France, 2009–2017

Benoît Travert,¹ Antoine Dossier,¹ Matthieu Jamme, Aurélie Cointe, Yahsou Delmas, Sandrine Malot, Alain Wynckel, Amélie Seguin, Claire Presne, Miguel Hie, Ygal Benhamou, David Ribes, Gabriel Choukroun, Steven Grangé, Alexandre Hertig, Emilie Cornec-Le Gall, Lionel Galicier, Eric Daugas, Lila Bouadma, François-Xavier Weill, Elie Azoulay, Fadi Fakhouri, Agnès Veyradier, Stéphane Bonacorsi, Julien Hogan, Véronique Frémeaux-Bacchi, Eric Rondeau, Patricia Mariani-Kurkdjian, Paul Coppo, Centre de Référence des Microangiopathies Thrombotiques²

We conducted a retrospective study on hemolytic uremic syndrome caused by Shiga toxin–producing *Escherichia coli* (STEC) in 96 adults enrolled in the cohort of the National Reference Center for Thrombotic Microangiopathies network in France during 2009–2017. Most infections were caused by STEC strains not belonging to the O157 or O104 serogroups. Thirty (31.3%) patients had multiple risk factors for thrombotic microangiopathy. In total, 61 (63.5%) patients required dialysis, 50 (52.1%) had a serious neurologic complication, 34 (35.4%) required mechanical ventilation, and 19 (19.8%) died during hospitalization. We used multivariate analysis to determine that the greatest risk factors for death were underlying immunodeficiency (hazard ratio 3.54) and severe neurologic events (hazard ratio 3.40). According to multivariate analysis and propensity score-matching, eculizumab treatment was not associated with survival. We found that underlying conditions, especially immunodeficiency, are strongly associated with decreased survival in adults who have hemolytic uremic syndrome caused by STEC.

Shiga toxin–producing *Escherichia coli* (STEC) infection is an environmental foodborne or waterborne disease that causes bloody diarrhea. Approximately 5%–20% of cases are complicated by hemolytic uremic syndrome (HUS) (1,2). Shiga toxins (Stx) can cause acute microvascular injury, leading to thrombotic microangiopathy (TMA), which is characterized by hemolytic anemia and thrombocytopenia, and in the scenario of HUS, associated with acute kidney injury (3). Researchers estimate that the global prevalence of STEC infection is ≈ 43.1 acute illnesses/100,000 person-years, causing $\approx 3,890$ annual cases of STEC-associated HUS (4). STEC-associated HUS occurs mostly in children; sporadic cases are rare in adults.

Among children, STEC-associated HUS is the most frequent form of TMA and the leading cause of acute renal failure (3). In France, surveillance for STEC-associated HUS in children <15 years of age

Author affiliations: Centre de Référence des Microangiopathies Thrombotiques, Paris, France (B. Travert, A. Dossier, M. Jamme, Y. Delmas, S. Malot, A. Wynckel, A. Seguin, C. Presne, M. Hie, Y. Benhamou, G. Choukroun, S. Grangé, A. Hertig, L. Galicier, E. Azoulay, F. Fakhouri, A. Veyradier, V. Frémeaux-Bacchi, E. Rondeau, P. Coppo); Université de Paris, Paris (B. Travert, A. Dossier, A. Cointe, L. Galicier, E. Daugas, L. Bouadma, E. Azoulay, A. Veyradier, S. Bonacorsi, J. Hogan, V. Frémeaux-Bacchi, P. Mariani-Kurkdjian); Hôpital Bichat—Claude Bernard, Paris (B. Travert, A. Dossier, E. Daugas, L. Bouadma); Sorbonne-Université, Paris (M. Jamme, M. Hie, A. Hertig, E. Rondeau, P. Coppo); Hôpital Tenon, Paris (M. Jamme, E. Rondeau); Hôpital Robert-Debré, Paris (A. Cointe, S. Bonacorsi, J. Hogan, P. Mariani-Kurkdjian); Centre Hospitalier Universitaire de Bordeaux, Bordeaux, France (Y. Delmas); Hôpital Maison Blanche, Reims, France (A. Wynckel); Centre Hospitalier Universitaire de Caen, Caen, France (A. Seguin); Centre

Hospitalier Universitaire d'Amiens, Amiens, France (C. Presne, G. Choukroun); Groupement Hospitalier Pitié-Salpêtrière, Paris (M. Hie, A. Hertig); Centre Hospitalier Universitaire de Rouen, Rouen, France (Y. Benhamou, S. Grangé); Centre Hospitalier Universitaire de Toulouse, Toulouse, France (D. Ribes); Centre Hospitalier Universitaire de Brest, Brest, France (E. Cornec-Le Gall); Hôpital Saint-Louis, Paris (L. Galicier, E. Azoulay); Institut Pasteur, Paris (F.-X. Weill); Centre Hospitalier Universitaire de Nantes, Nantes, France (F. Fakhouri); Hôpital Lariboisière, Paris (A. Veyradier); Hôpital Européen Georges Pompidou, Paris (V. Frémeaux-Bacchi); Hôpital Saint Antoine, Paris (P. Coppo)

DOI: <https://doi.org/10.3201/eid2707.204638>

¹These first authors contributed equally to this article.

²Members of this group are listed at the end of this article.

has existed since 1996. This surveillance system comprises 32 pediatric healthcare centers, including all 21 university hospital units specializing in pediatric nephrology. These centers notify public health authorities of cases of STEC-associated HUS. The National Reference Center for *Escherichia coli*, *Shigella* and *Salmonella* at the Institut Pasteur (NRC-Ec; Paris, France) and its associated laboratory at the Robert Debré University Hospital (Paris, France) confirm and characterize STEC infections in children and adults. This surveillance network estimated the annual incidence of HUS in France to be 1.00 case/100,000 child-years, causing a ≈1% death rate during 2007–2016 (5).

Despite the much lower incidence of HUS among adults than children, most deaths caused by STEC-associated HUS occur among persons ≥60 years of age (2,6). The French national health authorities do not have a dedicated surveillance system for STEC-associated HUS in adults. In 2011, a large STEC outbreak in Europe sickened 3,816 persons in Germany, causing 845 cases of HUS and 54 deaths; 24 persons were affected in the Bordeaux region of France, including 9 who had HUS, 8 of whom were adults (7,8). The outbreak was linked to an atypical hybrid pathotype *E. coli* O104:H4 strain characterized by enteroaggregative and enterohemorrhagic virulence; the strain also produced an extended spectrum β-lactamase. Most (88%) patients involved in this outbreak, which was associated with consumption of organic fenu-greek sprouts, were adults, and the median age was 42 years. Publicity surrounding this outbreak raised awareness of STEC-associated HUS in adults. However, cases of STEC-associated HUS in adults remain rare (9,10). Hence, the clinical characteristics of adult STEC-associated HUS and the effects of therapeutic strategies on outcome remain uncertain. We describe the epidemiologic and clinical features of adults with STEC-associated HUS, identify predictors of patient outcomes, and assess the effectiveness of therapeutic interventions in this population.

Methods

Study Design, Settings, and Data Sources

We conducted a retrospective cohort study of STEC-associated HUS cases in adults registered during January 2009–December 2017 in France by the Centre National de Référence des Microangiopathies Thrombotiques (CNR-MAT; <https://www.cnr-mat.fr>). We reviewed all medical files from the CNR-MAT database. This work was part of the TMA study approved by our institutional review board (Comité pour la protection des personnes Ile-de-France; approval no.

CPP04807) in accordance with the Declaration of Helsinki and the French Data Protection Authority.

Diagnostic Criteria

The diagnosis of HUS required the coexistence of TMA (i.e., thrombocytopenia [platelet levels <150,000 cells/μL] and microangiopathic hemolytic anemia [hemoglobin levels <12 g/dL]) and an acute kidney injury (AKI). We included all TMA patients ≥18 years of age in the CNR-MAT cohort who had an AKI and a positive PCR result for the Stx genes *stx1*, *stx2*, or both. We considered patients to have fever if they had a temperature of ≥38°C within 24 hours after admission.

Microbiological Data

Participating laboratories conducted PCR specific for *stx1* and *stx2* on *E. coli* strains isolated from stool, blood, and urine samples. Laboratory technicians also cultured samples from *stx*-positive stools. To characterize the isolated STEC strains, technicians used an O-serogroup multiplex PCR selective for the 10 most frequent serogroups affecting humans in France: O157, O26, O145, O55, O103, O104, O111, O91, O121, and O80 (11). Strains belonging to other serogroups were characterized by PCR of the restriction fragment length polymorphism of the O operon, *rfb* (*rfb*-RFLP) (12). In April 2017, NRC-Ec and local laboratories also began to characterize strains using whole-genome sequencing, when available. If a strain was *stx*-positive but its serogroup was not identified by culture, we classified that strain as not isolated.

Variables

Participating laboratories and physicians submitted data on each patient's medical history, clinical and biological features, microbiological findings, and treatment at admission and during hospitalization (13). We retrospectively calculated each patient's age-weighted Charlson Comorbidity Index (CCI) (14) and classified AKI according to the Kidney Disease: Improving Global Outcomes (KDIGO) criteria published by the International Society of Nephrology (15). We investigated ADAMTS13 and complement alternative pathway (CAP) activity as previously described (16).

Treatments and Outcomes

Treatment consisted mainly of therapeutic plasma exchange (TPE) or best supportive care (BSC) according to the discretion of the treating physician. The C5 complement blocker eculizumab (Soliris; Alexion Pharmaceuticals, Inc., <https://alexion.com>) also

was given at the discretion of the treating physician; however, physicians were encouraged to discuss eculizumab use with a member of the CNR-MAT team. The primary outcome of this study was patient survival at the time of most recent follow-up.

Statistics

We reported qualitative variables as frequencies and percentages; we reported quantitative discrete and continuous variables as medians and interquartile ranges (IQRs). We estimated survival using the Kaplan-Meier method. We used Cox proportional hazards regression to identify factors independently associated with survival. The proportional hazard assumption was supported by a nonsignificant relationship between scaled Schoenfeld residuals and time and refuted by a significant relationship using an alpha (α) risk set at 5%. We reported the results using hazard ratios (HRs) and 95% CIs, using an α risk set at 5% statistical significance. To quantify the effect of eculizumab on survival, we calculated and compared the propensity scores of patients who did and did not use eculizumab (Appendix, <https://wwwnc.cdc.gov/EID/article/27/7/20-4638-App1.pdf>). We used R software version 3.6.1 (The R Project for Statistical Computing, <https://www.r-project.org>) for statistical analysis. For propensity score analysis, we used MatchIt package (17).

Results

Of the 4,048 patients in the CNR-MAT cohort, we first identified 61 adult STEC-associated HUS patients with complete data during January 2009–December 2017. After comparing the NRC-Ec and CNR-MAT surveillance data, we identified 35 additional patients to be included in the study cohort. In total, the study cohort comprised 96 patients (Appendix Figure 1). This cohort included patients from hospitals throughout France, most of which were part of the CNR-MAT network (Figure 1, panel A). The women-to-men ratio was 1.7 and median age was 60.5 years (IQR 47.0–71.0 years) (Figure 1, panel B). Geographic, temporal, and microbiological characteristics of the cases suggested an outbreak among 13 patients (Figure 1). The cohort also included 8 patients affected by the 2011 O104:H4 outbreak in France described previously (8). We found a patient in our cohort who was infected in a family cluster of STEC-associated HUS in 2014, but the strain could not be identified. We also found 4 patients (2 in Marne, 1 in Nord, 1 in Paris) who tested positive for STEC O91 in summer 2013 but did not share a known infection source.

In total, 69 (71.9%) patients had underlying conditions; the median CCI was 2.00 (IQR 1.00–4.25) (Table

1). Of the 96 patients, 27 (28.1%) had an underlying immunodeficiency and 30 (31.3%) had ≥ 1 condition that might contribute to TMA.

Most (83.3%) patients had diarrhea and nearly half (49.0%) had bloody diarrhea; 11 patients had severe colitis, including 4 who required emergency surgery (Table 2). All patients had renal impairment. In 2011, 2 patients with STEC O104:H4 infection had proteinuria (i.e., >1 g/L) but not serum creatinine elevation; these patients also had microangiopathic hemolytic anemia and peripheral thrombocytopenia (8). The other 94 patients all had AKI stage 1 or higher according to KDIGO criteria, of which 61 (63.5%) required dialysis. Of 12 patients who underwent kidney biopsy, 11 showed signs of TMA. Most (76%) patients had neurologic symptoms, mainly confusion (56.3%) and headache (18.8%). Approximately half (52.1%) of patients had a serious neurologic complication such as seizure, coma, or focal deficiency. In addition, 34 (35.4%) patients required mechanical ventilation. In total, 42 patients had high blood pressure ($\geq 150/90$ mm Hg) at admission; severe hypertension ($\geq 170/110$ mm Hg) subsequently developed in 11 patients and hypertensive retinopathy developed in 6 patients. Only 2 patients had hypotension ($\leq 90/60$ mm Hg) at admission. In total, 41 (42.7%) patients had cardiac events; in 26 of 43 cases with available data, patients had troponin levels above the defined threshold of their respective laboratory (Table 2).

CAP measurements during the acute phase of illness were recorded in 69 patients. Of these patients, 36 (52.2%) had values within the reference range (Table 2). Less than 10% of patients had low levels of C3, C4, factor H, or factor I, whereas 26 (38.8%) patients had low levels of CH50. CD46 levels were low in 65.7% (23/35) patients. Two patients had low levels of anti-factor H antibodies (242 and 800 arbitrary units) (Table 2). ADAMTS13 activity was detectable ($\geq 10\%$) in all 69 patients in whom it was tested.

Among the 84 cases in which *stx* type was detected, *stx1*-/*stx2*+ was the most common genotype (85.7%). The *stx1*+/*stx2*- genotype was significantly associated with increased CCI and immunodeficiency (Appendix Table 1). As expected, the most common STEC isolation site was stool (93.8%), whereas only 10 patients had STEC-positive urine or blood samples. Seven (7.3%) patients had STEC-positive urine samples, including 5 who had a urologic infection without associated colitis. Four patients had STEC-positive blood samples, including 1 patient for whom STEC was identified in blood samples only. In total, 5 patients had a multisite infection.

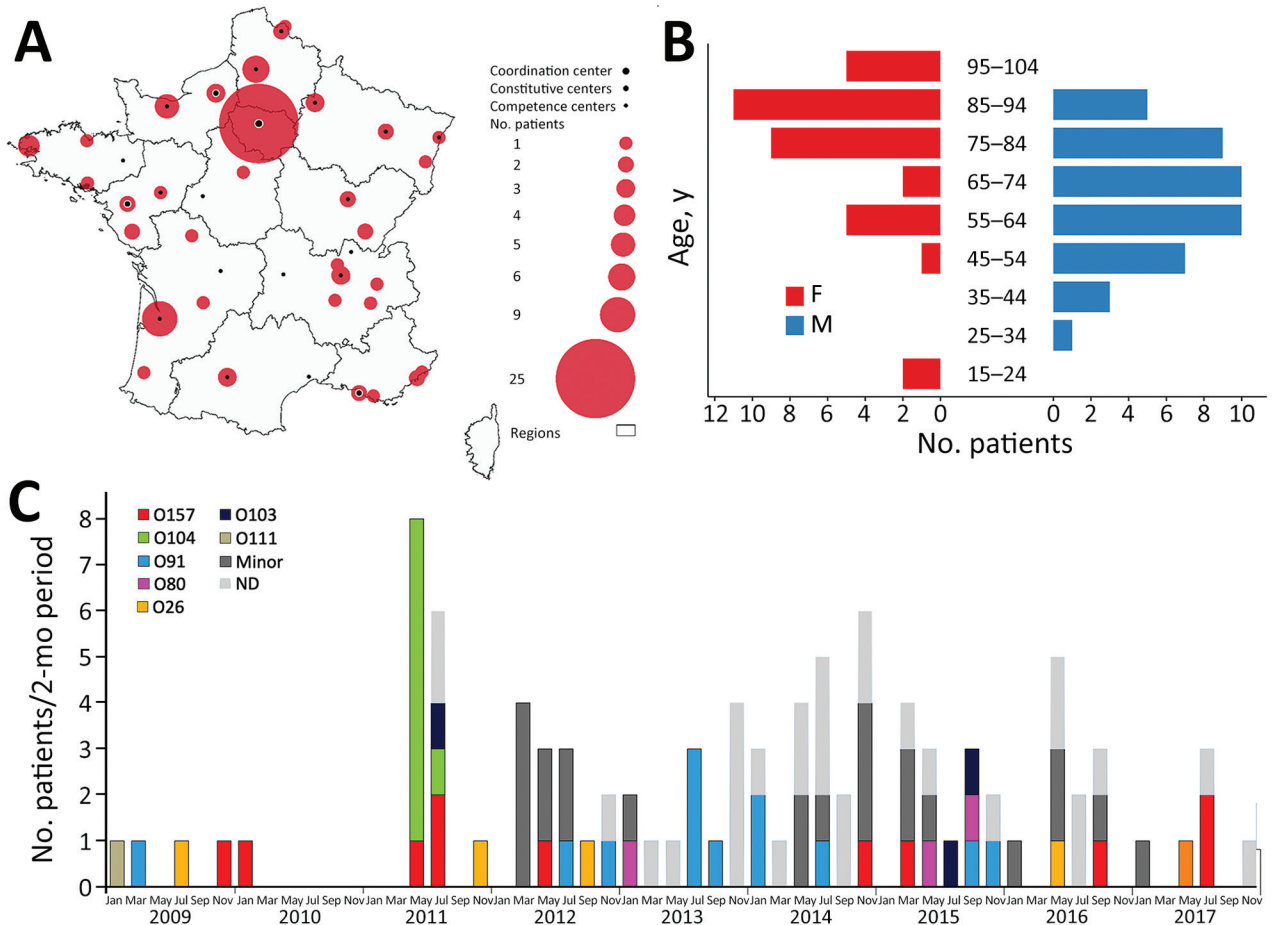


Figure 1. Distribution of adults with Shiga toxin–associated hemolytic uremic syndrome, France, 2009–2017. A) Geographic distribution of cases and thrombotic microangiopathy reference centers. The Centre National de Référence des Microangiopathies Thrombotiques is a national network comprising 1 coordination center, 5 constitutive centers, and 21 competence centers. B) Age and sex distribution of cases. C) Bimonthly distribution of cases according to serogroup. Of patients with minor serogroups, 4 had strains belonging to O106, 3 to O128, 3 to O174, 2 to O113, 1 to O100, 1 to O126, 1 to O148, 1 to O177, 1 to O78, 1 to O84, and 7 to an O serogroup not typable at the time of identification. ND, not determined.

Most (60; 62.5%) had a serogroup typable by the NRC-Ec; 7 (7.3%) patients had an untypable serogroup. The most common serogroups were O91 (12; 17.9%) and O157 (10; 14.9%) (Table 2). The STEC isolates from urine samples belonged to the O104, O91, O106, O126, O174, and O148 serogroups; isolates from blood samples belonged to the O80, O103, and O128 serogroups (Appendix Table 2).

In total, 19 (19.8%) patients died during hospitalization (Figure 2, panel A; Appendix Table 3). Patients died 3–152 days after admission and had a median follow-up period of 112 days (IQR 49–238). After follow-up, 1 patient had HELLP (hemolysis, elevated liver enzymes, low platelets) syndrome; the patient was STEC-negative at the time of the episode. None of the surviving patients had a further episode of TMA during follow-up.

Patients were treated mainly with BSC, TPE, or eculizumab; 3 patients also received immunoadsorption treatment (Appendix Table 4). Of the 61 patients who required dialysis, 17 (25.4%) died. At the end of the follow-up period, 6 (9.8%) patients still required dialysis, including 4 who had a follow-up period of >90 days. Patients who received dialysis were treated for a median duration of 13.5 days (IQR 8–28 days); 38 patients no longer required dialysis at the end of the follow-up period. After a median follow-up period of 34 days (IQR 23–75 days), the median serum creatinine value was 92 $\mu\text{mol/L}$ (IQR 74–124 $\mu\text{mol/L}$). Of the 50 patients with a severe neurologic complication, 14 (28.0%) died. Of the 25 surviving patients with available data, 8 (32.0%) patients had neurologic sequelae, including persistent sensorimotor deficit (7, 28.0%), epilepsy (2, 8.0%), and cognitive impairment (2, 8.0%).

RESEARCH

In total, 26 (27.1%) patients were treated with macrolides, including 3 who received the treatment to prevent infectious meningococcal meningitis associated with eculizumab. Fifty-seven (59.4%) patients received β -lactam antimicrobial drugs, aminoglycosides, or quinolones; 22 (22.9%) patients received metronidazole.

After unadjusted analysis, we found that age (HR 1.04, 95% CI 1.01–1.07; $p = 0.01$), CCI (HR 1.15, 95% CI 1.03–1.28; $p = 0.02$) (Figure 2, panel B), underlying immunodeficiency (HR 4.36, 95% CI 1.72–11.07;

$p < 0.01$), and associated digestive disease (HR 4.07, 95% CI 1.63–10.14; $p < 0.01$) were significantly associated with death of all causes (Table 3). We also found that severe neurologic events (HR 2.90, 95% CI 1.04–8.06; $p = 0.04$), mechanical ventilation (HR 2.71, 95% CI 1.09–6.74; $p = 0.03$), and dialysis (HR 5.57, 95% CI 1.29–24.16; $p = 0.02$) were predictive of death. High troponin levels and *stx* types were not associated with survival (Table 3). Most patients who died had STEC strains belonging to non-O104 and non-O157

Table 1. Characteristics of adults with Shiga toxin–associated hemolytic uremic syndrome, France, 2009–2017*

Characteristic	Value
Median age, y (IQR)	60.5 (47.00–71.00)
Sex	
M	35 (36.5)
F	61 (63.5)
Median age-weighted Charlson Comorbidity Index (IQR)	2.00 (1.00–4.25)
Tobacco use within previous 3 y	12 (12.5)
>1 underlying condition	69 (71.9)
Cardiovascular disease	48 (50.0)
Arterial hypertension	38 (39.6)
Diabetes mellitus	12 (12.5)
Venous thromboembolic disease	11 (11.5)
Heart disease†	20 (20.8)
CKD‡	15 (15.6)
History of kidney transplant	5 (5.2)
Stage 2 CKD	4 (4.2)
Stage 3 CKD	8 (8.3)
Stage 4 CKD	3 (3.1)
Digestive disorder§	29 (30.2)
Gastrointestinal disorder	18 (18.8)
Biliopancreatic disorder	9 (9.4)
Hepatic disorder	4 (4.2)
Autoimmune or inflammatory disease¶	11 (11.5)
Immunodeficiency	27 (28.1)
History of bone marrow or solid organ transplant#	8 (8.3)
Hematologic disease**	8 (8.3)
Active cancer††	8 (8.3)
HIV‡‡	3 (3.1)
Primary immunodeficiency§§	2 (2.1)
Neuropsychiatric disorder¶¶	18 (18.8)
Treatment	
Immunosuppressive treatment	12 (12.5)
Corticosteroids	11 (11.5)
Calcineurin inhibitors	7 (7.3)
Azathioprine or mycophenolate mofetil	7 (7.3)

*Values are no. (%) patients except as indicated. CKD, chronic kidney disease; IQR, interquartile range.

†8 patients had hypertensive disease, 4 had ischemic disease, 4 had hypertensive and ischemic disease, 2 had valvular cardiopathy, 1 had pulmonary hypertension, and 1 had unspecified heart disease.

‡According to Kidney Disease Improving Global Outcomes guidelines (15).

§8 patients had gastric, small bowel, or colonic resection; 2 had history of bariatric surgery; 3 had chronic diarrhea from diverticulosis; 1 had graft-versus-host disease; 1 had colonic endometriosis; 1 had microscopic colitis; 1 had AA amyloidosis; 1 had neurovegetative disorder (1 each); 3 had recurrent pyogenic cholangitis; 1 had sclerosing cholangitis; 2 had a double kidney-pancreas transplantation; 3 had chronic pancreatitis; 2 had cirrhosis; 1 had history of liver transplant; and 1 had autoimmune hepatitis.

¶2 patients had mixed connective tissue disease, 1 had systemic sclerosis, 1 had sclerosing cholangitis, 1 had microscopic polyangiitis, 3 had type 1 diabetes, 1 had multiple sclerosis, and 2 had psoriasis.

#3 patients had a history of kidney, 2 of double kidney–pancreas, 2 of bone marrow, and 1 of liver transplant.

**1 patient had acute myeloid leukemia, 1 had chronic lymphocytic leukemia, 1 had Hodgkin's lymphoma, 1 had clonal B-cell lymphocytosis, 1 had monoclonal gammopathy of undetermined significance, 1 had Waldenström's disease, 1 had myeloproliferative disorder, and 1 had myelodysplastic syndrome.

††3 patients had breast cancer, 1 had metastatic lung cancer, 1 had a gastrointestinal stromal cell tumor, 1 had bladder cancer, 1 had cervical cancer, and 1 had gastric cancer.

‡‡3 patients had AIDS, including 2 patients who received HIV diagnoses during treatment.

§§2 patients had hypogammaglobulinemia, including 1 patient who had ICF1 syndrome caused by a DNMT3b germinal mutation.

¶¶5 patients had stroke sequelae, 3 had Parkinson's disease, 1 had multiple sclerosis, 4 had cognitive impairment (including 1 patient who had Korsakoff syndrome and 1 who had vascular dementia), 1 had epilepsy, 1 had chronic polyradiculoneuropathy, and 5 had major depressive or bipolar disorder.

serogroups (Figure 2, panel C; Appendix Table 3). We found that overall survival was comparable among patients treated by different combinations of BSC,

TPE, and eculizumab ($p = 0.43$ by log-rank test) (Table 3; Figure 2, panel D). The use of macrolides was not associated with survival ($p = 0.77$).

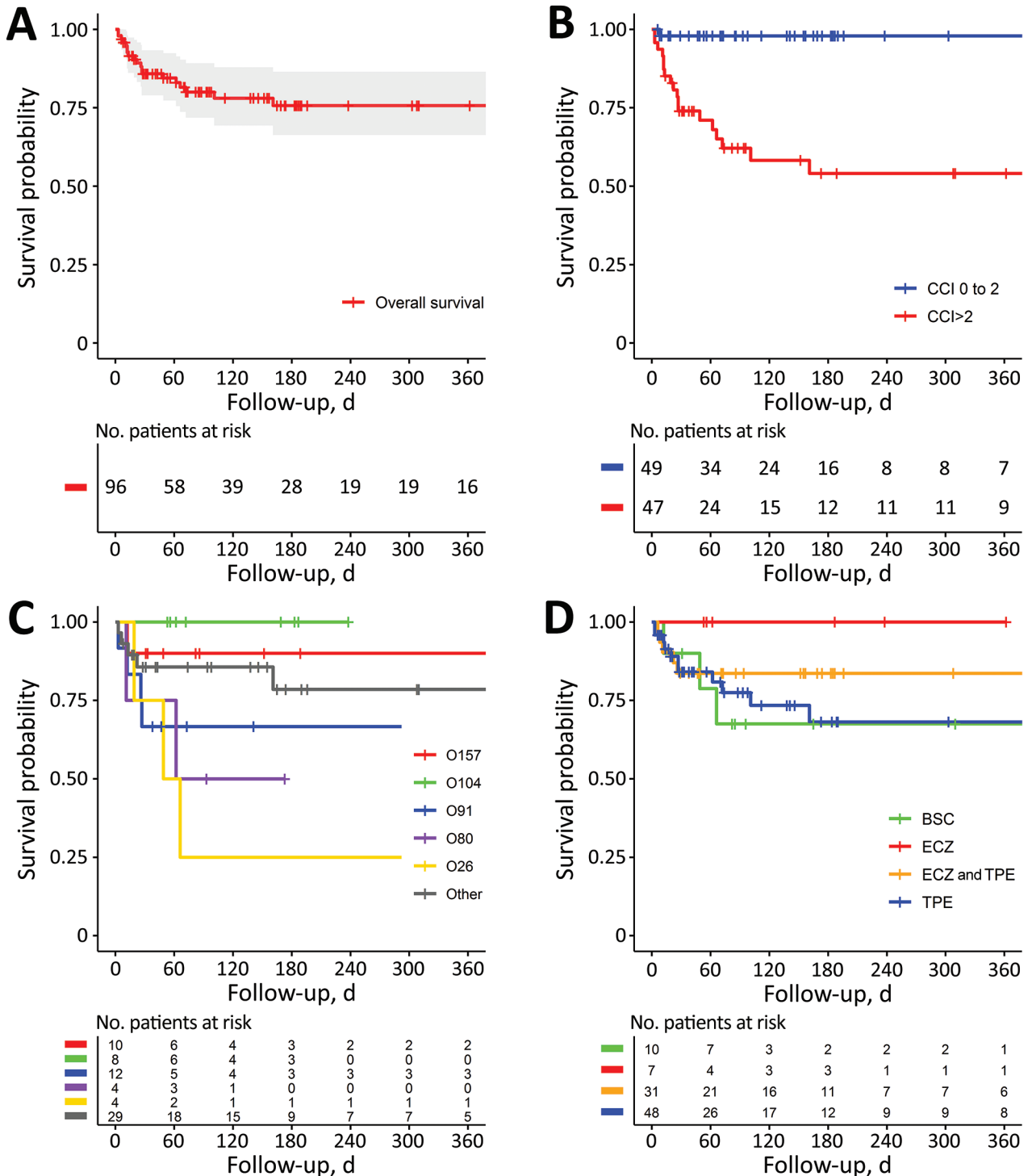


Figure 2. Kaplan-Meier survival plots of adults with Shiga toxin–associated hemolytic uremic syndrome, France, 2009–2017. A) Overall. B) By age-weighted Charlson comorbidity index. C) By STEC serogroup. D) By treatment. Plots show time from admission to death. p values determined using log-rank test. BSC, best standard of care; CCI, age-weighted Charlson comorbidity index; ECZ, eculizumab; TPE, therapeutic plasma exchange.

Multivariate analysis showed that underlying immunodeficiency (HR 3.54, 95% CI 1.24–10.14; $p = 0.02$) and severe neurologic events (HR 3.40, 95% CI 1.05–11.04; $p = 0.04$) were negatively associated with survival (Table 3). After adjustment of determinants retained for the multivariate analysis, we found that eculizumab was not associated with survival (HR 0.77, 95% 0.25–2.33; $p = 0.64$). Propensity score-matching also indicated that eculizumab was not associated with survival ($p = 0.34$) (Appendix Table 5, Figure 2).

Discussion

We found that 20% of adults who had STEC-associated HUS died during hospitalization, in agreement with previous findings (9,10); however, <1% of children who had STEC-associated HUS died in France during the same years, 2007–2016 (5). In addition, adults had cerebral involvement 3 times more frequently than children (2); 52.1% of adult patients had severe neurologic manifestations, similar to the observations of Karpac et al. (10). Renal recovery was slow and inconsistent; 4 patients still required dialysis 90 days after hospitalization (9). One third of patients required mechanical ventilation. These findings emphasize that, in adults, STEC-associated HUS is a severe systemic disease that can cause multiple organ failure. However, inclusion in the CNR-MAT registry relied on voluntary physician reporting; thus, this case series is not exhaustive and might disproportionately reflect the most severe cases. As previously observed for children (18), most cases in this cohort were sporadic and, for unclear reasons, in women. In regard to age distribution, STEC-associated HUS has a U curve from birth to old age (6,9,10,19). During the study period, 1,095 STEC-associated HUS cases in children were reported to Santé Publique France through the country's pediatric surveillance network (5). By comparison, this disease appears to be much rarer among adults, although underreporting is probable.

Our findings on underlying conditions and deaths by age group resemble those of the FoodNet registry of elderly adults with STEC-associated HUS (9). The risk for death from STEC-associated HUS increases for persons age >40 years, suggesting that young and middle-aged adults have similar clinical courses to those observed in children. We found a strong association between underlying conditions and decreased survival, especially for patients with immunodeficiency (9,20–23). The prevalence of antibodies against Stx decreases for persons >40 years of age (24), which might account for the more severe forms of STEC-associated HUS in elderly persons. The expression of glomerular globotriaosylceramide (Gb3), the main receptor of Stx,

was thought to decrease with age; however, researchers now believe that expression levels remain stable throughout a person's lifetime (25). Renal and neurologic signs similar to those caused by HUS develop in immunocompromised mice after STEC inoculation or Stx exposure, whereas wild-type mice are naturally resistant to this disease (26,27). Together, these findings highlight the role of the immune system in preventing STEC-associated HUS. Immunodeficiency probably contributes to disease severity.

The 2011 outbreak in Europe illustrated that microbiological characteristics play a key role in STEC-associated HUS (7). The distribution of serotypes among adults in our study was slightly different than in a study on pediatric HUS in France in the same timeframe (5). Non-O157 strains were more prevalent in the pediatric series (5) and in ours, whereas O157 and O26 were more commonly observed among children than adults (23% among children vs. 15% among adults for O157; 11% among children vs. 6% among adults for O26) (5). A similar overall distribution was observed among children and adults with STEC infection in Norway (23% for O157, 10% for O26) (28). By contrast, serogroups O91 and O104 have been mainly found among adults (29,30). The data might have been skewed by the 2011 outbreak caused by a strain belonging to the O104 serogroup; this outbreak caused infections in younger persons who had fewer underlying conditions, which could account for the better outcomes of those patients. Other serogroups, especially O80, O26, and O91, are emerging and might be associated with increased pathogenicity (2,18). STEC O91 was also the most common serogroup among adults with STEC infections in Germany (30), which raises the question of increased pathogenicity in adults and in persons >40 years of age.

In agreement with previous reports of STEC-associated HUS in adults (29,31), we found that *stx1+*/*stx2-* strains were more prevalent among adults (14.3%) than had been previously documented among children (2.0%) (5). One possible explanation for this distribution might be that in some patients, HUS was concurrent with but unrelated to infection or colonization by *stx1+*/*stx2-* STEC; however, this scenario is unlikely because STEC-positive patients had typical features of HUS in an infectious context. We cannot exclude the possibility that the *stx2* gene could have been lost in human hosts during infection or ex vivo during subculture, as already described for STEC O26 (32). In this series, all *stx1+*/*stx2-* strains belonged to non-O157 serogroups. These findings are similar to those of Käppeli et al. (29), who found that 15.8% of cases of non-O157 STEC-associated HUS were caused

by *stx1+*/*stx2-* strains, which could suggest that different serogroups might pose different risks for HUS associated with particular *stx* genotypes. Last, most (83%) patients with *stx1+*/*stx2-* genotypes had underlying immunodeficiency; one explanation could be that immunodeficient patients are more susceptible to Stx1. The alleles *stx1* and *stx2c* have been associated with a lower risk for severe STEC infection and HUS (28). However, *stx1a* is associated with higher risk for severe STEC infection (33). We did not have data on *stx* subtypes in our study.

We observed CAP abnormalities similar to those previously reported in a cohort of 113 cases of STEC-associated HUS in children (16). We found that 65.7% of patients had low CD46 and 38.8% had low CH50 levels. However, a decrease in the concentration of complement factors, the interpretation of which remains equivocal, might be attributable to kidney damage and STEC-associated HUS (16). The presence of an inflammatory syndrome further complicates the interpretation of these data. In contrast to atypical HUS, pediatric STEC-associated HUS has not been linked to a constitutional or acquired dysregulation of the CAP. Screening for variants in complement genes is not usually conducted among children with STEC-associated HUS. Similarly, it seems unlikely that STEC infection reveals underlying CAP abnormalities in many adults.

We found that 7% of patients had STEC-positive urine samples, an underrecognized finding documented by Lavrek et al. (34). Although urine samples might be easily contaminated, especially in patients who have diarrhea, these findings encourage systematic STEC-specific PCR screening and culture confirmation of stool or other biological samples (in the event of extraintestinal *E. coli* infection) from adult TMA patients (2,34).

Because the effectiveness of specific treatments remains unclear, BSC is the cornerstone of STEC-associated HUS treatment (2,35,36). Univariate analysis indicated that TPE was not associated with overall survival improvement, although other studies have concluded differently (37–39). However, considering the substantial overlap between the signs and symptoms of STEC-associated HUS in adults and TMA of other etiologies, some researchers believe that plasma therapy should be given until TTP or atypical HUS are ruled out (13,40). Whether TPE should be continued after the determination of *stx* status remains unclear. As previously reported, we did not find a clear survival benefit from eculizumab (38,41). However, the small sample size and the strong differences between patients who did and did not receive eculizumab treatment preclude definitive conclusions.

The benefits of antimicrobial drugs in treating STEC-associated HUS are unclear (42,43). Previous studies suggest that the use of antimicrobial drugs during early stages of STEC infection is associated with the development of HUS. However, the effects of antimicrobial drugs administered after HUS diagnosis remain unknown (42). A retrospective study reported that azithromycin administered during STEC infection might reduce the duration of STEC carriage (43). We found that use of macrolides was not associated with survival. This observation might have been confounded by possible unreported administration of antimicrobial drugs before hospitalization, treatment for unstandardized indications at the discretion of the practitioner, or other variables. We also found that the prescription of multiple antimicrobial drugs was a common practice, especially in cases of severe infection.

In conclusion, STEC-associated HUS is rarer among adults than among children but causes more severe disease and death. Underlying conditions, especially immunodeficiency, are strongly associated with decreased survival. The severity of the disease, a probably underestimated prevalence, and the risk for outbreaks of emerging STEC-associated HUS provide strong arguments for active epidemiologic and microbiological surveillance of this disease.

Members of the Reference Center for Thrombotic Microangiopathies team: Jean-François Augusto, Elie Azoulay, Virginie Barbay, Ygal Benhamou, Dominique Bordessoule, Christophe Charasse, Anne Charvet-Rumpler, Dominique Chauveau, Gabriel Choukroun, Jean-Philippe Coindre, Paul Coppo, Elise Corre, Yahsou Delmas, Georges Deschenes, Alain Devidas, Antoine Dossier, Olivier Fain, Fadi Fakhouri, Véronique Frémeaux-Bacchi, Lionel Galicier, Steven Grangé, Bertrand Guidet, Jean-Michel Halimi, Mohamed Hamidou, Raoul Herbrecht, Miguel Hié, Frédéric Jacobs, Bérange Joly, Tarik Kanouni, Gilles Kaplanski, Alexandre Lautrette, Véronique Le Guern, Bruno Moulin, Christiane Mousson, Mario Ojeda Uribe, Abdelkader Ouchenir, Nathalie Parquet, Frédéric Pène, Pierre Perez, Pascale Poullin, Claire Pouteil-Noble, Claire Presne, François Provôt, Eric Rondeau, Samir Saheb, Amélie Seguin, Aude Servais, Alain Stépanian, Agnès Veyradier, Cécile Vigneau, Alain Wynckel, and Patricia Zunic.

Acknowledgments

We thank Eric Alamartine, Sandrine Bedon Carte, Gilles Bernardin, Severin Cabasson, Vincent Cadiergue, Jean-François Cerfon, Thomas Crepin, Vincent Das, Philippe De Swardt, Geneviève Dumont, Alexandre Hertig, Jean Claude Lacherade, Olivier Leroy, Julie Le Scanff, Didier Perez, Emilie Pinçon, Jean-Pierre Quenot, Felipe Suarez, Rachel Tetaz,

Olivier Thauvat, Jean Marc Thouret, and Xavier Valette for their valuable collaboration with the CNR-MAT. These physicians played an active role in care of patients with STEC-associated HUS.

This work was partly funded by grants from the French Ministry of Health (Programme Hospitalier de Recherche Clinique [grant nos. P120118 and AOM12259]). B.T. has received a research grant from CSL Behring (<https://www.cslbehring.com>). E.R. and C.P. are members of the Advisory Board for Alexion Pharmaceuticals, Inc. (<https://alexion.com>). Y.D. has received lecture fees from Alexion Pharmaceuticals, Inc. and honorarium as advisory board member for Sanofi (<https://www.sanofi.com>). F.F. has received consultancy fees and speaker honoraria from F. Hoffmann-La Roche Ltd (<https://www.roche.com>); Alexion Pharmaceuticals, Inc.; Apellis Pharmaceuticals (<https://apellis.com>); Achillion Pharmaceuticals; Novartis AG (<https://www.novartis.com>); and Alnylam Pharmaceuticals, Inc. (<https://www.alnylam.com>). P.C. is member of the advisory boards of Alexion Pharmaceuticals, Inc.; Sanofi; Shire P.C.C. (<http://shirepcc.com>); Takeda Pharmaceutical Company Limited (<https://www.takeda.com>); and Octapharma AG (<https://www.octapharma.com>); he has received consultancy fees and speaker honoraria from Sanofi; Alexion Pharmaceuticals, Inc.; and Takeda Pharmaceutical Company Limited.

About the Author

Dr. Travert is a medical intern at Assistance Publique-Hôpitaux de Paris in Paris, France. His research interests include the epidemiology of STEC-associated HUS and environmental factors contributing to autoimmune diseases.

References

- Karmali MA, Steele BT, Petric M, Lim C. Sporadic cases of haemolytic-uraemic syndrome associated with faecal cytotoxin and cytotoxin-producing *Escherichia coli* in stools. *Lancet*. 1983;321:619–20. [https://doi.org/10.1016/S0140-6736\(83\)91795-6](https://doi.org/10.1016/S0140-6736(83)91795-6)
- Joseph A, Coite A, Mariani Kurkdjian P, Rafat C, Hertig A. Shiga toxin-associated hemolytic uremic syndrome: a narrative review. *Toxins (Basel)*. 2020;12:67. <https://doi.org/10.3390/toxins12020067>
- Fakhouri F, Zuber J, Frémeaux-Bacchi V, Loirat C. Haemolytic uraemic syndrome. *Lancet*. 2017;390:681–96. [https://doi.org/10.1016/S0140-6736\(17\)30062-4](https://doi.org/10.1016/S0140-6736(17)30062-4)
- Majowicz SE, Scallan E, Jones-Bitton A, Sargeant JM, Stapleton J, Angulo FJ, et al. Global incidence of human Shiga toxin-producing *Escherichia coli* infections and deaths: a systematic review and knowledge synthesis. *Foodborne Pathog Dis*. 2014;11:447–55. <https://doi.org/10.1089/fpd.2013.1704>
- Bruyand M, Mariani-Kurkdjian P, Le Hello S, King L-A, Van Cauteren D, Lefevre S, et al.; Réseau Français Hospitalier de Surveillance du Shu Pédiatrique. Paediatric haemolytic uraemic syndrome related to Shiga toxin-producing *Escherichia coli*, an overview of 10 years of surveillance in France, 2007 to 2016. *Euro Surveill*. 2019;24:24. <https://doi.org/10.2807/1560-7917.ES.2019.24.8.1800068>
- Gould LH, Demma L, Jones TF, Hurd S, Vugia DJ, Smith K, et al. Hemolytic uremic syndrome and death in persons with *Escherichia coli* O157:H7 infection, foodborne diseases active surveillance network sites, 2000–2006. *Clin Infect Dis*. 2009;49:1480–5. <https://doi.org/10.1086/644621>
- Frank C, Werber D, Cramer JP, Askar M, Faber M, an der Heiden M, et al.; HUS Investigation Team. Epidemic profile of Shiga-toxin-producing *Escherichia coli* O104:H4 outbreak in Germany. *N Engl J Med*. 2011;365:1771–80. <https://doi.org/10.1056/NEJMoa1106483>
- Delmas Y, Vendrely B, Clouzeau B, Bachir H, Bui H-N, Lacraz A, et al. Outbreak of *Escherichia coli* O104:H4 haemolytic uraemic syndrome in France: outcome with eculizumab. *Nephrol Dial Transplant*. 2014;29:565–72. <https://doi.org/10.1093/ndt/gft470>
- Gould LH, Jordan JG, Dunn J, Apostol M, Griffin PM; Emerging Infections Program FoodNet Working Group. Postdiarrheal hemolytic uremic syndrome in persons aged 65 and older in FoodNet sites, 2000–2006. *J Am Geriatr Soc*. 2011;59:366–8. <https://doi.org/10.1111/j.1532-5415.2011.03269.x>
- Karpac CA, Li X, Terrell DR, Kremer Hovinga JA, Lämmle B, Vesely SK, et al. Sporadic bloody diarrhoea-associated thrombotic thrombocytopenic purpura-haemolytic uraemic syndrome: an adult and paediatric comparison. *Br J Haematol*. 2008;141:696–707. <https://doi.org/10.1111/j.1365-2141.2008.07116.x>
- Soysal N, Mariani-Kurkdjian P, Smail Y, Liguori S, Gouali M, Loukiadis E, et al. Enterohemorrhagic *Escherichia coli* hybrid pathotype O80:H2 as a new therapeutic challenge. *Emerg Infect Dis*. 2016;22:1604–12. <https://doi.org/10.3201/eid2209.160304>
- Coimbra RS, Grimont F, Lenormand P, Burguière P, Beutin L, Grimont PA. Identification of *Escherichia coli* O-serogroups by restriction of the amplified O-antigen gene cluster (rfb-RFLP). *Res Microbiol*. 2000;151:639–54. [https://doi.org/10.1016/S0923-2508\(00\)00134-0](https://doi.org/10.1016/S0923-2508(00)00134-0)
- Coppo P, Schwarzinger M, Buffet M, Wynckel A, Clabault K, Presne C, et al.; French Reference Center for Thrombotic Microangiopathies. Predictive features of severe acquired ADAMTS13 deficiency in idiopathic thrombotic microangiopathies: the French TMA reference center experience. *PLoS One*. 2010;5:e10208. <https://doi.org/10.1371/journal.pone.0010208>
- Charlson M, Szatrowski TP, Peterson J, Gold J. Validation of a combined comorbidity index. *J Clin Epidemiol*. 1994;47:1245–51. [https://doi.org/10.1016/0895-4356\(94\)90129-5](https://doi.org/10.1016/0895-4356(94)90129-5)
- International Society of Nephrology. KDIGO clinical practice guideline for acute kidney injury. 2012 [cited 2020 Jul 31]. <https://kdigo.org/wp-content/uploads/2016/10/KDIGO-2012-AKI-Guideline-English.pdf>
- Frémeaux-Bacchi V, Sellier-Leclerc A-L, Vieira-Martins P, Limou S, Kwon T, Lahoche A, et al. Complement gene variants and Shiga toxin-producing *Escherichia coli*-associated hemolytic uremic syndrome. *Clin J Am Soc Nephrol*. 2019;14:364–77. <https://doi.org/10.2215/CJN.05830518>
- Ho D, Imai K, King G, Stuart EA. MatchIt: nonparametric preprocessing for parametric causal inference. *J Stat Softw*. 2011;42:1–28. <https://doi.org/10.18637/jss.v042.i08>
- Griffin PM, Karmali MA. Emerging public health challenges of Shiga toxin-producing *Escherichia coli* related to changes in the pathogen, the population, and the environment. *Clin Infect Dis*. 2017;64:371–6. <https://doi.org/10.1093/cid/ciw708>

19. Carter AO, Borczyk AA, Carlson JAK, Harvey B, Hockin JC, Karmali MA, et al. A severe outbreak of *Escherichia coli* O157:H7–associated hemorrhagic colitis in a nursing home. *N Engl J Med*. 1987;317:1496–500. <https://doi.org/10.1056/NEJM198712103172403>
20. Ville S, Ydee A, Garandeau C, Canet E, Tissot A, Cantarovich D, et al. Shiga toxin-producing *Escherichia coli*–associated hemolytic uremic syndrome in solid organ transplant recipients. *Kidney Int*. 2019;96:1423–4. <https://doi.org/10.1016/j.kint.2019.08.024>
21. Farina C, Gavazzeni G, Caprioli A, Remuzzi G. Hemolytic uremic syndrome associated with verocytotoxin-producing *Escherichia coli* infection in acquired immunodeficiency syndrome. *Blood*. 1990;75:2465. <https://doi.org/10.1182/blood.V75.12.2465a.2465a>
22. Vera-Aguilera J, Duma N, Gast K, Alkhateeb H, Tande A, Leung N, et al. Hemolytic uremic syndrome associated with *Escherichia coli* O157 infection in an allogenic stem cell transplant recipient. *Mayo Clin Proc Innov Qual Outcomes*. 2018;2:387–91. <https://doi.org/10.1016/j.mayocpiqo.2018.07.001>
23. Manière L, Domenger C, Camara B, Giovannini D, Malvezzi P, Rostaing L. An atypical case of Shiga toxin producing-*Escherichia coli* hemolytic and uremic syndrome (STEC-HUS) in a lung transplant recipient. *Case Rep Transplant*. 2019;2019:9465040. <https://doi.org/10.1155/2019/9465040>
24. Karmali MA, Mascarenhas M, Petric M, Dutil L, Rahn K, Ludwig K, et al. Age-specific frequencies of antibodies to *Escherichia coli* verocytotoxins (Shiga toxins) 1 and 2 among urban and rural populations in southern Ontario. *J Infect Dis*. 2003;188:1724–9. <https://doi.org/10.1086/379726>
25. Ergonul Z, Clayton F, Fogo AB, Kohan DE. Shiga toxin-1 binding and receptor expression in human kidneys do not change with age. *Pediatr Nephrol*. 2003;18:246–53. <https://doi.org/10.1007/s00467-002-1025-9>
26. Brando RJF, Miliwebsky E, Bentancor L, Deza N, Baschkier A, Ramos MV, et al. Renal damage and death in weaned mice after oral infection with Shiga toxin 2–producing *Escherichia coli* strains. *Clin Exp Immunol*. 2008;153:297–306. <https://doi.org/10.1111/j.1365-2249.2008.03698.x>
27. Karpman D, Connell H, Svensson M, Scheutz F, Aim P, Svanborg C. The role of lipopolysaccharide and Shiga-like toxin in a mouse model of *Escherichia coli* O157:H7 infection. *J Infect Dis*. 1997;175:611–20. <https://doi.org/10.1093/infdis/175.3.611>
28. Brandal LT, Wester AL, Lange H, Løbersli I, Lindstedt B-A, Vold L, et al. Shiga toxin-producing *Escherichia coli* infections in Norway, 1992–2012: characterization of isolates and identification of risk factors for haemolytic uremic syndrome. *BMC Infect Dis*. 2015;15:324. <https://doi.org/10.1186/s12879-015-1017-6>
29. Käppeli U, Hächler H, Giezendanner N, Beutin L, Stephan R. Human infections with non-O157 Shiga toxin-producing *Escherichia coli*, Switzerland, 2000–2009. *Emerg Infect Dis*. 2011;17:180–5. <https://doi.org/10.3201/eid1702.100909>
30. Werber D, Beutin L, Pichner R, Stark K, Fruth A. Shiga toxin-producing *Escherichia coli* serogroups in food and patients, Germany. *Emerg Infect Dis*. 2008;14:1803–6. <https://doi.org/10.3201/eid1411.080361>
31. Adams NL, Byrne L, Smith GA, Elson R, Harris JP, Salmon R, et al. Shiga toxin-producing *Escherichia coli* O157, England and Wales, 1983–2012. *Emerg Infect Dis*. 2016;22:590–7. <https://doi.org/10.3201/eid2204.151485>
32. Bielaszewska M, Prager R, Köck R, Mellmann A, Zhang W, Tschäpe H, et al. Shiga toxin gene loss and transfer in vitro and in vivo during enterohemorrhagic *Escherichia coli* O26 infection in humans. *Appl Environ Microbiol*. 2007;73:3144–50. <https://doi.org/10.1128/AEM.02937-06>
33. Byrne L, Adams N, Jenkins C. Association between Shiga toxin–producing *Escherichia coli* O157:H7 stx gene subtype and disease severity, England, 2009–2019. *Emerg Infect Dis*. 2020;26:2394–400. <https://doi.org/10.3201/eid2610.200319>
34. Lavrek D, Lava SAG, Milani GP, Simonetti GD, Bianchetti MG, Giannini O. Hemolytic-uremic syndrome after *Escherichia coli* urinary tract infection in humans: systematic review of the literature. *J Nephrol*. 2018;31:919–24. <https://doi.org/10.1007/s40620-018-0543-x>
35. Michael M, Elliott EJ, Ridley GF, Hodson EM, Craig JC. Interventions for haemolytic uraemic syndrome and thrombotic thrombocytopenic purpura. *Cochrane Database Syst Rev*. 2009;1:CD003595. <https://doi.org/10.1002/14651858.CD003595.pub2>
36. Grisar S, Xie J, Samuel S, Hartling L, Tarr PI, Schnadower D, et al.; Alberta Provincial Pediatric Enteric Infection Team. Associations between hydration status, intravenous fluid administration, and outcomes of patients infected with Shiga toxin–producing *Escherichia coli*: a systematic review and meta-analysis. *JAMA Pediatr*. 2017;171:68–76. <https://doi.org/10.1001/jamapediatrics.2016.2952>
37. Keenswijk W, Raes A, De Clerck M, Vande Walle J. Is plasma exchange efficacious in Shiga toxin–associated hemolytic uremic syndrome? A narrative review of current evidence. *Ther Apher Dial*. 2019;23:118–25. <https://doi.org/10.1111/1744-9987.12768>
38. Kielstein JT, Beutel G, Fleig S, Steinhoff J, Meyer TN, Hafer C, et al.; Collaborators of the DGfN STEC-HUS registry. Best supportive care and therapeutic plasma exchange with or without eculizumab in Shiga-toxin–producing *E. coli* O104:H4 induced haemolytic-uraemic syndrome: an analysis of the German STEC-HUS registry. *Nephrol Dial Transplant*. 2012;27:3807–15. <https://doi.org/10.1093/ndt/gfs394>
39. Padmanabhan A, Connelly-Smith L, Aquil N, Balogun RA, Klingel R, Meyer E, et al. Guidelines on the use of therapeutic apheresis in clinical practice—evidence-based approach from the Writing Committee of the American Society for Apheresis: the eighth special issue. *J Clin Apher*. 2019;34:171–354. <https://doi.org/10.1002/jca.21705>
40. Joseph A, Rafat C, Zafrani L, Mariani-Kurkdjian P, Veyradier A, Hertig A, et al. Early differentiation of Shiga toxin–associated hemolytic uremic syndrome in critically ill adults with thrombotic microangiopathy syndromes. *Crit Care Med*. 2018;46:e904–11. <https://doi.org/10.1097/CCM.0000000000003292>
41. Lapeyraque A-L, Malina M, Fremaux-Bacchi V, Boppel T, Kirschfink M, Oualha M, et al. Eculizumab in severe Shiga-toxin–associated HUS. *N Engl J Med*. 2011;364:2561–3. <https://doi.org/10.1056/NEJMc1100859>
42. Wong CS, Mooney JC, Brandt JR, Staples AO, Jelacic S, Boster DR, et al. Risk factors for the hemolytic uremic syndrome in children infected with *Escherichia coli* O157:H7: a multivariable analysis. *Clin Infect Dis*. 2012;55:33–41. <https://doi.org/10.1093/cid/cis299>
43. Nitschke M, Sayk F, Härtel C, Roseland RT, Hauswaldt S, Steinhoff J, et al. Association between azithromycin therapy and duration of bacterial shedding among patients with Shiga toxin–producing enteroaggregative *Escherichia coli* O104:H4. *JAMA*. 2012;307:1046–52. <https://doi.org/10.1001/jama.2012.264>

Address for correspondence: Paul Coppo, AP-HP.6 and Sorbonne Université, 184 rue du Faubourg Saint-Antoine, Assistance Publique–Hôpitaux de Paris, 75012 Paris, France; email: paul.coppo@aphp.fr

Shiga Toxin–Associated Hemolytic and Uremic Syndrome in Adults, France, 2009–2017

Appendix

Appendix Methods

We used logistic regression to estimate the distance measure for propensity score-matching. We used the nearest neighbor method with caliper index equal to 0.3 to conduct 1:1 matching, because this method balanced the number of unmatched patients and acceptable comparability between matched groups. We created a random computerized loop to optimize best matching for our final analysis. Other methods, including exact, stratification matching, or many-to-one matching on the propensity score, did not enable the formation of acceptable comparison groups because patients treated with eculizumab had different characteristics that appeared to be the main determinants of survival (Appendix Table 6; Appendix Figure 2).

Appendix Table 1. Clinical and biological characteristics of 96 adults with Shiga toxin–associated hemolytic uremic syndrome by infecting strain genotype, France, 2009–2017*

Characteristic	stx genotype			p value¶
	stx1+/stx2-†	stx1-/stx2+‡	stx1+/stx2+§	
Total	12	63	9	
Clinical features				
Sex				
M	5 (41.7)	24 (38.1)	3 (33.3)	
F	7 (58.3)	39 (61.9)	6 (66.7)	1.00
Median age, y (IQR)	52.50 (42.75–60.25)	62.00 (51.00–71.00)	56.00 (43.00–64.00)	0.24
Median age-weighted	6.00 (4.75–7.25)	2.00 (0.50–4.00)	2.00 (0.00–3.00)	<0.01
Charlson comorbidity index (IQR)				
≥1 Underlying condition	12 (100.0)	46 (73.0)	6 (66.7)	0.07
Immunodeficiency	10 (83.3)	15 (23.8)	1 (11.1)	<0.01
Digestive disorder	8 (66.7)	15 (23.8)	3 (33.3)	0.02
Cardiovascular disease	10 (83.3)	29 (46.0)	4 (44.4)	0.06
Diarrhea	9 (75.0)	54 (85.7)	6 (66.7)	0.12
Bloody diarrhea	5 (41.7)	29 (46.0)	5 (55.6)	0.87
Isolation site				
Stool	10 (83.3)	61 (96.8)	8 (88.9)	0.10
Urine	1 (8.3)	4 (6.3)	1 (11.1)	0.63
Blood	2 (16.7)	1 (1.6)	0	0.08
Events and outcomes				
Dialysis	9 (75.0)	40 (63.5)	5 (55.6)	0.60
Stroke, coma, or seizure	6 (50.0)	33 (52.4)	5 (55.6)	1.00
Any cardiac event	6 (50.0)	23 (36.5)	6 (66.7)	0.22
Death	4 (33.3)	13 (20.6)	1 (11.1)	0.45

*Values are no. (%), except as indicated.

†Three stx1-positive strains belonged to serogroup O103, 3 to O26, 1 to O111, 1 to O126, 1 to O128, 1 to O78, and 1 to O84; 1 strain was not typable.

‡Twelve stx2-positive strains belonged to serogroup O91, 9 to O157, 8 to O104, 4 to O106, 4 to O80, 2 to O113, 2 to O128, 2 to O174, 1 to O100, 1 to O148, 1 to O177, and 1 to O26; 3 strains were not typable. Remaining strains were not isolated or not available.

§One stx1-positive/stx2-positive strain belonged to serogroup O157 and 1 belonged to O174; 3 strains were not typable.

¶P values were determined by Fisher exact test for categorical variables and Kruskal-Wallis test for continuous variables. Remaining strains were not isolated or not available.

Appendix Table 2. Clinical and biological characteristics of 96 adults with Shiga toxin–associated hemolytic uremic syndrome by serogroup, France, 2009–2017*

Characteristic	Serogroup											
	O91	O157	O104	O26	O80	O106	O103	O128	O174	Other	Not isolated†	Not available‡
Total	12	10	8	4	4	4	3	3	3	16	17	12
Clinical features												
Sex												
M	5 (41.7)	4 (40.0)	2 (25.0)	3 (75.0)	4 (100.0)	2 (50.0)	1 (33.3)	1 (33.3)	0	5 (31.3)	5 (29.4)	3 (25.0)
F	7 (58.3)	6 (60.0)	6 (75.0)	1 (25.0)	0	2 (50.0)	2 (66.7)	2 (66.7)	3 (100.0)	11 (68.8)	12 (70.6)	9 (75.0)
Median age, y (IQR)	70.0 (67.5–74.75)	63.5 (47.0–71.25)	41.0 (36.25–47.5)	48.0 (37.25–59.25)	59.0 (52.5–62.75)	69.5 (61.75–76.75)	56.0 (54.0–56.5)	61.0 (46.5–80.0)	44.0 (38.0–65.5)	55.5 (46.0–62.5)	66.0 (54.0–73.0)	62.5 (55.75–72.5)
Median age-weighted	3.0 (2.0–4.25)	3.5 (0.25–4.0)	0.0	6.0 (5.5–6.5)	3.5 (2.25–4.5)	2.5 (1.75–3.75)	4.0 (2.5–5.0)	2.0 (1.0–5.5)	0.0 (0.0–4.0)	3.0 (1.0–5.0)	2.0 (1.0–5.0)	2.0 (1.0–3.25)
Charlson comorbidity index (IQR)												
≥1 Underlying condition	12 (100.0)	6 (60.0)	1 (12.5)	4 (100.0)	4 (100.0)	3 (75.0)	3 (100.0)	2 (66.7)	2 (66.7)	13 (81.3)	14 (82.4)	5 (41.7)
Immunodeficiency	3 (25.0)	1 (10.0)	0	4 (100.0)	2 (50.0)	0	2 (66.7)	1 (33.3)	0	6 (37.5)	7 (41.2)	1 (8.3)
Digestive disorder	3 (25.0)	0	0	1 (25.0)	2 (50.0)	1 (25.0)	2 (66.7)	1 (33.3)	1 (33.3)	7 (43.8)	8 (47.1)	3 (25.0)
Cardiovascular disease	8 (66.7)	4 (40.0)	0	2 (50.0)	2 (50.0)	3 (75.0)	3 (100.0)	0	1 (33.3)	12 (75.0)	8 (47.1)	5 (41.7)
Diarrhea	8 (66.7)	10 (100.0)	8 (100.0)	4 (100.0)	4 (100.0)	3 (75.0)	3 (100.0)	2 (66.7)	2 (66.7)	9 (56.3)	16 (94.1)	11 (91.7)
Bloody diarrhea	4 (33.3)	8 (80.0)	5 (62.5)	2 (50.0)	3 (75.0)	0	2 (66.7)	1 (33.3)	1 (33.3)	4 (25.0)	9 (52.9)	8 (66.7)
Isolation site												
Stool	12 (100.0)	10 (100.0)	8 (100.0)	4 (100.0)	4 (100.0)	3 (75.0)	3 (100.0)	2 (66.7)	2 (66.7)	14 (87.5)	17 (100.0)	11 (91.7)
Urine	1 (8.3)	0	1 (12.5)	0	0	1 (25.0)	0	0	1 (33.3)	2 (12.5)	0	1 (8.3)
Blood	0	0	0	0	1 (25.0)	0	1 (33.3)	1 (33.3)	0	0 (0.0)	0	1 (8.3)
stx genotypes												
stx1+/stx2–	0	0	0	3 (75.0)	0	0	3 (100.0)	1 (33.3)	0	5 (31.3)	0	NA
stx2+/stx1–	12 (100.0)	9 (90.0)	8 (100.0)	1 (25.0)	4 (100.0)	4 (100.0)	0	2 (66.7)	2 (66.7)	8 (50.0)	13 (76.5)	NA
stx1+/stx2+	0	1 (10.0)	0	0	0	0	0	0	1 (33.3)	3 (18.8)	4 (23.5)	NA
Events and outcomes												
Dialysis	11 (91.7)	6 (60.0)	2 (25.0)	3 (75.0)	3 (75.0)	3 (75.0)	3 (100.0)	2 (66.7)	2 (66.7)	7 (43.8)	12 (70.6)	7 (58.3)
Stroke, coma, or seizure	8 (66.7)	5 (50.0)	3 (37.5)	1 (25.0)	3 (75.0)	3 (75.0)	2 (66.7)	2 (66.7)	1 (33.3)	10 (62.5)	6 (35.3)	6 (50.0)
Any cardiac event	4 (33.3)	3 (30.0)	3 (37.5)	2 (50.0)	2 (50.0)	2 (50.0)	1 (33.3)	0	1 (33.3)	9 (56.3)	8 (47.1)	6 (50.0)
Death	4 (33.3)	1 (10.0)	0	3 (75.0)	2 (50.0)	1 (25.0)	1 (33.3)	0	0	3 (18.8)	3 (17.6)	1 (8.3)

*Values are no. (%), except as indicated. Major serogroups (i.e., O91, O157, O26, O80, O103, O111) comprised 62.1% of identified strains.

†Strain not identified.

‡Identification not performed.

Appendix Table 3. Characteristics of 19 adults who died of Shiga toxin–associated hemolytic uremic syndrome, France, 2009–2017*

Pt	Age, y	Sex	Underlying conditions	Serogroup	stx type	Minimum platelet count, 10 ⁹ cells/L	Packed red blood cells, nb†	Dialysis	Mechanical ventilation	Severe neurologic complications	Cardiac event	Tr	Days to death	Cause(s) of death
1	73	M	Colonic surgery, short bowel syndrome, hypogammaglobulinemia	O91	stx2	75	0	+	+	Coma, stroke	AHF	TPE	3	MOF
2	87	F	COPD, pulmonary hypertension, AH	O91	stx2	20	2	+				TPE	12	Sepsis
3	69	M	Mixed connective tissue disease	O91	stx2	19	4	+	+	Seizure, coma, stroke	Elevated troponin	TPE, ECZ	26	SNS
4	80	M	AH, COPD, ICM	O91	stx2	8	NA	+	+	Seizure, coma		TPE	27	Sepsis, VAP
5	73	F	Parkinson's disease with dementia	O157	stx2	15	4	+		Seizure, coma	Elevated troponin	BSC	12	SNS
6	57	M	Bone marrow transplantation (for refractory anemia with excess of blasts), digestive graft-versus-host disease	O80	stx2	3	+	+	+	Coma	HArr, AHF	TPE, ECZ	11	MOF, gastrointestinal bleeding
7	68	M	Bone marrow transplant (for acute myeloid leukemia), digestive graft-versus-host disease	O80	stx2	NA	0					TPE	62	Progressive graft-versus-host disease
8	78	F	Kidney transplantation (for ANCA-associated vasculitis), AH, ICM, renal insufficiency, depression, basocellular carcinoma	O26	stx1	63	2	+				TPE	19	MOF, sepsis, and gastrointestinal bleeding
9	53	M	Waldenström macroglobulinemia, COPD, cachexia	O26	stx2	77	0		+	Seizure, coma		BSC	66	SNS
10	20	M	Congenital immunodeficiency (immunodeficiency, centromeric region instability, facial anomalies syndrome; DNMT3b mutation), chronic colitis, autoimmune hepatitis, AH, renal insufficiency	O26	stx1	61	6	+			Elevated troponin, AHF	BSC	49	MOF
11	57	F	Liver transplant (for hepatitis B and C), CAFIB, AH, COPD, depression	O103	stx1	39	17	+		Stroke	HArr	TPE	152	SNS, persistent renal failure
12	79	F	Gastric cancer, stroke	O106	stx2	NA	0	+		Seizure, coma, stroke		TPE	3	SNS
13	65	F	AH, DM, ICM, CAFIB, primary sclerosing cholangitis	O177	stx2	29	+	+	+	Seizure, coma	Elevated troponin, AHF	TPE, ECZ	6	MOF
14	60	F	Cervical cancer, DM	Onew H27	stx2	32	3	+	+	Seizure, coma	AHF	TPE	44	MOF
15	68	M	Colon surgery, AH	NA	stx	42	2	+	+	Seizure, coma, stroke	Elevated troponin	TPE, ECZ	8	SNS
16	53	M	Chronic lymphocytic leukemia, chronic biliary disease, renal insufficiency, AH	NA	stx1	13	+	+	+	Seizure, coma, stroke	Elevated troponin	TPE, ECZ	22	SNS
17	80	F	Breast and endometrial cancer, AH, DM, ICM, CAFIB	NA	stx2	16	5	+			HArr, AHF	TPE	27	AHF caused by TPE
18	78	F	Parkinson's disease with severe dysautonomia	NA	stx1+2	32	9	+	+	Seizure, coma, stroke	Elevated troponin	TPE	72	SNS and VAP
19	73	F	Lung cancer, depression	NA	stx2	67	11	+	+	coma		TPE	101	Sepsis, VAP, cancer

*AH, arterial hypertension; AHF, acute heart failure; BSC, best standard of care; CAFIB, chronic atrial fibrillation; COPD, chronic obstructive pulmonary disease; DM, diabetes mellitus; ECZ, eculizumab; HArr, heart arrhythmia; ICM, ischemic cardiomyopathy; MOF, multiple organ failure; NA, not available; Pt, patient; SNS, severe neurologic sequelae; TPE, therapeutic plasma exchange; Tr, treatment; VAP, ventilation-associated pneumonia.

†Values reported where available.

Appendix Table 4. Clinical characteristics and outcomes of adults with Shiga toxin–associated hemolytic uremic syndrome by treatment, France, 2009–2017*

Characteristic	Best supportive care	TPE only	ECZ only	ECZ and TPE	ECZ, TPE, and immunoadsorption
Total	10 (100.0)	48 (100.0)	7 (100.0)	28 (100.0)	3 (100.0)
Medical history					
Median age, y (IQR)	64.5 (48.5–75.25)	62.0 (55.75–73.0)	38.0 (35.5–44.0)	57.5 (48.75–68.0)	64.0 (59.0–64.50)
Sex					
M	4 (40.0)	20 (41.7)	2 (28.6)	9 (32.1)	0
F	6 (60.0)	28 (58.3)	5 (71.4)	19 (67.9)	3 (100.0)
Median age-weighted Charlson Comorbidity Index (IQR)	3.5 (2.25–5.75)	3.0 (1.0–5.25)	0.0 (0.0–0.0)	2.0 (0.0–3.25)	2.0 (1.5–2.50)
≥1 Underlying condition					
Digestive disorder	3 (30.0)	20 (41.7)	0	6 (21.4)	0
Cardiovascular disease	8 (80.0)	27 (56.3)	1 (14.3)	12 (42.9)	0
Renal disease	5 (50.0)	7 (14.6)	0	3 (10.7)	0
Immunodeficiency	4 (40.0)	15 (31.3)	1 (14.3)	7 (25.0)	0
Autoimmune or inflammatory disease	4 (40.0)	17 (35.4)	1 (14.3)	8 (28.6)	1 (33.3)
Transplant	0	6 (12.5)	0	2 (7.1)	0
Neuropsychiatric disorder	3 (30.0)	9 (18.8)	2 (28.6)	4 (14.3)	0
Clinical features					
Diarrhea	7 (70.0)	40 (83.3)	7 (100.0)	23 (82.1)	3 (100.0)
Bloody diarrhea	3 (30.0)	24 (50.0)	4 (57.1)	14 (50.0)	2 (66.7)
Laboratory features					
Median platelet count, 10 ⁹ cells/L (IQR)†	77.5 (53.75–98.75)	51.0 (27.0–95.0)	241.0 (197.5–255.5)	55.0 (33.0–89.0)	36.0 (30.0–162.50)
Median minimum platelet count, 10 ⁹ cells/L (IQR)†	55.0 (35.0–73.0)	32.0 (18.0–50.0)	84.0 (46.0–86.5)	29.5 (21.75–46.75)	29.0 (22.0–33.0)
Median hemoglobin, g/dL (IQR)†	11.55 (9.67–12.25)	10.0 (9.05–11.7)	14.70 (12.65–16.08)	11.0 (8.5–12.4)	12.1 (10.35–13.15)
Renal manifestations					
Median serum creatinine, μmol/L (IQR)†	320.5 (142.0–385.0)	200.0 (157.0–328.0)	83.0 (66.75–217.75)	283.5 (160.5–452.75)	195.0 (130.0–218.00)
Stage 3 acute kidney injury‡	8 (80.0)	34 (70.8)	3 (42.9)	27 (96.4)	2 (66.7)
Required dialysis	4 (40.0)	30 (62.5)	2 (28.6)	24 (85.7)	1 (33.3)
Neurologic events					
Any neurologic events	4 (40.0)	39 (81.3)	3 (42.9)	24 (85.7)	3 (100.0)
Stroke, coma, or convulsions	4 (40.0)	23 (47.9)	2 (28.6)	18 (64.3)	3 (100.0)
Convulsions	2 (20.0)	12 (25.0)	1 (14.3)	12 (42.9)	3 (100.0)
Coma	2 (20.0)	16 (33.3)	0	15 (53.6)	3 (100.0)
Focal neurologic deficit	0	11 (22.9)	2 (28.6)	11 (39.3)	1 (33.3)
Abnormal brain imaging§	2 (66.7)	12 (41.4)	0	8 (40.0)	1 (33.3)
Required mechanical ventilation	1 (10.0)	16 (33.3)	0	14 (50.0)	3 (100.0)
Cardiac events					
Any cardiac event	7 (70.0)	18 (37.5)	1 (14.3)	12 (42.9)	3 (100.0)
High troponin¶	4 (100.0)	11 (57.9)	1 (33.3)	9 (60.0)	1 (50.0)
Serotype O104:H4	0	0	5 (71.4)	2 (7.1)	1 (33.3)
Isolation site					
Stool	9 (90.0)	45 (93.8)	7 (100.0)	26 (92.9)	3 (100.0)
Urine	1 (10.0)	3 (6.3)	1 (14.3)	2 (7.1)	0
Blood	0	3 (6.3)	0	1 (3.6)	0
>1 site	0	3 (6.3)	1 (14.3)	1 (3.6)	0
Other treatments					
Macrolides	1 (10.0)	9 (18.8)	5 (71.4)	9 (32.1)	2 (66.7)
Corticoids	2 (20.0)	11 (22.9)	0	2 (7.1)	1 (33.3)
Outcomes					
Deceased	3 (30.0)	11 (22.9)	0	5 (17.9)	0
Median duration of hospitalization, d (IQR)	19.0 (11.0–24.5)	30.0 (13.0–48.0)	16.0 (11.0–21.0)	38.0 (28.0–47.0)	40.0 (37.5–61.0)
Duration of dialysis >90 d#	1 (50.0)	2 (11.1)	0	1 (5.3)	0
Neurologic sequelae at the end of follow-up **	0	4 (36.4)	0	2 (18.2)	2 (66.7)

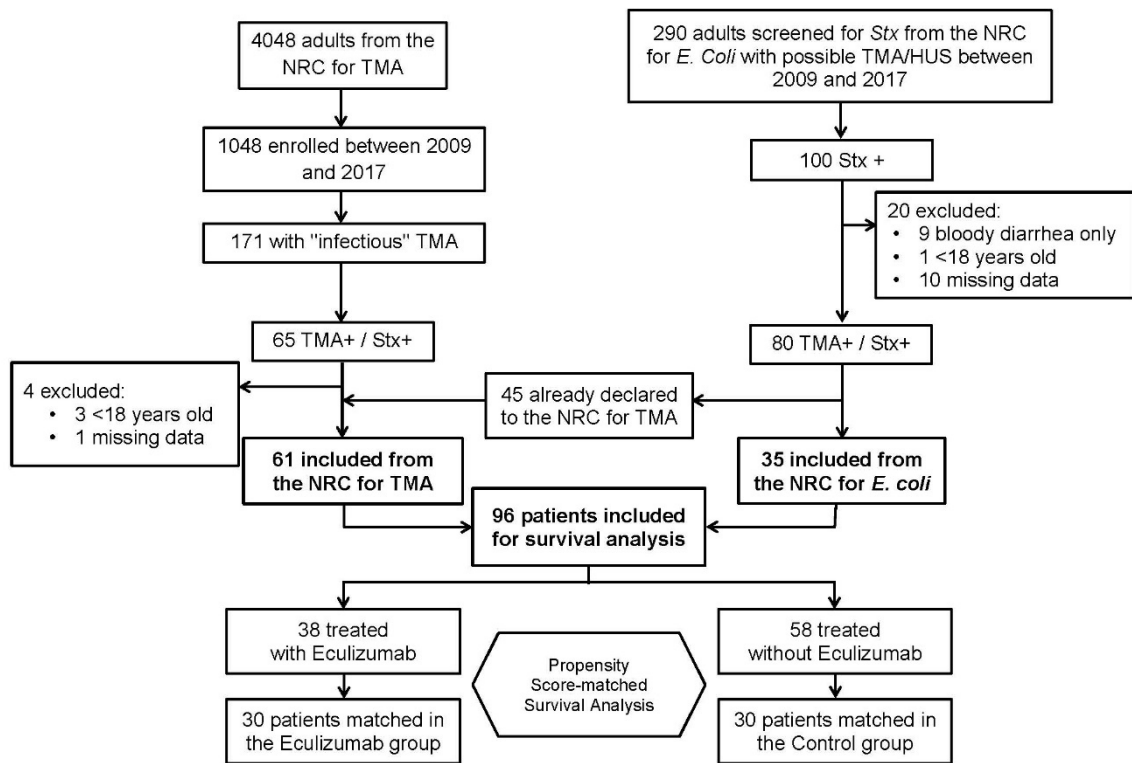
Characteristic	Best supportive care	TPE only	ECZ only	ECZ and TPE	ECZ, TPE, and immunoadsorption
*Values are no. (%), except as indicated. ECZ, eculizumab; TPE, therapeutic plasma exchange.					
†Samples taken at admission.					
‡According to Kidney Disease Improving Global Outcomes criteria (15).					
§Of 58 patients who had brain imaging, 3 were treated with the best standard of care, 29 with TPE only, 3 with ECZ only, 20 with ECZ and TPE, and 3 with ECZ, TPE, and immunoadsorption.					
¶Of 53 patients who had known blood troponin levels, 4 were treated with the best standard of care, 19 with TPE only, 3 with ECZ only, 15 with ECZ and TPE, and 2 with ECZ, TPE, and immunoadsorption.					
#Of 42 surviving dialysis patients with available data on the duration of dialysis, 2 were treated with the best standard of care, 18 with TPE only, 2 with ECZ only, 19 with ECZ and TPE, and 1 with ECZ, TPE, and immunoadsorption.					
**Of 25 surviving patients with available data who had stroke, coma or convulsions, 11 were treated with TPE only, 11 with ECZ and TPE, and 3 with ECZ, TPE, and immunoadsorption.					

Appendix Table 5. Comparison of Shiga toxin–associated hemolytic uremic syndrome patients treated with and without eculizumab, France, 2009–2017*

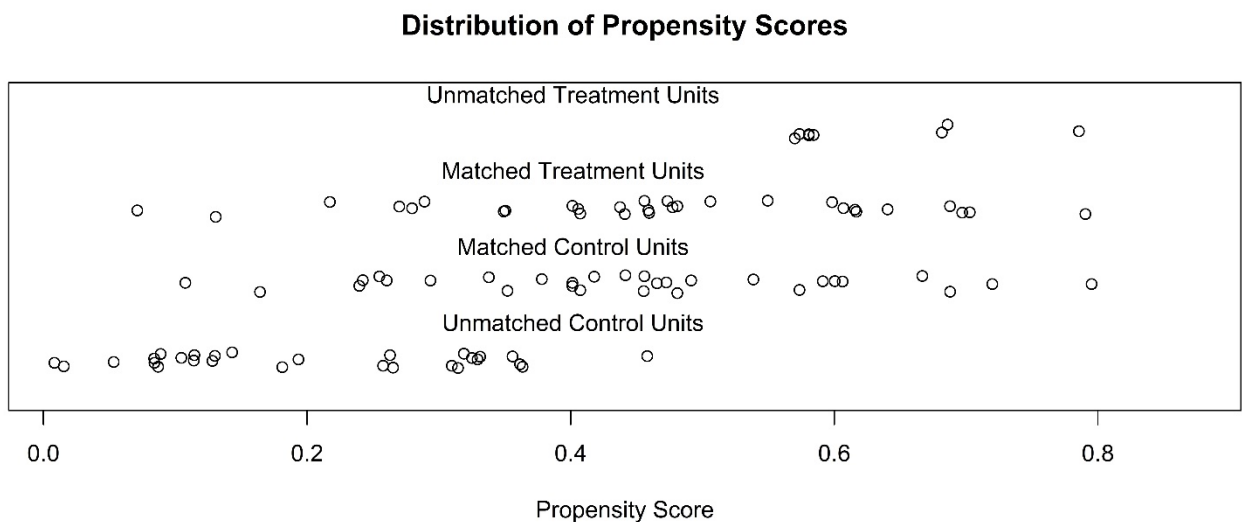
Characteristic	Eculizumab		p value†
	No	Yes	
Total	58	38	
Matched	30	30	
Unmatched	28	8	
Propensity score variables			
Median age, y (IQR)	59.50 (48.75–71.00)	54.50 (41.00–64.75)	0.19
Median age-weighted Charlson comorbidity index (IQR)	2.00 (0.00–3.75)	1.00 (0.00–3.75)	0.41
Immunodeficiency	8 (26.7)	8 (26.7)	1.00
Dialysis	21 (70.0)	19 (63.3)	0.79
Stroke, coma, or convulsions	17 (56.7)	16 (53.3)	1.00
Therapeutic plasma exchange	27 (90.0)	23 (76.7)	0.30
Other variables			
Death attributable to Shiga toxin–associated hemolytic uremic syndrome	7 (23.3)	4 (13.3)	0.51
Median follow-up time, d (IQR)	99.50 (31.75–181.25)	156.00 (56.00–227.50)	0.38

*Values are no. (%), except as indicated. Values are out of matched scores. Comparison subgroups were made after matching with propensity score (nearest neighbor method; caliper index equal to 0.3).

†p values for categorical variables determined by Fisher exact test; p values for continuous variables determined by Kruskal-Wallis test.



Appendix Figure 1. Design of study on adults with Shiga toxin–associated hemolytic uremic syndrome, France, 2009–2017. NRC, National Reference Centre; *E. coli*, *Escherichia coli*; HUS, hemolytic uremic syndrome; TMA, thrombotic microangiopathy.



Appendix Figure 2. Distribution of propensity scores of adults with Shiga toxin–associated hemolytic uremic syndrome, France, 2009–2017.