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nuttalli parasites are as virulent as *E. histolytica* parasites in animal models, it remains unclear whether they are as virulent in humans (4). We recommend that caretakers of NHPs be screened on a regular basis and be provided with appropriate medications if needed.

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Alaria alata Mesocercariae among Feral Cats and Badgers, Denmark

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To the Editor: The digenean trematode *Alaria alata* is considered an emerging zoonotic parasite in Europe because of increased findings in wild boars during *Trichinella* inspection. No human illness caused by *A. alata* mesocercariae (infective larvae) has been reported, but concern remains because the closely related North American species *A. americana* has caused illnesses among humans, including 1 death (1).

In Denmark, high prevalence of A. alata trematodes in final hosts has been shown (2), but limited data on potential paratenic hosts are available. Therefore, samples from 406 domestic pigs, 130 wild boars, 9 badgers, and 99 cats were collected by convenience sampling during October 2013-September 2014. We used pig and wild boar samples from multiple geographic areas of Denmark were leftover tissue samples from ongoing Trichinella spp. surveillance. Badgers had died naturally or were hit by vehicles (8 from Jutland, 1 from Zealand) and collected as part of a wildlife monitoring program. Cats (all from Zealand) were either feral (n = 92)or domesticated (n = 7) and had been euthanized as part of a national control program. Carcasses were necropsied in our laboratory; we collected 30 g of tissue samples according to Riehn et al. (3). All samples were analyzed by the modified A. alata mesocercariae migration technique (3). In brief, the sample was cut into ≈ 0.5 -cm edge pieces, wrapped in gauze, and suspended for 90 min by 2 wooden sticks in a conical glass with ≈ 300 mL of water (46°C–48°C). Approximately 15 mL of sediment was collected from the bottom of the glass by suction by using a glass pipette and examined by microscopy (magnification $\times 20$).

A. alata mesocercariae were isolated from 3 cats and 6 badgers (online Technical Appendix Table 1, http:// wwwnc.cdc.gov/EID/article/20/10/14-1817-Techapp1.pdf). All 3 cats were female (2 pregnant, 1 lactating); prevalence was significantly higher in pregnant or lactating females (3/12) than other intact females (0/24) (p = 0.031 by Fisher exact test). This finding might be related to increased exposure because an increase in predation by the cats during pregnancy and lactation to meet higher protein and energy demand. However, because *A. marcianae* mesocercariae can be

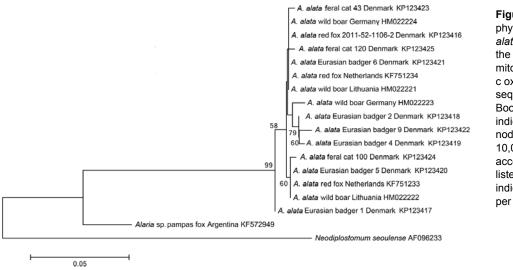


Figure. Neighbor-joining phylogenetic tree of *Alaria alata* isolates based on the analysis of partial mitochondrial cytochrome c oxidase subunit 1 gene sequences (332 bp). Bootstrap values are indicated to the left of the nodes and are based on 10,000 replicates. GenBank accession numbers are listed to the right. Scale bar indicates base substitutions per site.

transmitted through milk in cats (4), lactating females may also be predisposed to an increased chance for *A. alata* mesocercariae reaching their offspring. Examination of the intestines of all cats by sedimentation and counting technique (5) revealed no *A. alata* adults. Although *A. alata* adults have been found in cats in Uruguay (6), reports from Europe are lacking, and thus it is still uncertain whether cats can act as amphiparatenic or final hosts. Natural infection of cats with other *Alaria* spp. has been reported in the United States (7), indicating biologic differences among *Alaria* spp.

Zoonotic risk for *A. alata* infection through ingestion of cat meat is probably minimal in Europe but may be important in Asia and South America, where cats are occasionally consumed. Badgers are, however, sometimes consumed as game meat or road kill meat in Europe. In Russia, 10.6% of trichinellosis outbreaks during 1998–2002 were caused by consumption of badger meat (8). Thus, the zoonotic potential of infections in this animal, although a protected species, should not be ignored.

Negative findings in domestic pigs and wild boars in this study may reflect underestimation because those samples were below the recommended 30 g and often taken from sites that are not typically infected with mesocercariae (3). A follow-up study with better sampling strategy would be of value to determine the risk for *A. alata* transmission from domestic pigs and wild boars.

Identification of isolated mesocercariae was confirmed by PCR and sequencing of a fragment (332 bp) of the mitochondrial cytochrome c oxidase subunit 1 gene (*cox*1) (9). By neighbor-joining analysis (10), the consensus *cox*1 sequences were compared with the trematode *Neodiplostomum seoulense* (outgroup), 1 *A. alata* isolate from a Danish red fox, and all 7 *cox*1 sequences of *Alaria* spp. available in GenBank as of October 2014. (Sequences from this study have been deposited into GenBank under accession nos. KP123417–KP123422 [badgers] KP123423–KP123425 [feral cats].) The inferred phylogenetic tree (Figure) showed marked genetic variation among *A. alata* isolates from Denmark and other parts of Europe but no apparent separation of most *A. alata* isolates from Europe based on host species or country, except for that from badger 1 (on-line Technical Appendix Table 2). This animal originated from Northern Jutland, where host and parasite populations are geographically isolated by a large fjord separating the region from the rest of the country. The marked genetic variation within *cox*1 sequences suggests the usefulness of this marker, but additional genetic markers should be included in future studies to explore the genetic flow of *A. alata* within natural hosts.

In conclusion, *A. alata* mesocercariae seem to favorably infect pregnant or lactating cats, thereby increasing the chance of vertical transmission. Further, detection of *A. alata* infection in numerous badgers suggests potential high zoonotic risk associated with ingestion of such exotic meat. These results should, however, be interpreted with caution because of the small sample size and unknown efficacy of the modified *A. alata* mesocercariae migration technique.

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Human Infections with *Pseudoterranova cattani* Nematodes, Chile

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To the Editor: Anisakidosis is an emerging foodborne zoonosis caused by nematode larvae of the Anisakinae

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subfamily, which includes the genera Anisakis, Pseudoterranova, and Contracecum (1). In natural cycles, anisakid larvae are transmitted to marine mammals or piscivorous birds when they eat raw saltwater fish or squid. In the human incidental host, larvae attach to the mucosa of the gastrointestinal tract, causing clinical features ranging from asymptomatic carriage to severe abdominal pain with complications, such as gastric perforation (2). Microscopical diagnosis is hampered by the lack of distinguishing morphologic characteristics in larval stages (1). Recently, molecular genetic techniques have shown that the main species, Anisakis simplex and Pseudoterranova decipiens, are in fact species groups with distinct geographic and biologic characteristics (3,4). The *P. decipiens* complex consists of at least 6 sibling species (online Technical Appendix Table, http://wwwnc. cdc.gov/EID/article/21/10/14-1848-Techapp1.pdf). We report 4 human infections with P. cattani diagnosed during 2012-2014.

The case-patients were adults 22-59 years of age; 2 were female, and all lived in Santiago, Chile. Additional anamnestic and clinical data were available for 3 patients: all spontaneously regurgitated the parasites without having other gastrointestinal complaints. All 3 reported eating ceviche, a dish made of raw marine fish marinated in lemon juice. One patient reported a tingling sensation and coughs before the expulsion of a highly motile larva (Video, http:// wwwnc.cdc.gov/article/21/10/14-1848-V1.htm). This patient was awaiting oral surgery after a bicycle accident and had eaten the last raw fish dish 2 weeks previously. Initially, parasites were identified by morphologic criteria. Larvae were 20 mm long, were of whitish to reddish color, and had 3 anterior lips (online Technical Appendix Figure 1). Because of the presence of an anteriorly directed cecum (online Technical Appendix Figure 2), they were assigned to Pseudoterranova species.

For further molecular identification, DNA samples were extracted by using a DNeasy Blood and Tissue Kit (QIAGEN K.K., Tokyo, Japan). The rRNA gene containing 2 internal transcribed spacer (ITS) regions was amplified by PCR using primers NC5 and NC2, as previously described (5). PCR products were sequenced by using a BigDye Terminator Cycle Sequencing Kit (Applied Biosystems Inc., Foster City, CA, USA) on an automated sequencer (ABI3100, Applied Biosystems). Sequence similarities were determined by a BLAST search of DDBJ (http://blast. ddbj.nig.ac.jp/top-j.html). The GENETYX-WIN program version 7.0 (Software Development Co., Tokyo, Japan) facilitated sequence alignment and comparison. Within the 4 ITS sequences of amplicons obtained, all were 100% identical, and alignment with the other P. cattani sequence differed only in 1 nt. ITS sequences of 2 isolates are available in GenBank (accession nos. KF781284 and KF781285). All P. cattani sequences showed a previously described

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Technical Appendix

Technical Appendix Table 1. Information on domestic animals and wildlife examined for Alaria alata by using modified Alaria alata migration technique, Denmark October 2013-September 2014*

Origin	Examination period	amination period Sample type		Prevalence (%) (95% Cl)	
Z	Oct 2013–Mar 2014	Diaphragm or foreleg	260	0	
Z, F, J, ND	Oct 2013–Mar 2014	Diaphragm or foreleg	146	0	
Z	Oct 2013–Mar 2014	Diaphragm or foreleg	39	0	
J	Nov 2013–Jan 2014	Diaphragm, foreleg, or intercostal muscles	58	0	
ND	Oct 2013 – Mar 2014	Diaphragm or foreleg	33	0	
J	Feb–Sep 2014	Tissue sample mixture†	9	66.7 (29.9–92.5)	
Z	Mar–May 2014	Tissue sample mixture	99	3.0 (0.6–8.6)	
	Z Z, F, J, ND Z J ND	Z Oct 2013–Mar 2014 Z, F, J, ND Oct 2013–Mar 2014 Z Oct 2013–Mar 2014 J Nov 2013–Jan 2014 ND Oct 2013 – Mar 2014 J Feb–Sep 2014	ZOct 2013–Mar 2014Diaphragm or forelegZ, F, J, NDOct 2013–Mar 2014Diaphragm or forelegZOct 2013–Mar 2014Diaphragm or forelegJNov 2013–Jan 2014Diaphragm, foreleg, or intercostal musclesNDOct 2013 – Mar 2014Diaphragm or forelegJFeb–Sep 2014Tissue sample mixture†	ZOct 2013–Mar 2014Diaphragm or foreleg260Z, F, J, NDOct 2013–Mar 2014Diaphragm or foreleg146ZOct 2013–Mar 2014Diaphragm or foreleg39JNov 2013–Jan 2014Diaphragm, foreleg, or intercostal muscles58NDOct 2013 – Mar 2014Diaphragm or foreleg33JFeb–Sep 2014Tissue sample mixture†9	

*Z, Zealand; F, Funen; J, Jutland; ND, no data available. †30 g tissue sample mixture of diaphragm, tongue, and masticatory muscles;

Technical Appendix Table 2. Characteristics of Alaria alata mesocercariae-positive animal tissue samples identified by using modified Alaria alata migration technique, Denmark

				No. of mesocercariae		
Identity code	Species	Sex	Origin	per 30 g sample	GenBank accession number	
Cat 43	Cat (feral)	Female (pregnant)	Zealand	23	KP123423	
Cat 100	Cat (feral)	Female (pregnant)	Zealand	2	KP123424	
Cat 120	Cat (feral)	Female (lactating)	Zealand	12	KP123425	
Badger 1	Badger	Not recorded	Northern Jutland	1*	KP123417	
Badger 2	Badger	Female	Central Jutland	2	KP123418	
Badger 4	Badger	Female	Southern Jutland	1	KP123419	
Badger 5	Badger	Male	Central Jutland	2	KP123420	
Badger 6	Badger	Male	Central Jutland	8	KP123421	
Badger 9	Badger	Female	Zealand	2	KP123422	

*30 g tissue sample mixture of diaphragm, tongue, and masticatory muscles were used.