

References

1. Kamar N, Bendall R, Legrand-Abravanel F, Xia NS, Ijaz S, Izopet J, et al. Hepatitis E. *Lancet*. 2012;379:2477–88. [http://dx.doi.org/10.1016/S0140-6736\(11\)61849-7](http://dx.doi.org/10.1016/S0140-6736(11)61849-7)
2. Drexler JF, Seelen A, Corman VM, Fumie Tateno A, Cottontail V, Melim Zerbinati R, et al. Bats worldwide carry hepatitis E virus-related viruses that form a putative novel genus within the family *Hepeviridae*. *J Virol*. 2012;86:9134–47. <http://dx.doi.org/10.1128/JVI.00800-12>
3. He J, Binn LN, Tsarev SA, Hayes CG, Frean JA, Isaacson M, et al. Molecular characterization of a hepatitis E virus isolate from Namibia. *J Biomed Sci*. 2000;7:334–8. <http://dx.doi.org/10.1007/BF02253253>
4. van Cuyck H, Juge F, Roques P. Phylogenetic analysis of the first complete hepatitis E virus (HEV) genome from Africa. *FEMS Immunol Med Microbiol*. 2003;39:133–9. [http://dx.doi.org/10.1016/S0928-8244\(03\)00241-4](http://dx.doi.org/10.1016/S0928-8244(03)00241-4)
5. Nicand E, Armstrong GL, Enouf V, Guthmann JP, Guerin JP, Caron M, et al. Genetic heterogeneity of hepatitis E virus in Darfur, Sudan, and neighboring Chad. *J Med Virol*. 2005;77:519–21. <http://dx.doi.org/10.1002/jmv.20487>
6. Kaba M, Colson P, Musongela JP, Tshilolo L, Davoust B. Detection of hepatitis E virus of genotype 3 in a farm pig in Kinshasa (Democratic Republic of the Congo). *Infect Genet Evol*. 2010;10:154–7. <http://dx.doi.org/10.1016/j.meegid.2009.09.011>
7. John R, Plenge-Bönig A, Hess M, Ulrich RG, Reetz J, Schielke A. Detection of a novel hepatitis E-like virus in faeces of wild rats using a nested broad spectrum RT-PCR. *J Gen Virol*. 2010;91:750–8. <http://dx.doi.org/10.1099/vir.0.016584-0>
8. Mizuo H, Suzuki K, Takikawa Y, Sugai Y, Tokita H, Akahane Y, et al. Polyphyletic strains of hepatitis E virus are responsible for sporadic cases of acute hepatitis in Japan. *J Clin Microbiol*. 2002;40:3209–18. <http://dx.doi.org/10.1128/JCM.40.9.3209-3218.2002>
9. Pfefferle S, Frickmann H, Gabriel M, Schmitz N, Günther S, Schmidt-Chanasit J. Fatal course of an autochthonous hepatitis E virus infection in a patient with leukemia in Germany. *Infection*. 2012;40:451–4. <http://dx.doi.org/10.1007/s15010-011-0220-7>
10. Lu L, Li C, Hagedorn CH. Phylogenetic analysis of global hepatitis E virus sequences: genetic diversity, subtypes and zoonosis. *Rev Med Virol*. 2006;16:5–36. <http://dx.doi.org/10.1002/rmv.482>

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Novel Respiratory Syncytial Virus Subtype ON1 among Children, Cape Town, South Africa, 2012

To the Editor: Human respiratory syncytial virus (RSV) is a common cause of severe acute lower respiratory tract infection in young children, accounting for ≈160,000 deaths/year worldwide (1,2). As part of an RSV nosocomial transmission study, we detected RSV genotype ON1, which was identified during November 2010–February 2011 as a novel genotype in Ontario, Canada, in samples from children in a tertiary pediatric hospital in Cape Town, South Africa during 2012. The genotype described in Canada was characterized by a 72-nt sequence duplication within the second variable domain of the envelope glycoprotein. The 72-nt duplication within the second variable domain in ON1 was the largest sequence duplication described in this virus (3).

RSV is divided into 2 genetically distinct groups, RSV A and B, based on the viral envelope glycoprotein nucleotide sequences (4). Sequence variability in the C-terminal variable domain of the glycoprotein gene is commonly used to determine RSV phylogeny (3,5). To date, 11 RSV A (ON1, GA1–GA7, SAA1, NA1, and NA2) and 17 RSV B (GB1–GB4, SAB1–SAB3, and BA1–BA10) genotypes have been identified (3,6).

As part of the aforementioned molecular epidemiology study surveying RSV infection in a pediatric hospital, (University of Cape Town research ethics study no. 305/2012), we sequenced the RSV glycoprotein second variable domain of nucleic acid extracts derived from RSV-positive respiratory secretion samples from 160 young children hospitalized for treatment of respiratory tract infections. The techniques used have been described (7). During January–April, in an area where NA1 was the dominant circulating RSV genotype, 119 (74%) of 160 RSV isolates were RSV A. We noted the presence, albeit at a low incidence, of the novel ON1 genotype cluster (8 viral isolates) (online Technical Appendix Figure, wwwnc.cdc.gov/EID/pdfs/12-1465-Techapp.pdf) in specimens collected during February–April.

Children in the RSV ON1-infected cohort were brought to health care facilities during February 24–April 25, 2012 (Figure and online Technical Appendix Figure), where they received a diagnosis of bronchiolitis or bronchiopneumonia (online Technical Appendix Table). With the exception of 1 patient, child 8, who had been hospitalized before onset of this illness, all ON1 isolates were community acquired. Seven of the 8 ON1 isolates were obtained from infants <4 months of age (median 7 weeks), who were younger than the 152 children who were not infected with the ON1 genotype (median age 3.5 months). The RSV ON1-infected children lived within a 2.5-km radius of one another (online Technical Appendix Table). The children who were not infected with RSV ON1 lived in a much wider geographic area; >90% lived within an 18-km radius of one another. These spatial associations with disease prevalence suggested that the ON1-infected children represented a localized cluster of transmission.

None of the children were infected with HIV, although 3 had antenatal

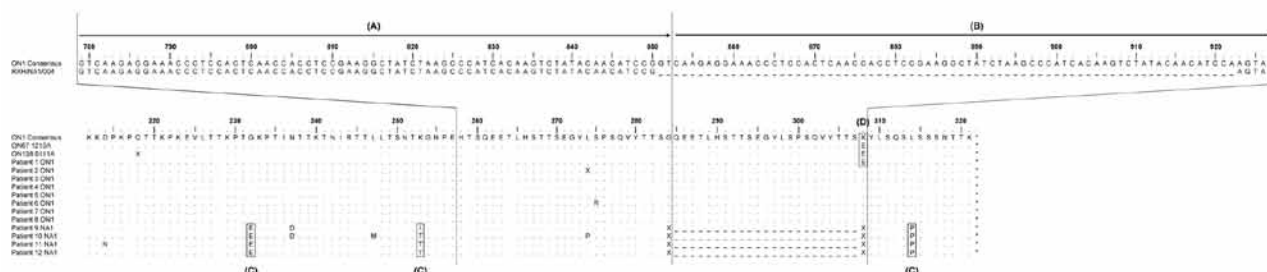


Figure. Alignment of deduced amino acid sequences for ON1 isolates from South Africa (patients 1–8, accession nos. JX885730–JX885737) and Canada (ON67 and ON138) with NA1 isolates (patients 9–12) from South Africa. A) Variable domain sequence copied. B) Duplicated sequence inserted into variable domain. C) Characteristic amino acid substitutions that distinguish ON1 from NA1. D) Amino acid substitution (E308K) (position 284 before insertion) that distinguishes between most ON1 isolates from South Africa (patients 2–8) from those from Canada (ON67 and ON138) ON1.

exposure to HIV. Co-infection with adenovirus and rhinovirus was noted in 3 of the patients. Although 3 of the patients were hospitalized for a prolonged period and required ventilatory support, the severity and outcome of the RSV ON1 infections were similar to RSV infections caused by other genotypes among children of the same age.

Sequence analyses revealed that ON1 isolates identified in South Africa are essentially identical to those isolated in Canada, possessing characteristic amino acid substitutions at positions E232G, T253K, and P314L that distinguish the genotype from circulating NA1 genotypes (3). However, 7 of 8 ON1 isolates from South Africa possess a unique E308K (position 284 before insertion) amino acid change at the 3' border of the duplicated gene segment not present in the ON1 isolates identified in Canada (Figure). The conservation of the E308K mutation within ~90% of the isolates from South Africa that we studied suggests a possible functional role for the positively charged lysine residue.

The capacity of the RSV glycoprotein to accommodate large insertions and remain functional was first demonstrated with the RSV B, BA genotype (Buenos Aires, Argentina 1999). This genotype contains a 60-nt duplication in the second variable

domain, which, similar to ON1, did not cause serious clinical outcomes (6,8–10). Longitudinal analyses during 12 epidemic seasons (1996–97 through 2007–08) of international RSV subtype distribution revealed that since its initial detection in 1999, BA prevalence has gradually increased to become the dominant RSV group B virus genotype in circulation (10). Because RSV A has traditionally been the dominant RSV type in circulation, if the large insertion in ON1 confers similar selection advantage as seen in BA, the potential dominance of a single ON1 genotype within this group might promote bias on RSV type distribution toward RSV A.

The novel ON1 genotype was first described in Ontario, Canada (3). Our subsequent findings in South Africa suggest extensive distribution of this genotype, which was assumed to have arisen before winter 2010–11 (3). To understand whether ON1 in South Africa occurred as a result of importation or natural evolution within locally circulating NA1 genotypes, further research is required.

Acknowledgments

We thank the staff of the Groote Schuur National Health laboratory Service, who provided RSV- positive archived cDNA samples. We thank Heidi Smuts for critical review of the manuscript and research guidance.

Study funding was provided by the University of Cape Town.

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DOI: <http://dx.doi.org/10.3201/eid1904.121465>

References

- Selwyn BJ. The epidemiology of acute respiratory tract infection in young children: comparison of findings from several developing countries. Coordinated Data Group of BOSTID Researchers. *Rev Infect Dis.* 1990;12(Suppl 8):S870–88. http://dx.doi.org/10.1093/clinids/12.Supplement_S870
- World Health Organization (WHO). Initiative for Vaccine Research (IVR). Respiratory syncytial virus and parainfluenza virus. Disease burden. Geneva: The Organization; 2009.
- Eshaghi A, Duvvuri VR, Lai R, Nadarajah JT, Li A, Patel SN, et al. Genetic variability of human respiratory syncytial virus A strains circulating in Ontario: a novel genotype with a 72 nucleotide G gene duplication. *PLoS ONE.* 2012;7:e32807. <http://dx.doi.org/10.1371/journal.pone.0032807>

4. Johnson PR, Spriggs MK, Olmsted RA, Collins PL. The G glycoprotein of human respiratory syncytial viruses of subgroups A and B: extensive sequence divergence between antigenically related proteins. *Proc Natl Acad Sci U S A*. 1987;84:5625–9. <http://dx.doi.org/10.1073/pnas.84.16.5625>
5. Cane PA. Molecular epidemiology of respiratory syncytial virus. *Rev Med Virol*. 2001;11:103–16. <http://dx.doi.org/10.1002/rmv.305>
6. Daplat IC, Shobugawa Y, Sano Y, Saito R, Sasaki A, Suzuki Y, et al. New genotypes within respiratory syncytial virus group B genotype BA in Niigata, Japan. *J Clin Microbiol*. 2010;48:3423–7. <http://dx.doi.org/10.1128/JCM.00646-10>
7. Peret TC, Hall CB, Schnabel KC, Golub JA, Anderson LJ. Circulation patterns of genetically distinct group A and B strains of human respiratory syncytial virus in a community. *J Gen Virol*. 1998;79:2221–9.
8. Trento A, Galiano M, Videla C, Carballal G, Garcia-Barreno B, Melero JA, et al. Major changes in the G protein of human respiratory syncytial virus isolates introduced by a duplication of 60 nucleotides. *J Gen Virol*. 2003;84:3115–20. <http://dx.doi.org/10.1099/vir.0.19357-0>
9. Trento A, Viegas M, Galiano M, Videla C, Carballal G, Mistchenko AS, et al. Natural history of human respiratory syncytial virus inferred from phylogenetic analysis of the attachment (G) glycoprotein with a 60-nucleotide duplication. *J Virol*. 2006;80:975–84. <http://dx.doi.org/10.1128/JVI.80.2.975-984.2006>
10. Trento A, Casas I, Calderón A, Garcia-Garcia ML, Calvo C, Perez-Breña P, et al. Ten years of global evolution of the human respiratory syncytial virus BA genotype with a 60-nucleotide duplication in the G protein gene. *J Virol*. 2010;84:7500–12. <http://dx.doi.org/10.1128/JVI.00345-10>

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Henipaviruses and Fruit Bats, Papua New Guinea

To the Editor: In 2010, detection of henipavirus (Hendra or Nipah virus) and rubulavirus (Tioman or Menangle virus) antibodies in fruit bats in Papua New Guinea (PNG) was reported (1). To explore changes in henipavirus dynamics in fruit bats, we compare and contrast this finding with serologic findings from 10 years earlier (2; H. Field et al., unpub. data).

In these earlier studies, blood samples were collected from 182 wild-caught fruit bats of mixed species, age, and sex from 3 locations in PNG: Madang (1996), New Britain (1997), and Lae (1999) (2; H. Field et al., unpub. data) (Figure). The 20 samples from Madang were collected as blood spots on filter paper and forwarded to the (then) Department of Primary Industries Animal Research Institute in Brisbane, Australia, where they were eluted and screened by ELISA for antibodies against Hendra virus (3). Serum from 59 samples

from New Britain and 103 from Lae were forwarded to the Commonwealth Scientific and Industrial Research Organisation's Australian Animal Health Laboratory in Geelong, Australia. Samples from New Britain were screened for antibodies against Hendra virus by virus neutralization test (VNT) (3). Positive samples were subsequently screened by VNT for antibodies against Nipah virus. Samples from Lae were screened by VNT for Hendra, Nipah, and Menangle viruses (3). A reciprocal VNT titer of >5 was considered indicative of antibodies.

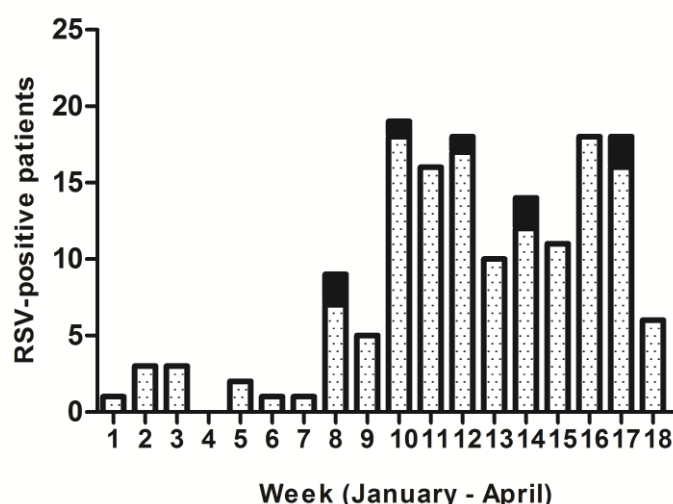
Of the 20 samples from Madang, 2 (10%) reacted in the Hendra virus ELISA. Of the 147 samples from New Britain and Lae that yielded definitive VNT results, 11 (7.5%) yielded neutralizing antibodies to Hendra virus and 5 (3.4%) to Nipah virus. All samples with antibodies against Nipah virus also had antibodies against Hendra virus; titers against Hendra virus were greater (4 samples) or equivalent (1 sample) to those against Nipah virus. Reciprocal titers against Hendra virus were 5–160 (median 10) and against Nipah



Figure. Sampling locations and henipavirus antibody prevalence, Papua New Guinea 1996–1999, among 182 wild-caught fruit bats from Madang (1996), New Britain (1997), and Lae (1999), Papua New Guinea. HeV, Hendra virus; NiV, Nipah virus; MenV, Menangle virus.

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Technical Appendix



Technical Appendix Figure. Respiratory syncytial virus (RSV) cases among hospitalized children, Cape Town, South Africa. Stippled bar sections indicate non-ON1 genotype infections. Black bar sections indicate ON1 genotype infections.

Technical Appendix Table. Epidemiologic observations for patients infected with respiratory syncytial virus ON1 in a hospital pediatric unit, Cape Town, South Africa*

Pt no.	Sex/age, wk	LOS, d	HIV exposure/status	ICU/HDU care	Respiratory support	Discharge diagnosis	Concurrent conditions	Co-infection
1	F/15	2	U/-	No	No	Bronchial pneumonia	Preterm birth	None
2	M/1.5	1	U/-	No	No	Bronchiolitis	Neonatal jaundice	None
3	F/4	7	U/-	HDU	CPAP	Bronchiolitis	None	None
4	F/3	1	E/-	No	No	Bronchiolitis	None	Adenovirus
5	F/10	2	E/-	No	No	Lobar pneumonia	Neonatal sepsis	Rhinovirus
6	F/8	38	U/-	ICU	IPPV	Bronchiopneumonia	Preterm birth	None
7	F/6.5	12	E/-	HDU	CPAP	Bronchiopneumonia	Preterm birth	None
8	M/60	1	U/-	No	No	Bronchiolitis obliterans	Recurrent LRTI after adenovirus infection	Rhinovirus

*Pt, patient; LOS, length of stay in hospital; ICU, intensive care unit; HDU, high dependency unit; U, unexposed; -, negative; E, exposed; LRTI, lower respiratory tract infection; CPAP, continuous positive airway pressure; IPPV, intermittent positive pressure ventilation.