

9. Plyusnin I, Kant R, Jääskeläinen AJ, Sironen T, Holm L, Vapalahti O, et al. Novel NGS pipeline for virus discovery from a wide spectrum of hosts and sample types. *Virus Evol.* 2020;6:veaa091. <https://doi.org/10.1093/ve/veaa091>
10. Laenen L, Vergote V, Calisher CH, Klempa B, Klingström J, Kuhn JH, et al. Hantaviridae: current classification and future perspectives. *Viruses.* 2019;11:788. <https://doi.org/10.3390/v11090788>

Address for correspondence: Nathaniel Mull, Shawnee State University, 940 Second St, Portsmouth, OH 45662, USA; email: nmull@shawnee.edu

Zoonotic Marine Nematode Infection of Fish Products in Landlocked Country, Slovakia

Mikuláš Oros, Daniel Barčák, Daniela Antolová, Miroslava Fecková, Tomáš Scholz

Author affiliations: Institute of Parasitology, Slovak Academy of Sciences, Košice, Slovakia (M. Oros, D. Barčák, D. Antolová, M. Fecková); Institute of Parasitology, Biology Centre of the Czech Academy of Sciences, České Budějovice, Czech Republic (T. Scholz); University of South Bohemia Faculty of Science, České Budějovice (T. Scholz)

DOI: <https://doi.org/10.3201/eid2912.230674>

Fish products in Slovakia have been heavily infected with *Anisakis* spp. larvae, which causes human anisakiasis. We found larvae in all tested samples of frozen Atlantic herring. Anisakid allergen t-Ani s7 testing revealed 2 positive cases in humans, signaling need for health authorities to closely monitor zoonotic marine parasites, even in inland areas.

Food safety is an unquestioned global public health imperative. Strict controls on food before release to markets are intended to prevent disease caused by agents of infectious diseases, including parasitoses such as human anisakiasis. Anisakiasis, an emerging zoonosis, is caused mainly by marine nematodes of the genus *Anisakis*. They mature in dolphins and whales, but their third-stage larvae, which reside in the flesh, gonads, and body cavities of marine fish,

can infect humans who consume raw or undercooked fish (1). Most human cases are reported in Japan, Spain, and South Korea (1).

Zoonotic diseases caused by marine parasites have been largely confined to coastal regions, but surveillance by health authorities in landlocked countries is lacking. However, global trade and the increasing popularity of raw fish consumption have contributed to emergence of that disease. We provide data on extensive infection of fish products with *Anisakis* larvae in Slovakia, a landlocked country in central Europe. We also report seropositive cases in a group of volunteers regularly eating fish products. We conducted this study in accordance with ethics standards in the 2013 revision of the Declaration of Helsinki of 1975. It was approved by the ethics committee of the Institute of Parasitology, Slovak Academy of Sciences (EC/01/2018; December 14, 2018).

We examined 100 frozen Atlantic herring (*Clupea harengus*) provided by a fish product supplier and 18 packages of ready-to-eat pickled herring from local supermarkets for anisakid larvae. We found 4,163 larvae in frozen Atlantic herring at an intensity of infection of 2–368 (mean 42) larvae/fish (Figure, panels A–C, E–H). Although we found most larvae in the abdominal cavity, we also found them in the muscles of 1/3 and the gonads of 1/5 of fish we examined. Although all larvae were dead, even dead parasites or their residues in contaminated fish products can cause allergic reaction in sensitized persons (2,3). In addition, we found anisakids in 1/3 of ready-to-eat pickled herring (1–9 larvae/fish) (Figure, panel D).

We morphologically identified larvae as *Anisakis* spp. and used several larvae for genotyping based on the ~800 bp-long internal transcribed spacer region. Sequences of all isolates were identical to those of *A. simplex* sensu stricto (Appendix, <https://wwwnc.cdc.gov/EID/article/29/12/23-0674-App1.pdf>), considered the predominant agent of human anisakiasis (4).

We used a Trisakis 170, *A. simplex* IgE-ELISA kit (Laboratorio de Parasitología, Facultad de Farmacia, Universidad de Santiago de Compostela, Santiago de Compostela, Spain) to test IgE sensitization to *Anisakis* spp. in a human population. The kit detects IgE to the recombinant allergens r-Ani s 1 and t-Ani s 7 in human serum, which we collected in 2020 from 91 volunteers who regularly ate fish products. The allergen tests are highly specific (up to 100%), as deduced also from primary amino acid sequences of both allergens that have no significant homologies with other allergens to which humans are known to be sensitized. Sensitivity reaches 61.1% for Ani s 1 and 93.9% for Ani s 7; many researchers consider serum truly

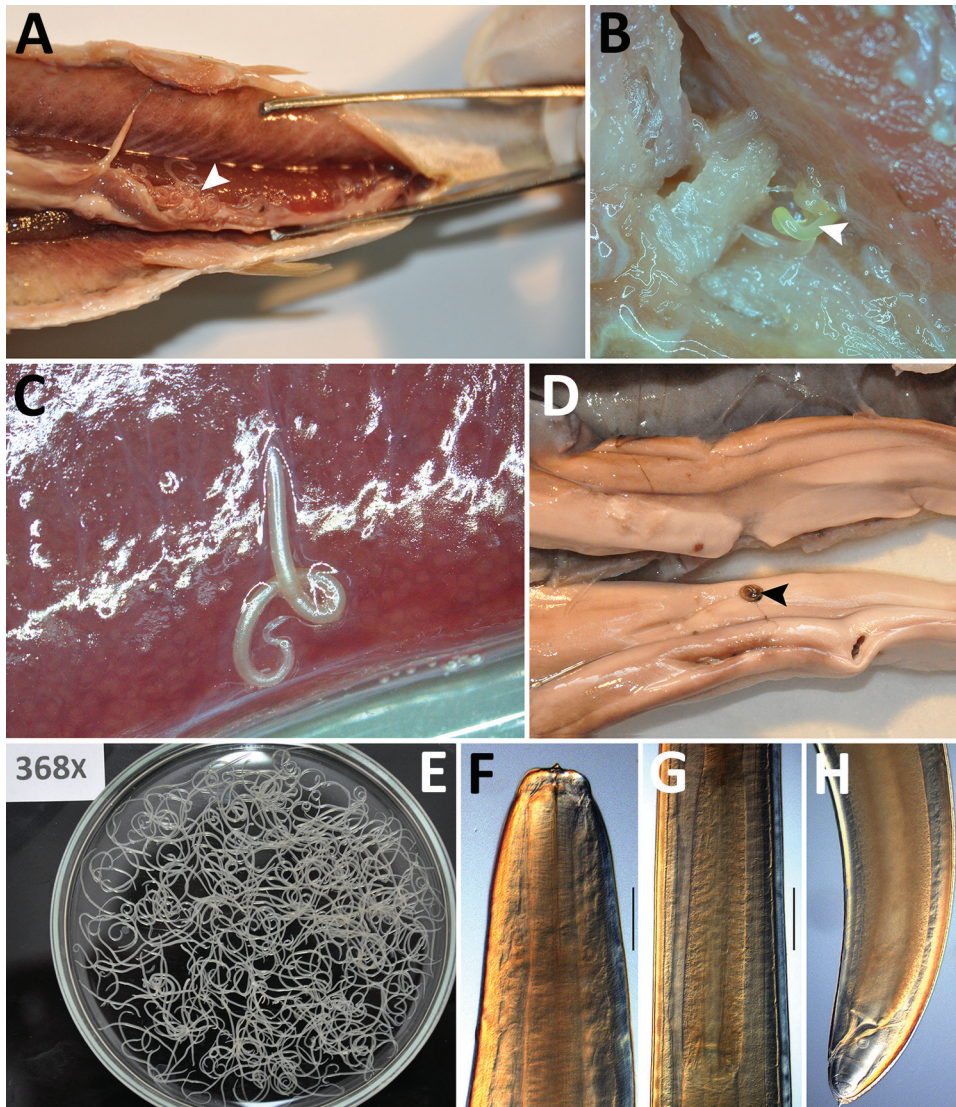


Figure. Anisakid larvae found in frozen (A–C, E–H) and ready-to-eat (D) Atlantic herrings (*Clupea harengus*), Slovakia. A) Cluster of larvae on the viscera (arrowhead). B) Larva in dorsal muscle (arrowhead). C) Larva partly embedded in the hard roe. D) Larva on the surface of the soft roe (arrowhead). E) All larvae isolated from single fish. Original magnification as indicated. F–H) Light microscope photographs of head part (F), ventriculus (G), and tail part (H) of an *Anisakis simplex* sensu stricto nematode. Scale bars indicate 200 µm.

positive only when testing positive to Ani s 1 or Ani s 7 (5,6). Moreover, Ani s 7 and, probably, Ani s 1 are excretory-secretory allergens that are recognized by the host immune system only in the course of an *Anisakis* infection (5). Of 91 serum samples collected in 2020, sensitization to *Anisakis* allergen t-Ani s 7 was detected in 2 (2.2%) samples. The positive case-patients, both women, had no clinical symptoms of sensitization to *Anisakis* but had experienced allergy symptoms on several occasions in the past.

Any report of sensitization to *Anisakis* in Slovakia is unusual in that it is a landlocked country. Given the low level of raw fish consumption in Slovakia, the 2.2% rate of detected positivity was relatively high. In a similar study in Norway, 0/993 blood donors and 1/414 (0.2%) patients with high IgE levels tested positive for anisakiasis (6). In a study in Croatia in

which the same ELISA method was used as in our study, 3.5% positivity was found in persons living on islands, but only 1.5% in persons in urban areas on the coast (7). A 15.4% positivity rate was detected in an adult population in Spain (8), whereas seroprevalence was only 0.4% in blood donors in Galicia, in northwestern Spain (9). In the future, IgE for Ani s 1, Ani s 4, Ani s 5, and Ani s 9, heat-resistant allergens that cause most clinical episodes of the allergic form of anisakiasis, should be evaluated.

Monitoring fish products intended for human consumption for parasites currently receives insufficient attention. Although all parasites we found were dead, frequent presence of *Anisakis* spp. in herring poses a potential risk to sensitive persons who might suffer a hyperallergic reaction. In addition, some *Anisakis* larvae can survive freezing (10), so risk of infection

remains even in fish products frozen for a short time. In conclusion, the results of our study signal the need for health authorities to closely monitor marine parasites with zoonotic potential, even in inland areas.

Acknowledgements

We thank fish product supplier who generously provided frozen herrings for parasitologic examination.

This study was supported by the Slovak Research and Development Agency (APVV SK-CZ-RD-21-0078 to M.O.) and Ministry of Education, Youth and Sports of the Czech Republic (LUASK22045 to T.S.).

About the Author

Dr. Oros is a senior researcher at the Institute of Parasitology, Slovak Academy of Sciences. His research focuses on the integrative taxonomy, systematics, and phylogeny of multicellular parasites of bony fishes, including etiological agents of fish-borne human diseases.

References

- Shamsi S, Barton DP. A critical review of anisakidosis cases occurring globally. *Parasitol Res.* 2023;122:1733–45. <https://doi.org/10.1007/s00436-023-07881-9>
- Nieuwenhuizen NE, Lopata AL. *Anisakis* – a food-borne parasite that triggers allergic host defences. *Int J Parasitol.* 2013;43:1047–57. <https://doi.org/10.1016/j.ijpara.2013.08.001>
- Bao M, Pierce GJ, Strachan JC, Pascual S, González-Muñoz M, Levsen A. Human health, legislative and socioeconomic issues caused by the fish-borne zoonotic parasite *Anisakis*: challenges in risk assessment. *Trends Food Sci Technol.* 2019;86:298–310. <https://doi.org/10.1016/j.tifs.2019.02.013>
- Mattiucci S, Cipriani P, Levsen A, Paoletti M, Nascetti G. Molecular epidemiology of *Anisakis* and anisakiasis: an ecological and evolutionary road map. *Adv Parasitol.* 2018;99:93–263. <https://doi.org/10.1016/bs.apar.2017.12.001>
- Anadón AM, Rodríguez E, Gárate MT, Cuéllar G, Romarís F, Chivato T, et al. Diagnosing human anisakiasis: recombinant Ani s 1 and Ani s 7 allergens versus the UniCAP 100 fluorescence enzyme immunoassay. *Clin Vaccine Immunol.* 2010;17:496–502
- Lin AH, Nepstad I, Florvaag E, Egaas E, Van Do T. An extended study of seroprevalence of anti-*Anisakis* simplex IgE antibodies in Norwegian blood donors. *Scand J Immunol.* 2014;79:61–7. <https://doi.org/10.1111/sji.12130>
- Mladineo I, Poljak V, Martínez-Sernández V, Ubeira FM. Anti-*Anisakis* IgE seroprevalence in healthy Croatian coastal population and associated risk factors. *PLoS Negl Trop Dis.* 2014;8:e2673. <https://doi.org/10.1371/journal.pntd.0002673>
- Ubeira FM, Anadón AM, Salgado A, Carjaval A, Ortega S, Aguirre C, et al. Synergism between prior *Anisakis simplex* infections and intake of NSAIDs, on the risk of upper digestive bleeding: a case-control study. *PLoS Negl Trop Dis.* 2011;5:e1214.
- Valiñas B, Lorenzo S, Eiras A, Figueiras A, Sanmartín ML, Ubeira FM. Prevalence of and risk factors for IgE sensitization to *Anisakis simplex* in a Spanish population. *Allergy.* 2001;56:667–71.
- Wharton DA, Alders O. The response of *Anisakis* larvae to freezing. *J Helminthol.* 2002;76:363–8. <https://doi.org/10.1079/JOH2002149>

Address for correspondence: Mikuláš Oros, Institute of Parasitology, Slovak Academy of Sciences, Hlinkova 3, 040 01, Košice, Slovak Republic; email: oros@saske.sk

Detection of African Swine Fever Virus from Wild Boar, Singapore, 2023

Eileen Y. Koh, Adrian K.S. Tan, Darren Yeo, Clara Lau, Li Ying Tan, Oi Wing Ng, Jasmine Ong, Stacy Chong, Steffie Toh, Jing Chen, Wai Kwan Wong, Brian Z.Y. Tan, Christine He-Lee, Zhan Pei Heng, Ian Liang, Charlene Judith Fernandez, Siow Foong Chang, Kenneth B.H. Er

Author affiliation: National Parks Board, Singapore

DOI: <https://doi.org/10.3201/eid2912.230966>

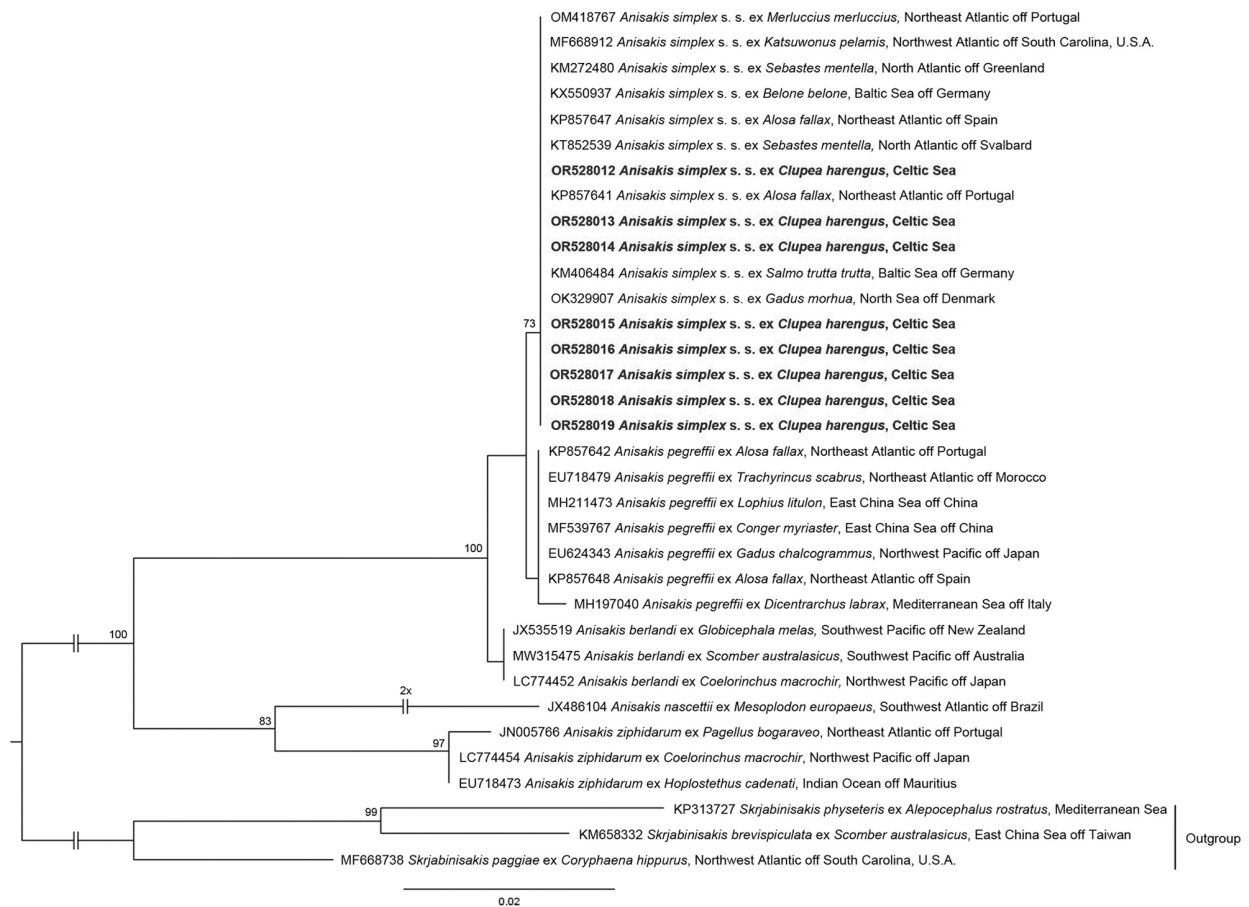
We detected African swine fever virus (ASFV) from a wild boar in Singapore. In ≤ 72 hours, we confirmed and reported ASFV p72 genotype II, CD2v serogroup 8, and IGR-II variant by using a combination of real-time PCR and whole-genome sequencing. Continued biosurveillance will be needed to monitor ASFV in Singapore.

African swine fever (ASF), a nonzoonotic, World Organisation for Animal Health (WOAH) notifiable disease, is a devastating hemorrhagic infectious disease of domestic (*Sus domesticus*) and wild (*Sus scrofa*) swine populations (1). ASF was identified in Kenya in 1921 and subsequently spread through >50 countries (1). ASF is known to be spread by *Ornithodoros* soft ticks, vectors of ASF virus (ASFV), as well as contact with infected swine or contaminated vehicles or equipment and by consuming of infected carcasses (1). ASF often is associated with high illness and death rates in suids. However, wild suid species in Africa, such as warthogs (*Phacochoerus aethiopicus*), act as reservoir hosts for the virus but reportedly

EID cannot ensure accessibility for supplementary materials supplied by authors. Readers who have difficulty accessing supplementary content should contact the authors for assistance.

Zoonotic Marine Nematode Infection of Fish Products in Landlocked Country, Slovakia

Appendix



Appendix Figure. Phylogeny inferred from ITS rDNA dataset, maximum likelihood estimation by ultrafast bootstrap approximation with 1,000 replicates in IQ-TREE, version 2.0.5. Bootstrap supports over 70 shown. Newly generated sequences of *A. simplex* from fish products in Slovakia are bolded. Scale bar indicates number of substitutions per site.