Seroprevalence of Zika and Dengue Virus Antibodies among Migrant Workers, Taiwan, 2017

Guey Chuen Perng, Tzu-Chuan Ho, Hsin-I Shih, Chia-Hua Lee, Pei-Wen Huang, Chih-Huan Chung, Nai-Ying Ko, Wen-Chien Ko, Yu-Wen Chien

A serosurvey of 600 workers newly arrived in Taiwan from 4 Southeast Asia countries showed that 18 (3%) were positive for Zika virus IgM; 6 (1%) fulfilled the World Health Organization criteria for laboratory-confirmed recent Zika virus infection. The incidence of Zika virus infection in Southeast Asia might be underestimated.

Approximately 690,000 migrant workers live in Taiwan; most are from 4 countries in Southeast Asia: Indonesia, the Philippines, Thailand, and Vietnam. Although these migrants are valuable to the workforce, they may also bring the risk of spreading mosquitoborne diseases. Most imported cases of Zika virus and dengue virus (DENV) infections in Taiwan come from Southeast Asia. Although screening for fever at international airports was implemented in Taiwan in 2003 (*1*), most Zika virus and DENV infections are inapparent and cannot be detected.

For assessing the true disease burden of flavivirus infections, seroprevalence studies are effective. Recently, several serosurveys of Zika virus infections were conducted in Africa, Oceania, Latin America, and the Caribbean (*2*). Although small outbreaks of Zika virus infection have been reported in Singapore, Vietnam, and Thailand (*3–5*), seroprevalence in Southeast Asia countries remains largely unknown. To estimate the incidence of Zika virus and DENV infections in Southeast Asia and to evaluate the risk of importing these viruses into Taiwan, we investigated seroprevalence of IgM and IgG against these viruses among newly arrived migrant workers in Taiwan.

Author affiliations: National Cheng Kung University College of Medicine, Tainan, Taiwan (G.C. Perng, T.-C. Ho, H.-I. Shih, C.-H. Lee, P.-W. Huang, N.-Y. Ko, W.-C. Ko, Y.-W. Chien); National Cheng Kung University Hospital, Tainan (H.-I. Shih, N.-Y. Ko, W.-C. Ko, Y.-W. Chien); Kuo General Hospital, Tainan (C.-H. Chung)

The Study

Migrant workers are required by law to undergo preemployment health examinations within 3 days of arrival in Taiwan. For this study, we recruited 600 newly arrived migrant workers from Indonesia, the Philippines, Thailand, and Vietnam (150 workers from each country) who received preemployment examinations at a regional hospital in Tainan, Taiwan, during June–August 2017. Workers who were >20 years of age and willing to participate were eligible without specific exclusion criteria. We used commercial ELISAs (https://focusdx.com) to test for IgM and IgG against dengue virus, anti-Zika virus IgG ELISA (Euroimmun AG, https://www.euroimmun.com) to test for IgG against Zika virus, and InBios Zika Detect IgM Capture ELISA (http://www.inbios.com) to test for IgM against Zika virus (because this assay seems to have higher sensitivity) (*6*). All tests were performed and interpreted according to manufacturers' instructions. We performed plaque reduction neutralization tests (PRNTs) on a subgroup of samples to detect neutralizing antibodies against 4 DENV serotypes (DENV-1, strain Hawaii; DENV-2, strain 16681; DENV-3, strain H87; DENV-4, strain H241) and 2 Zika virus strains (strain MR766 and 1 clinical isolate from a patient who acquired infection in Thailand). We calculated titers required to reduce dengue and Zika viral plaques by 50% ($PRNT_{50}$) and 90% ($PRNT_{90}$) (Appendix, https://wwwnc.cdc.gov/EID/article/25/4/18-1449-App1.pdf). For persons positive for Zika virus IgM, we used the interim case definition of laboratory-confirmed cases of recent Zika virus infection defined by the World Health Organization to see whether they fulfilled these criteria: IgM antibody against ZIKV positive and $PRNT_{90}$ for ZIKV with titer ≥20 and ZIKV $PRNT_{90}$ titer ratio ≥4 compared to other flaviviruses (*7*). This study was approved by the institutional review board of National Cheng Kung University Hospital (approval no. A-ER-106-045).

Most migrant workers were young adults 20–39 years of age (Appendix Table 1). Of the 600 workers, 18 (3.0%) were positive for Zika virus IgM and 233 (38.8%) were positive for Zika virus IgG. Only 3 (0.5%) workers had detectable DENV IgM, but 484 (80.7%) had DENV IgG. Seroprevalence of IgG against Zika virus and DENV was much lower in Vietnam than in the other 3 countries (Appendix Tables 2, 3).

Table 1. Zika virus and DENV serostatus for 600 migrant workers from Southeast Asia, Taiwan, 2017*

| | No. (%) workers | | | | |
| | Zika virus IgG+ | | Zika virus IgG– | | |
Serostatus	Zika virus IgM+	Zika virus IgM–	Zika virus IgM+	Zika virus IgM–	Total
DENV IgG+					
DENV IgM+	0	3 (0.5)	0	0	3 (0.5)
DENV IgM–	11 (1.8)	213 (35.5)	6 (1.0)	251 (41.8)	481 (80.2)
DENV IgG–					
DENV IgM+	0	0	0	0	0
DENV IgM–	0	6 (1.0)	1 (0.2)	109 (18.2)	116 (19.3)
Total	11 (1.8)	222 (37.0)	7 (1.2)	360 (60.0)	600 (100)

*DENV, dengue virus; +, positive; –, negative.

Among all workers, 227 (37.8%) had IgG against both Zika virus and DENV, 6 (1.0%) had only Zika virus IgG, 257 (42.8%) had only DENV IgG, and 110 (18.3%) were negative for both (Table 1). All 18 workers positive for Zika virus IgM were negative for DENV IgM, indicating that false-positive results from cross-reactivity with DENV IgM or polyclonal B cell stimulation were unlikely. We found that 6 (1%) workers positive for Zika virus IgM had Zika virus $PRNT_{90}$ titers ≥20, and their Zika virus $PRNT_{90}$ titer ratio was ≥4 compared with that of 4 serotypes of DENV (Table 2; Appendix Table 4). Although we did not perform PRNT for flaviviruses other than Zika virus and DENV, we assumed that these 6 workers fulfilled the World Health Organization criteria of confirmed Zika virus infection. All 3 workers with positive DENV IgM had detectable $PRNT_{50}$ and $PRNT_{90}$ titers against single or multiple DENV serotypes; thus, the positive ELISA results for DENV IgM were considered true positives. Among 6 participants with positive Zika virus IgG but negative DENV IgG, 5 had a high $PRNT_{50}$ titer against Zika virus (Appendix Table 4).

Conclusions

Zika virus IgM persists for ≈12 weeks (8); therefore, our results suggest that 1% of the workers had confirmed Zika virus infection within 3 months before blood collection, implying that the incidence of Zika virus infection in Southeast Asia might be severely underestimated. The median duration of viremia is 2 weeks (9); thus, some workers might have entered Taiwan during their viremia period and had the potential to spread Zika virus through *Aedes* mosquitoes in Taiwan. In addition, Zika virus can be detected in semen up to 6 months after symptom onset (10); thus, Zika virus transmission through sexual contact with these workers, who are at a sexually active age, is also possible. Furthermore, 3 female workers with confirmed Zika virus infection were of childbearing age, raising concerns about the risk for congenital infection.

The infectiousness of persons with asymptomatic Zika virus infection remains unknown. However, a recent study of DENV showed that asymptomatic persons could infect mosquitoes despite their lower average level of viremia (11). A recent modeling analysis estimated that 84% of DENV transmission is attributable to persons with inapparent infections because these persons are more mobile (12). If the infection characteristics of Zika virus are similar to those of DENV, the ability of fever screening programs at international airports and ports to prevent importation of Zika virus from migrant workers and travelers will be limited.

In this study, we may have overestimated Zika virus IgG seroprevalence (38.8%) because of false positivity resulting from cross-reactivity; nevertheless, the observed

Table 2. Serostatus and neutralizing antibody titers for Zika virus and DENV among 11 migrant workers from Southeast Asia who were IgM and IgG positive for Zika virus, Taiwan, 2017*

| | ELISA | | | | 90% PRNT titer | | | | | |
Worker no., age, y/sex	Zika virus IgM	Zika virus IgG	DENV IgM	DENV IgG	DENV-1	DENV-2	DENV-3	DENV-4	Thai	MR766
ID01, 21/F†	+	+	–	+	10	10	<10	<10	149	399
ID02, 37/F	+	+	–	+	609	1,401	40	<10	10	<10
ID03, 28/M	+	+	–	+	138	505	11	149	438	<10
VN01, 34/M†	+	+	–	+	<10	40	<10	<10	40	160
PH01, 28/F†	+	+	–	+	124	440	147	<10	1486	>2,560
PH02, 24/M†	+	+	–	+	<10	10	<10	<10	227	639
PH03, 24/F†	+	+	–	+	<10	<10	<10	<10	800	1279
TH01, 42/M	+	+	–	+	<10	40	<10	<10	107	143
TH02, 46/M†	+	+	–	+	<10	513	<10	<10	593	>2,560
TH03, 43/F	+	+	–	+	405	310	40	40	10	1,215
TH04, 31/M	+	+	–	+	>2,560	1,360	1,600	10	1,468	1,599

*DENV, dengue virus; DENV-1–4, DENV serotypes 1–4; MR766, African Zika virus strain MR766; PRNT, plaque reduction neutralization test; Thai, 1 Zika virus isolate from a worker with an imported case acquired in Thailand.
†Six persons positive for Zika virus IgM fulfilled the criteria for laboratory confirmation of recent Zika virus infection adopted with definition according to the World Health Organization (7).

seroprevalence is comparable with that in Martinique (42.2%) and French Polynesia (49%) but lower than that in Brazil (63.3%) and Micronesia (73%) (*2*). It remains unclear why only very few cases of Zika virus–related microcephaly have been reported in Southeast Asia (*13*) despite such high seroprevalence. Possible explanations are differences in virus strains, differences in host factors, and limitations of the surveillance system (*14*).

To our surprise, seroprevalence of DENV IgM was lower than that of Zika virus IgM. In DENV-hyperendemic countries, children may have been exposed to multiple DENV serotypes and then acquired immunity; therefore, the incidence of dengue in adults is relatively low. Also of note, we observed much lower seroprevalences of Zika virus and DENV among workers in Taiwan from Vietnam, which may be because most of these workers originally came from rural areas in subtropical northern Vietnam, where the population density and climate are not suitable for establishing endemic transmission cycles of mosquitoborne viruses (*15*).

Our finding that 1% of migrant workers from Southeast Asia had laboratory-confirmed recent Zika virus infection suggests that the incidence of Zika virus infection in this region is underestimated. Given the convenience of flight for global travel, the risk for international dissemination of Zika virus by workers and travelers originating from Southeast Asia cannot be neglected.

About the Author

Dr. Perng is a professor at the Department of Microbiology and Immunology, College of Medicine, National Cheng Kung University, Tainan, Taiwan. His primary research interests are the role and fate of stem and progenitor cells during the acute stages of DENV infection, their development in asymptomatic and persistent DENV infection, and the epidemiology of Zika virus and DENV infections.

References

1. Kuan MM, Chang FY. Airport sentinel surveillance and entry quarantine for dengue infections following a fever screening program in Taiwan. BMC Infect Dis. 2012;12:182. http://dx.doi.org/10.1186/1471-2334-12-182
2. Fritzell C, Rousset D, Adde A, Kazanji M, Van Kerkhove MD, Flamand C. Current challenges and implications for dengue, chikungunya and Zika seroprevalence studies worldwide: a scoping review. PLoS Negl Trop Dis. 2018;12:e0006533. http://dx.doi.org/10.1371/journal.pntd.0006533
3. Ho ZJM, Hapuarachchi HC, Barkham T, Chow A, Ng LC, Lee JMV, et al.; Singapore Zika Study Group. Outbreak of Zika virus infection in Singapore: an epidemiological, entomological, virological, and clinical analysis. Lancet Infect Dis. 2017; 17:813–21. http://dx.doi.org/10.1016/S1473-3099(17)30249-9
4. Chu DT, Ngoc VTN, Tao Y. Zika virus infection in Vietnam: current epidemic, strain origin, spreading risk, and perspective. Eur J Clin Microbiol Infect Dis. 2017;36:2041–2. http://dx.doi.org/10.1007/s10096-017-3030-8
5. Khongwichit S, Wikan N, Auewarakul P, Smith DR. Zika virus in Thailand. Microbes Infect. 2018;20:670–5.
6. Granger D, Hilgart H, Misner L, Christensen J, Bistodeau S, Palm J, et al. Serologic testing for Zika virus: comparison of three Zika virus IgM-screening enzyme-linked immunosorbent assays and initial laboratory experiences. J Clin Microbiol. 2017; 55:2127–36. http://dx.doi.org/10.1128/JCM.00580-17
7. Ward MJ, Alger J, Berrueta M, Bock H, Buekens P, Cafferata ML, et al. Zika virus and the World Health Organization criteria for determining recent infection using plaque reduction neutralization testing. Am J Trop Med Hyg. 2018;99:780–2. http://dx.doi.org/10.4269/ajtmh.18-0237
8. Landry ML, St George K. Laboratory diagnosis of Zika virus infection. Arch Pathol Lab Med. 2017;141:60–7. http://dx.doi.org/10.5858/arpa.2016-0406-SA
9. Paz-Bailey G, Rosenberg ES, Doyle K, Munoz-Jordan J, Santiago GA, Klein L, et al. Persistence of Zika virus in body fluids–preliminary report. N Engl J Med. 2017;379. h ttp://dx.doi.org/10.1056/NEJMoa1613108
10. Nicastri E, Castilletti C, Liuzzi G, Iannetta M, Capobianchi MR, Ippolito G. Persistent detection of Zika virus RNA in semen for six months after symptom onset in a traveller returning from Haiti to Italy, February 2016. Euro Surveill. 2016;21:pii:30314. http://dx.doi.org/10.2807/1560-7917.ES.2016.21.32.30314
11. Duong V, Lambrechts L, Paul RE, Ly S, Lay RS, Long KC, et al. Asymptomatic humans transmit dengue virus to mosquitoes. Proc Natl Acad Sci U S A. 2015;112:14688–93. http://dx.doi.org/10.1073/pnas.1508114112
12. Ten Bosch QA, Clapham HE, Lambrechts L, Duong V, Buchy P, Althouse BM, et al. Contributions from the silent majority dominate dengue virus transmission. PLoS Pathog. 2018;14:e1006965. http://dx.doi.org/10.1371/journal.ppat.1006965
13. Wongsurawat T, Athipanyasilp N, Jenjaroenpun P, Jun SR, Kaewnapan B, Wassenaar TM, et al. Case of microcephaly after congenital infection with Asian lineage Zika virus, Thailand. Emerg Infect Dis. 2018;24:1758–61. http://dx.doi.org/10.3201/eid2409.180416
14. Lim SK, Lim JK, Yoon IK. An update on Zika virus in Asia. Infect Chemother. 2017;49:91–100. http://dx.doi.org/10.3947/ic.2017.49.2.91
15. Rabaa MA, Simmons CP, Fox A, Le MQ, Nguyen TTT, Le HY, et al. Dengue virus in sub-tropical northern and central Viet Nam: population immunity and climate shape patterns of viral invasion and maintenance. PLoS Negl Trop Dis. 2013;7:e2581. http://dx.doi.org/10.1371/journal.pntd.0002581

Address for correspondence: Yu-Wen Chien, Department of Public Health, College of Medicine, National Cheng Kung University, No. 1, University Rd, Tainan 70101, Taiwan; email: yuwenchien@mail.ncku.edu.tw

Seroprevalence of Zika and Dengue Virus Antibodies among Migrant Workers, Taiwan

Appendix

Plaque Reduction Neutralization Test for Dengue Virus and Zika Virus

Plaque reduction neutralization tests (PRNT) were performed to detect the antibody neutralization titers for four serotypes of dengue virus (DENV1 – 4) and two strains of Zika virus (ZIKV). The viruses used in current PRNT tests were DENV1 (strain Hawaii), DENV2 (strain 16681), DENV3 (strain H87), and DENV4 (strain H241), and ZIKV (strain MR766 and one recent clinical isolate from an imported subject who was infected in Thailand). The PRNT protocols for DENV and ZIKV were previously described (*1*,*2*). Briefly, PRNTs were performed in baby hamster kidney fibroblasts (BHK-21) cells and Vero cells for DENV and ZIKV, respectively. A total of 7×10^5 cell per well of BHK-21 cells and Vero cells were seeded in 6-well plates 1 day before PRNT assays. The complement in human serum was inactivated at 56°C for 30 minutes. The inactivated serum was then subjected to 4-fold serial dilution from 1:10 to 1:2560 with 2% DMEM. Viruses at 10^4 PFU in 250uL were mixed with equal volume of diluted serum at 37°C for 30 minutes. The virus-serum mixtures were agitated every 15 minutes, and then 400uL of mixtures were added into each well of 6-well plates that had been seeded with cells. The plates were incubated at 37°C in 5% CO2 incubator for 2 hours, the inoculants were then aspirated, and overlaid with 3mL per well of 1% methyl cellulose solutions (2% FBS of DMEM, pH7.4~7.6). The plates were incubated at 37°C in 5% CO_2 incubator for 7 days. The overlay media were removed from plates after 7 days, and the cells were stained with 2mL of crystal violet for 2 hours and followed by rinsing with water. Plaque numbers were counted by the naked eye under the transmitted light box. Antibody titers were calculated based on reciprocal of the diluted serum titers, in which 50% and 90% reduction in plaque counts ($PRNT_{50}$ and $PRNT_{90}$) compared to controls were obtained by using GraphPad Prism version 7 for Windows (GraphPad Software, La Jolla, CA, USA). If the titer could not be calculated, it was

expressed as the reciprocal of the highest diluted serum titers by showing ≥50% or ≥90% reduction in plaque counts as compared with that of controls.

References

1. Morens DM, Halstead SB, Repik PM, Putvatana R, Raybourne N. Simplified plaque reduction neutralization assay for dengue viruses by semimicro methods in BHK-21 cells: comparison of the BHK suspension test with standard plaque reduction neutralization. J Clin Microbiol. 1985;22:250–4. PubMed

2. Müller JA, Harms M, Schubert A, Mayer B, Jansen S, Herbeuval JP, et al. Development of a high-throughput colorimetric Zika virus infection assay. Med Microbiol Immunol (Berl). 2017;206:175–85. PubMed http://dx.doi.org/10.1007/s00430-017-0493-2

Appendix Table 1. Sex and age distribution of migrant workers from four Southeast Asian countries.

Category	Indonesia	Philippines	Thailand	Vietnam	All
			N (% of total)		
Total	150	150	150	150	600
Sex					
Female	75 (50.0%)	129 (86.0%)	30 (20.0%)	63 (42.0%)	297 (49.5%)
Male	75 (50.0%)	21 (14.0%)	120 (80.0%)	87 (58.0%)	303 (50.5%)
Age					
20–29	66 (44.0%)	120 (80.0%)	52 (34.7%)	83 (55.3%)	321 (53.5%)
30–39	73 (48.7%)	29 (19.3%)	54 (36.0%)	57 (38.0%)	213 (35.5%)
≥40	11 (7.4%)	1 (0.7%)	44 (29.3%)	10 (6.7%)	66 (22%)

Appendix Table 2. Seroprevalence of Zika virus-specific IgG and IgM among migrant workers from four Southeast Asian countries.

| | | | | | No. of positive (% positive) | | | | | |
| Category | Indonesia | | Philippines | | Thailand | | Vietnam | | All | |
	IgM	IgG	IgM	IgG	IgM	IgG	IgM	IgG	IgM	IgG
Total	7 (4.7)	65 (43.3)	5 (3.3)	83 (55.3)	5 (3.3)	76 (50.7)	1 (0.7)	9 (6.0)	18 (3.0)	233 (38.8)
Sex										
Female	4 (5.3)	27 (36.0)	4 (3.1)	71 (55.0)	1 (3.3)	19 (63.3)	0 (0.0)	2 (3.2)	9 (3.0)	119 (40.1)
Male	3 (4.0)	38 (50.7)	1 (4.8)	12 (57.1)	4 (3.3)	57 (47.5)	1 (1.1)	7 (8.0)	9 (3.0)	114 (37.6)
Age										
20–29	2 (3.0)	31 (47.0)	5 (4.2)	68 (56.7)	0 (0.0)	22 (42.3)	0 (0.0)	6 (7.2)	7 (2.2)	127 (39.6)
30–39	5 (6.8)	30 (41.1)	0 (0.0)	15 (51.7)	1 (1.9)	31 (57.4)	1 (1.8)	3 (5.3)	7 (3.3)	79 (37.1)
≥40	0 (0.0)	4 (40.0)	0 (0.0)	0 (0.0)	4 (9.8)	23 (52.3)	0 (0.0)	0 (0.0)	4 (6.1)	27 (40.9)

Appendix Table 3. Seroprevalence of dengue virus-specific IgG and IgM among migrant workers from four Southeast Asian countries.

| | | | | | No. of positive (% positive) | | | | | |
| Category | Indonesia | | Philippines | | Thailand | | Vietnam | | All | |
	IgM	IgG	IgM	IgG	IgM	IgG	IgM	IgG	IgM	IgG
Total	0(0.0)	136 (90.7)	1 (0.7)	142 (94.7)	2 (1.3)	136 (90.7)	0 (0.0)	70 (46.7)	3 (0.5)	484 (80.7)
Sex										
Female	0 (0.0)	67 (89.3)	1 (0.8)	122 (94.6)	0 (0.0)	26 (86.7)	0 (0.0)	26 (41.3)	1 (0.3)	241 (81.1)
Male	0 (0.0)	69 (92.0)	0 (0.0)	20 (95.2)	2 (1.7)	110 (91.7)	0 (0.0)	44 (50.6)	2 (0.7)	243 (80.2)
Age										
20–29	0 (0.0)	61 (92.4)	1 (0.8)	115 (95.8)	0 (0.0)	42 (80.8)	0 (0.0)	32 (38.6)	1 (0.3)	250 (77.9)
30–39	0 (0.0)	66 (90.4)	0 (0.0)	26 (89.7)	2 (3.7)	52 (96.3)	0 (0.0)	33 (57.9)	2 (0.9)	177 (83.1)
≥40	0 (0.0)	9 (81.8)	0 (0.0)	1 (100.0)	0 (0.0)	42 (95.5)	0 (0.0)	5 (50.0)	0 (0.0)	57 (86.4)

Appendix Table 4. Serostatuses and titers of neutralizing antibodies for Zika virus and dengue virus infection among selected workers*

Worker No.	Age	Sex	ELISA				PRNT$_{90}$						PRNT$_{50}$					
			Zika IgM	Zika IgG	DEN IgM	DEN IgG	D1	D2	D3	D4	Thai	MR766	D1	D2	D3	D4	Thai	MR766
ID01†	21	F	+	+	-	+	10	10	<10	<10	149	399	78	40	36	33	150	400
ID02	37	F	+	+	-	+	609	1401	40	<10	10	<10	610	1402	640	39	102	38
ID03	28	M	+	+	-	+	138	505	11	149	438	<10	139	507	68	150	440	<10
VN01†	34	M	+	+	-	+	<10	40	<10	<10	160	<10	<10	153	<10	<10	253	1066
PH01†	28	F	+	+	-	+	124	440	147	<10	1486	>2560	126	441	149	10	>2560	>2560
PH02†	24	M	+	+	-	+	<10	10	<10	<10	227	639	<10	259	40	<10	160	640
PH03†	24	F	+	+	-	+	<10	<10	<10	<10	800	1279	<10	25	<10	<10	>2560	>2560
TH01	42	M	+	+	-	+	<10	40	<10	<10	107	143	<10	214	<10	35	109	160
TH02†	46	M	+	+	-	+	<10	513	<10	<10	593	>2560	38	515	31	38	640	>2560
TH03	43	F	+	+	-	+	405	310	40	40	10	1215	406	312	249	269	764	>2560
TH04	31	M	+	+	-	+	>2560	1360	1600	10	1468	1599	>2560	>2560	>2560	82	>2560	1600
ID04	31	F	+	-	-	+	<10	10	<10	<10	<10	<10	<10	40	<10	<10	<10	82
ID05	37	M	+	-	-	+	<10	10	40	<10	<10	<10	<10	40	40	35	<10	459
ID06	32	M	+	-	-	+	<10	10	34	<10	<10	40	<10	40	39	34	40	160
ID07	33	F	+	-	-	+	10	34	<10	<10	<10	<10	40	39	38	<10	<10	38
PH04	23	F	+	-	-	+	<10	40	<10	10	<10	29	<10	160	<10	40	<10	30
TH05	43	M	+	-	-	+	<10	92	141	<10	10	155	38	93	143	<10	101	>2560
PH05	22	F	-	+	+	+	<10	<10	10	10	10	579	<10	37	40	40	239	589
TH06	33	M	-	+	+	+	10	160	<10	<10	<10	<10	62	1102	28	38	40	776
TH07	37	M	-	+	+	+	<10	446	<10	513	<10	<10	10	448	<10	640	<10	<40
VN02	23	F	-	+	-	-	<10	<10	<10	<10	<10	<10	<10	<10	<10	<10	111	<10
PH06	33	M	-	+	-	-	<10	<10	<10	<10	144	40	<10	<10	<10	<10	145	640
TH08	48	M	-	+	-	-	<10	382	<10	<10	<10	356	<40	384	40	36	107	357
TH09	41	M	-	+	-	-	<10	40	<10	<10	<10	40	36	337	<10	<10	108	80
TH10	21	M	-	+	-	-	<10	<10	<10	<10	<10	<10	<10	<10	<10	<10	374	<10
TH11	30	M	-	+	-	-	<10	<10	<10	<10	<10	<10	<10	<10	10	<10	<10	10

*ELISA, enzyme-linked immunosorbent assay; PRNT, plaque reduction neutralization test; ZIKV, Zika virus; DENV, dengue virus; D1, dengue virus serotype 1; D2, dengue virus serotype 2; D3, dengue virus serotype 3; D4, dengue virus serotype 4; Thai, one Zika virus isolate from an imported cases who got infection in Thailand; MR766, the African Zika virus strain MR766.
†Six people with positive anti-ZIKV IgM fulfilled the criteria for laboratory confirmation of recent ZIKV infection.