Zika Virus-Associated Cognitive Impairment in Adolescent, 2016

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DOI: https://dx.doi.org/10.3201/eid2306.162029

Incidence of neurologic manifestations associated with Zika virus infection has been increasing. In 2016, neuropsychological and cognitive changes developed in an adolescent after travel to a Zika virus–endemic area. Single-photon emission computed tomography and neuropsychological testing raised the possibility that Zika virus infection may lead to neuropsychiatric and cognitive symptoms.

A lthough the full clinical spectrum of complications associated with Zika virus disease remains unclear, evidence of an association between Zika virus infection and diseases of the nervous system is growing (1). Increased incidence of neurologic manifestations has been reported, including cerebral birth abnormalities (2), Guillain-Barré Syndrome (3), meningoencephalitis (4), and memory loss (5). We describe a case of probable central nervous system (CNS) infection with Zika virus associated with the onset of neuropsychological and cognitive changes in an adolescent.

In summer 2016, an adolescent traveled to a Zika virus–endemic island in the southern Caribbean, where many insect bites occurred. Before the trip, the patient's medical history included mild depression treated with a selective serotonin reuptake inhibitor. Assessment after return to the United States indicated that mental and physical health were the same as before travel. One week after return, the patient experienced sore throat, headache, diffuse scarlatiniform rash (online Technical Appendix Figure 1, https:// wwwnc.cdc.gov/EID/article/23/6/16-2029-Techapp1.pdf), joint pain, confusion, and short-term memory loss; no fever or eye inflammation were noted.

Three days after symptom onset, the patient received care from a pediatrician; extensive laboratory testing revealed urine reverse transcription PCR (RT-PCR) results positive for Zika virus. Serologic results were positive for Epstein-Barr virus, consistent with the patient's history of infection 15 months earlier (online Technical Appendix Table 1). Five days after symptom onset, the sore throat, headache, rash and joint pain resolved; however, 8 days after symptom onset, the neuropsychiatric symptoms worsened and included excessive energy, decreased sleep, rapid and tangential speech, grandiose thinking, impulsivity, decreased inhibition, and behavioral regression suggestive of hypomania. The patient's psychiatrist prescribed an antipsychotic medication. An ambulatory and 24-hour electroencephalogram and magnetic resonance imaging of the brain without contrast showed no abnormalities.

Symptoms failed to improve, and 13 days after symptom onset, the patient was hospitalized for further work-up. Fourteen days after symptom onset, magnetic resonance imaging of the brain with contrast showed no abnormalities. Serum was positive for dengue virus IgG and IgM (online Technical Appendix Table 2). Repeat Zika virus testing was not performed because of the prior positivity of urine RT-PCR. Fifteen days after symptom onset, cerebrospinal fluid (CSF) obtained by lumbar puncture contained 8 leukocytes/µL, 1,000 erythrocytes/µL, 55 mg/dL glucose, and 30 mg/dL protein. CSF testing for Zika virus, performed at the Wadsworth Laboratory, New York City Department of Mental Health and Hygiene (New York, NY, USA), was positive for IgM but negative by RT-PCR.

After discharge, the patient's symptoms of regression, disinhibition, and cognitive impairment persisted. Seven weeks after symptom onset, single-photon emission computed tomography revealed mildly heterogeneous cerebral cortical perfusion, with focal moderateto-severe hypoperfusion in the inferior left frontal region (online Technical Appendix Figure 2); neuropsychological testing demonstrated evidence of superior intellectual ability (Wechsler Intelligence Scale for Children-fourth edition, General Ability Index), probably reflecting function before illness (online Technical Appendix Table 3). Processing speed was significantly slowed relative to other skills; the Processing Speed Index score was >2 SDs lower than the General Ability Index score. Performance on most memory tests and tests of executive function was within normal limits yet lower than expected given reported performance before illness. Immediate and delayed visual recall of the Rey complex figure was poor, reflecting primary difficulty encoding new visual information. On a standardized behavioral questionnaire, the patient self-reported psychiatric symptoms including anxiety, racing thoughts, and an inability to turn off thoughts. The patient had not experienced racing thoughts before the Zika virus infection, and the anxiety symptoms had worsened since the infection. In addition, the patient reported significant and functionally limiting fatigue. Nine weeks after onset, symptoms were better but not resolved; because of concerns that these symptoms were triggered by a postinfectious immune-mediated process, a trial of intravenous immunoglobulin was administered. Fifteen

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weeks after symptom onset, the patient's symptoms were better but not fully resolved.

Although we cannot entirely rule out Epstein-Barr virus as a possible trigger, the length of time between previous infection and onset of neuropsychological symptoms would be unusual. In addition, although it is impossible to exclude contributions of coinfection from other mosquitoborne viruses (e.g., dengue and chikungunya), given the Zika virus positivity on RT-PCR, the patient's condition met criteria for definitive Zika virus infection and the CSF IgM titer was consistent with CNS involvement of Zika virus. The changes on single-photon emission computed tomographs and neuropsychological test scores raise the possibility that Zika virus infection may trigger neuropsychiatric and cognitive symptoms. Although we cannot prove that the patient's symptoms were related to Zika virus, clinicians should be aware of this potential association and the value of closely monitoring patients with Zika virus infection.

K.T., who helped edit the manuscript, is a World Health Organization consultant on neurologic manifestations in the context of the Zika virus outbreak, receives an honorarium as chief consult editor on Zika virus for Medscape consults, and participates in Zika research through the Neuroviruses Emerging in the Americas Study. K.T. receives funding support from the National Institutes of Health, and J.Z. receives funding support from a National Institutes of Health training grant (T32 AI007531).

Dr. Zucker is a postdoctoral fellow in adult and pediatric infectious diseases at Columbia University Medical Center. His research interests include improving prevention and treatment for adolescents and young adults living with, or at risk for, HIV infection, hepatitis C, and sexually transmitted diseases.

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Highly Pathogenic Avian Influenza Virus (H5N8) Clade 2.3.4.4 Infection in Migratory Birds, Egypt

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DOI: https://dx.doi.org/10.3201/eid2306.162056

We isolated highly pathogenic avian influenza virus (H5N8) of clade 2.3.4.4 from the common coot (*Fulica atra*) in Egypt, documenting its introduction into Africa through migratory birds. This virus has a close genetic relationship with subtype H5N8 viruses circulating in Europe. Enhanced surveillance to detect newly emerging viruses is warranted.

A vian influenza is a highly contagious disease of poultry that continues to spread across the globe in bird populations. Occasionally, transmission of a highly pathogenic avian influenza virus (HPAIV) from infected poultry to humans results in a severe public health crisis (1).

In 2010, strains of HPAIV (H5N8) of clade 2.3.4.4 were first detected among wild birds in Asia and later spread to domestic birds across China, South Korea, and Japan (2,3). Most recently, a novel reassortant virus of sub-type H5N8 clade 2.3.4.4 was reported in Russia and further spread to many countries in Europe, Asia, and the Middle East (4,5). The spread of HPAIV (H5N8) strains has been linked to the overlapping flyways of migratory wild birds that come from different continents; this mingling of wild birds poses a major concern worldwide (4,6).

Egypt is one of the most notable migration spots for migratory birds crossing Europe, Asia, and Africa. In early winter each year, thousands of migrating waterfowl use Egypt as a resting stop before they continue their journey southward through the African continent through the East Africa/East Asia and Mediterranean/Black Sea migratory

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Technical Appendix

Technical Appendix Table 1. Outpatient laboratory evaluation of acute illness and mental status changes in an adolescent returning from a Zika endemic territory

| Date | Test (units) | Result | Reference |
|---------|--------------------------------------|----------|-----------|
| 8/6/16* | EBV EBNA (U/ml) | 75.4 (H) | 0-21.9 |
| | EBV VCA IgG (U/ml) | 100 (H) | 0-21.9 |
| | EBV VCA IgM (U/ml) | <10 | 0-43.9 |
| | EBV IgG (U/ml) | 9.3 | 0-10.9 |
| | Mycoplasma IgG (U/L) | .02 | <0.09 |
| | Mycoplasma IgM (U/L) | .21 U/L | <0.76 |
| | Parvovirus IgG (IV) | 5.73 (H) | <0.89 |
| | Parvovirus IgM (IV) | .24 | <0.89 |
| | Antinuclear antibody | Negative | Negative |
| | Antistreptolysin O Titer (IU/ml) | 63 | 0-330 |
| | DNase B antibody (U/ml) | 169 | 0–310 |
| | Sedimentation rate (mm/hr) | 8 | 0–24 |
| | Thyroid stimulating hormone (uIH/mL) | 1.2 | .4 – 4.2 |
| | Vitamin B12 (pg/ml) | 406 | 211–911 |
| | Folate (ng/mL) | 10.17 | 4-1000 |
| | Vitamin D 25 Hydroxy (NG/mI) | 42.5 | 30–100 |
| 8/6/16† | Lyme Western Blot IgM | Negative | Neg |
| | Lyme Western Blot IgG | Negative | Neg |
| | Serum N methyl D Aspartate Receptor | <1:10 | <1:10 |
| | Antibody | | |
| 8/6/16‡ | Zika Serum RT-PCR | Negative | Negative |
| | Zika Serology (IgM) | Negative | Negative |
| | Zika Urine RT-PCR | Positive | Negative |

*Labs performed at Mount Sinai Beth Israel Laboratory.

†Labs performed at an outside hospital laboratory. ‡Labs performed at New York Department of Health.

| Technical Appendix Table 2. Inpatient laboratory evaluation of acute illness and mental status changes in an adolescent returning | ١g |
|---|----|
| from a Zika endemic territory | |

| Date | Test | Result | Reference |
|----------|---|-----------|------------|
| Serum | Urine toxicology | Negative | Negative |
| 8/16/16* | | - | - |
| | C-Reactive Protein (mg/L) | <0.15 | 0–10 |
| | Sedimentation rate (mm/hr) | 10 | 0–20 |
| | Dengue Fever Virus Antibody IgM (IV) | 2.15 (H) | <1.64 |
| | Dengue Fever Virus Antibody IgG (IV) | 14 (H) | <1.64 |
| | Total IgG (mg/dL) | 941 ΄ | 714 - 1711 |
| CSF | CSF Cell Count (cells/microliter) | 8 WBCs | 0 - 5 |
| 8/18/16* | | 1000 RBCs | 0 |
| | CSF Glucose (mg/dL) | 55 (L) | 60 - 80 |
| | CSF IgG and Total Protein Ratio | 6.7 | 6–13 |
| | BioFire® Meningitis/Encephalitis RT- PCR Panel | Negative | Negative |
| | Oligoclonal Bands | Negative | Negative |
| 3/18/16* | CSF N methyl D Aspartate Receptor Antibody, IgG | Negative | Negative |
| 8/18/16† | CSF Zika RT-PCR | Negative | Negative |
| • | CSF Zika IgM | Positive | Negative |

*Labs performed at New York Presbyterian Columbia.

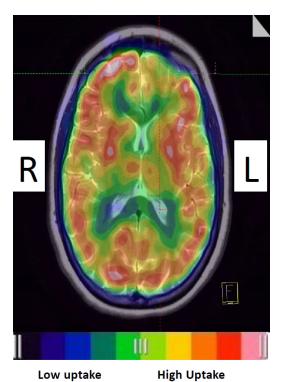
†Labs performed at New York Department of Health.

Technical Appendix Table 3. Neuropsychological test findings

| Standard Score 118 117 118 115 83 121 T/z-score 45 0 Scaled Score 12 13 T-score 29 24 27 Scaled Score 11 12 | Percentile 88 87 88 84 13 92 Percentile 34 50 Percentile 75 84 Percentile 2 1 1 Percentile |
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Technical Appendix Figure 1. Scarlitiform rash in a patient with urine positive for Zika virus on reverse transcription PCR. Picture taken 4 days after the onset of symptoms.



Technical Appendix Figure 2. This ^{99m}Tc-ethyl cysteinate dimer (ECD) fusion of brain SPECT and MRI shows the localization of focal left inferofrontal lobe moderate-severely decreased hypoperfusion (crosshairs), along with a non-specific heterogeneous global uptake pattern.