

Porcine Coronavirus HKU15 Detected in 9 US States, 2014

To the Editor: Porcine coronavirus (PorCoV) HKU15 is a single-stranded, positive-sense, enveloped RNA virus belonging to the genus *Deltacoronavirus* (family *Coronaviridae*). PorCoV HKU15 was first identified in 2012 in a surveillance study from China (1). Until February 2014, however, the role of this virus strain in clinical diseases of pigs had not been reported.

We recently reported the detection of PorCoV strain HKU15-OH1987 in feces samples from sows and intestine samples from piglets in Ohio, United States; the infected animals were from swine farms where outbreaks of diarrheal disease had occurred in late January and early February 2014 (2). Genetic analysis showed that HKU15-OH1987 is closely related to 2 deltacoronavirus strains that were detected in Hong Kong, China, in 2012: HKU15-155 and HKU15-44 (2). We also demonstrated the presence of histopathologic lesions in the small intestines of PorCoV HKU15-infected piglets with diarrhea (L. Wang, unpub. data). In April 2014, a novel swine enteric coronavirus disease caused by PorCoV HKU15 or porcine epidemic diarrhea virus was reported to the World Animal Health Organization by the US Department of Agriculture (http://www.oie.int/wahis_2/public/wahid.php/ReviewReport/Review?page_refer=MapFullEventReport&reportid=15133).

PorCoV HKU15 is now recognized as a key pathogenic cause of diarrheal diseases in pigs in the United States. However, the geographic distribution and genotype diversity of PorCoV HKU15 in this country are still not clear. To further our knowledge of the virus, we analyzed swine samples that had been submitted for

diagnosis of diarrheal disease from farms in 10 US states. We report the detection and phylogenetic analyses of PorCoV HKU15 strains from these samples.

Between February 7, 2014, when PorCoV HKU15-OH1987 was first identified in Ohio (2), and April 9, 2014, the Animal Disease Diagnostic Laboratory of the Ohio Department of Agriculture received >2,000 swine samples from farms in 10 US states for diagnosis of diarrheal disease. The states from which samples had been submitted were Minnesota, South Dakota, Nebraska, Illinois, Indiana, Michigan, Kentucky, Pennsylvania, Maryland, and Ohio. Among those samples, 435 were selected to be tested for the presence of PorCoV HKU15. A real-time reverse transcription PCR assay targeting the membrane protein gene was used to identify PorCoV HKU15. Samples with a cycle threshold value of <35 were considered positive on the basis of validation data using the cloned membrane protein gene (data not shown). Of the 435 samples, 109 (25%) from 9 states (all states mentioned above, excluding Maryland) were positive for PorCoV HKU15 by real-time reverse

transcription PCR (Figure). Of those 109 samples, 19 (17%) were also positive for porcine epidemic diarrhea virus. This result suggests that PorCoV HKU15 is prevalent among pig populations in the major pig-producing US states.

To determine the genetic diversity of PorCoV HKU15 strains from the 9 states, we conducted whole-genome sequencing for 1 strain from each state by using 16 pairs of previously described, overlapping primers (2). Strain names were designated by the state abbreviation and case number. The complete sequence for HKU15-OH1987 was reported previously (2). Sequence analysis showed that strains from South Dakota (SD3424), Nebraska (NE3579), Illinois (IL2768), Indiana (IN2847), Kentucky (KY4813), Michigan (MI6148), and Pennsylvania (PA3148) have the same genome size (25,422 nt) as OH1987, and whole-genome pairwise comparison showed that they share a high nucleotide similarity ($\geq 99.8\%$). Furthermore, all of the isolates share high nucleotide similarity (98.9%–99.2%) with the 2 PorCoV HKU15 strains in GenBank, HKU15-155 and HKU15-44.

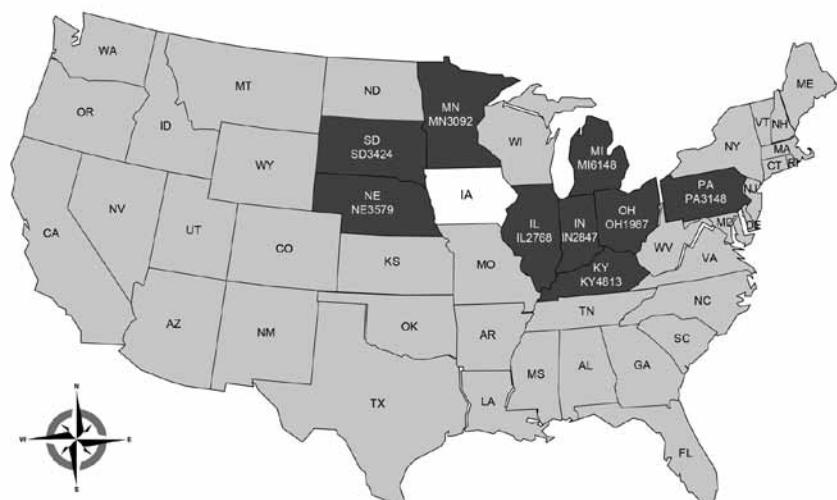


Figure. US states with swine samples positive for porcine coronavirus (PorCoV) HKU15, between February and April 2014. A total of 435 samples from 10 states were selected to be tested for the presence of PorCoV. Of those samples, 109 (25%) from 9 states (dark gray) were positive for PorCoV HKU15. Another recent article reported the presence of PorCoV HKU15 in Iowa (white) (3). Strain names are indicated below state abbreviations.

A phylogenetic tree constructed by using the entire sequence showed that all PorCoV HKU15 strains from the United States clustered together in 1 clade of the genus *Deltacoronavirus* with HKU15-155 and HKU15-44 (online Technical Appendix Figure, panel A, <http://wwwnc.cdc.gov/EID/article/20/9/14-0756-Techapp1.pdf>). This finding indicates that 1 genotype of PorCoV HKU15 is currently circulating in multiple US states. This result was further supported by phylogenetic trees constructed by using the full-length amino acids of spike and nucleocapsid proteins (online Technical Appendix Figure, panels B, C). Because of limitation of the samples received, only a partial genome sequence was determined for strain MN3092 from Minnesota. However, on the basis of the spike and nucleocapsid protein sequence analyses, it is highly likely that the entire genome of the Minnesota strain is genetically identical to that of the other 8 strains (online Technical Appendix Figure, panels B, C).

In addition to the 9 states reported in this study, Iowa has also had a recent detection of PorCoV HKU15 (3). Thus, PorCoV HKU15 has been detected in 10 of the 50 US states, and those 10 states mainly cluster in the midwestern United States (Figure). As with data collected for porcine epidemic diarrhea outbreaks by the US National Animal Health Laboratory Network (<http://www.nahln.org/default/>), data collected on the geographic location and numbers of PorCoV HKU15 cases is also required to be reported weekly.

Earlier reports support avian coronaviruses as the gene source for *Deltacoronavirus* spp. (1,4). To confirm this, surveillance for PorCoV should be carried out among birds. Moreover, effective control strategies, including vaccine development, should be in place for prevention and control of infections caused by PorCoV HKU15.

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References

1. Woo PC, Lau SK, Lam CS, Lau CC, Tsang AK, Lau JH, et al. Discovery of seven novel mammalian and avian coronaviruses in the genus *Deltacoronavirus* supports bat coronaviruses as the gene source of *Alphacoronavirus* and *Betacoronavirus* and avian coronaviruses as the gene source of *Gammacoronavirus* and *Deltacoronavirus*. *J Virol*. 2012;86:3995–4008. <http://dx.doi.org/10.1128/JVI.06540-11>
2. Wang L, Byrum B, Zhang Y. Detection and genetic characterization of a delta-coronavirus in pigs in the United States. *Emerg Infect Dis*. 2014;
3. Li G, Chen Q, Harmon KM, Yoon KJ, Schwartz KJ, Hoogland MJ, et al. Full-length genome sequence of porcine deltacoronavirus strain USA/IA/2014/8734. *Genome Announc*. 2014;2:e00278-14. <http://dx.doi.org/10.1128/genomeA.00278-14>
4. Chu DK, Leung CY, Gilbert M, Joyner PH, Ng EM, Tse TM, et al. Avian coronavirus in wild aquatic birds. *J Virol*. 2011;85:12815–20. <http://dx.doi.org/10.1128/JVI.05838-11>

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Schistosomiasis Haematobium, Corsica, France

To the Editor: In Europe, urinary schistosomiasis (*I*) has previously been detected only in Portugal, where this focus disappeared during the 1950s (2). However, freshwater snails of the species *Bulinus contortus*, *B. truncatus*, and *Planorbarius metidjensis*, which are recognized intermediate hosts for *Schistosoma haematobium* trematodes, have been found in Portugal (3), Spain (4), and Corsica (5,6). This finding suggested that autochthonous schistosomiasis could re-emerge in southern Europe if these mollusks become infected. We report a probable focus for transmission of schistosomiasis haematobium in Corsica, France.

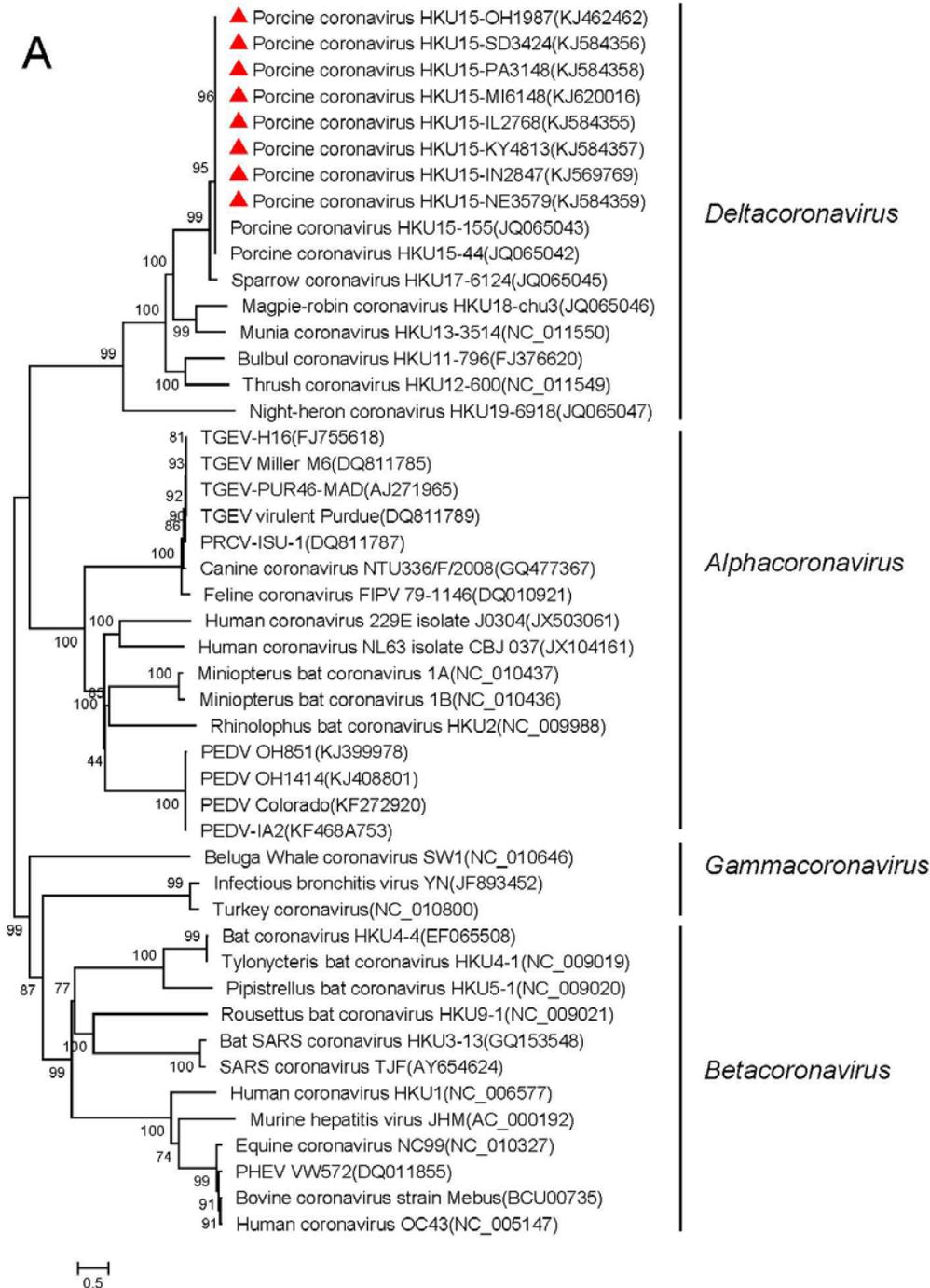
In March 2014, a 4-year-old girl (index case-patient) from France was referred to the Toulouse University Hospital (Toulouse, France), with gross hematuria. Ultrasonography and cystoscopic examination of the bladder detected a polyp. Examination of the polyp for parasites identified bodies that were consistent with schistosome eggs. Parasitologic examination of urine confirmed schistosomiasis by detecting viable *S. haematobium* eggs.

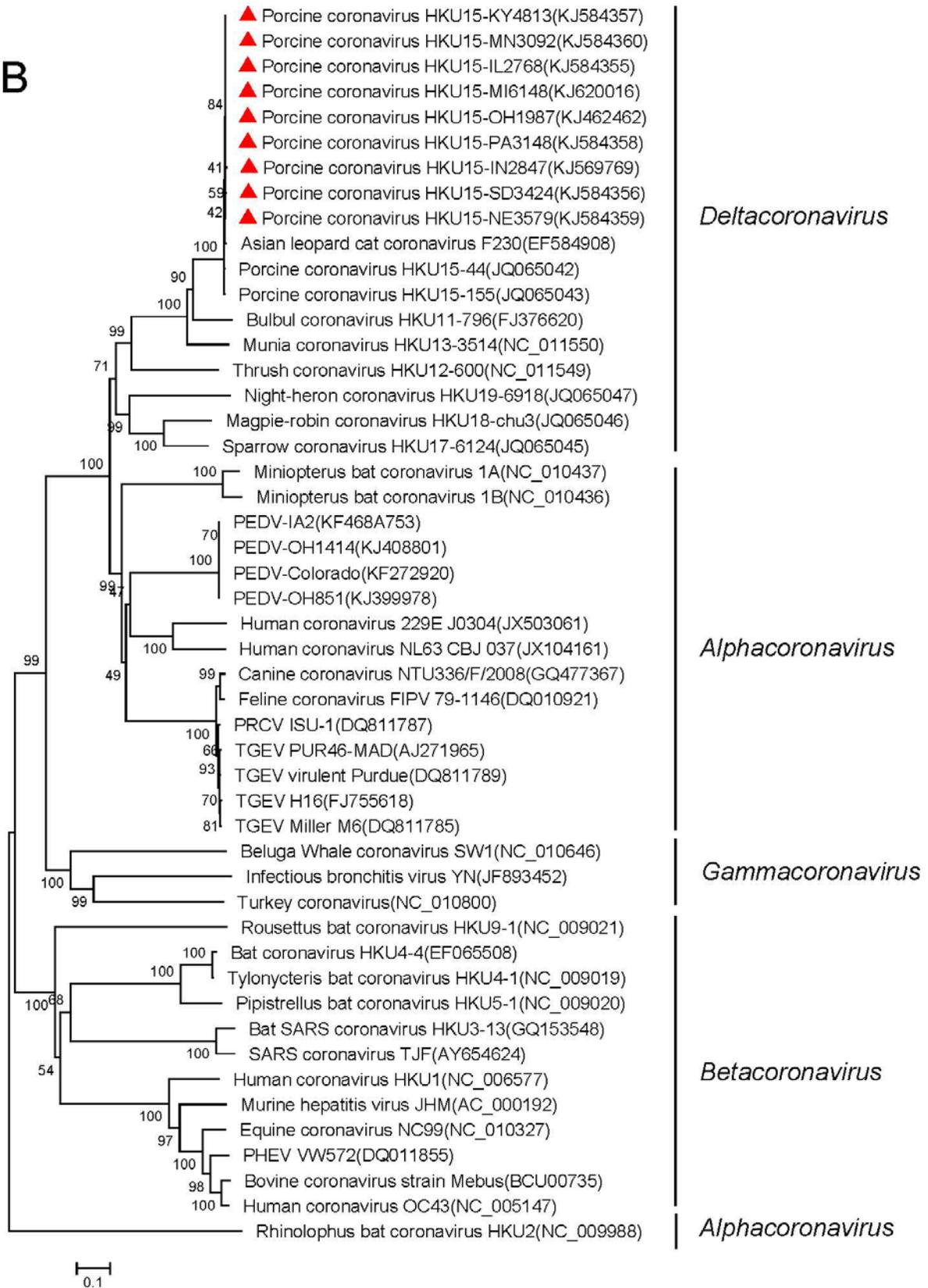
The parents of the girl (family A) did not report any stay or travel in an area to which urinary schistosomiasis was endemic; they reported summer holidays only in Mallorca in the Balearic Islands (Spain) and Corsica. However, her father reported that since 2012, he had experienced gross hematuria that had been evaluated by standard urologic investigations but not by cystoscopy; no etiology was determined. Parasitologic urinalysis in our hospital department showed numerous viable *S. haematobium* eggs in the father's urine.

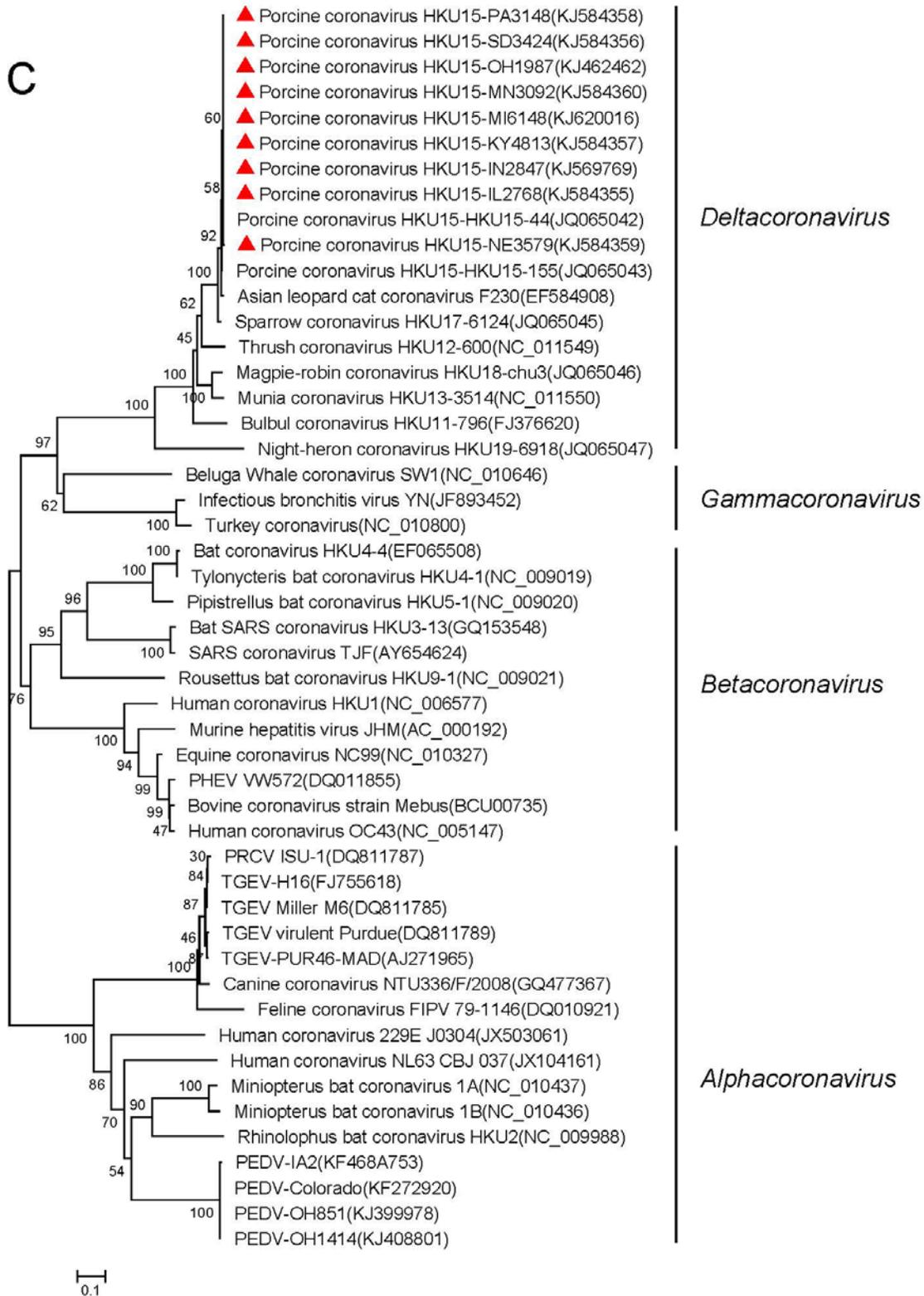
The parents of the index case-patient also reported that an 8-year-old boy in a friend's family (family B), who shared summer vacations with

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Technical Appendix



B



Technical Appendix Figure. Phylogenetic tree constructed on the basis of the whole genome sequences (A), spike protein (B) and nucleocapsid protein (C) of 4 genera of coronaviruses (*Alphacoronavirus*, *Betacoronavirus*, *Gammacoronavirus*, and *Deltacoronavirus*), including the 9 US strains of porcine coronavirus HKU15 (indicated by red triangle). The dendrogram was constructed by using the neighbor-joining method in the MEGA software package, version 6.05 (<http://www.megasoftware.net>). Bootstrap resampling (1,000 replications) was performed, and bootstrap values are indicated for each node. Reference sequences obtained from GenBank are indicated by strain name and accession number. Scale bar represents 0.5 nt (A) or 0.1 aa (B, C) substitutions per site. PEDV, porcine epidemic diarrhea virus; PHEV, porcine hemagglutinating encephalomyelitis virus; PRCV, porcine respiratory coronavirus; TGEV, transmissible gastroenteritis coronavirus; SARS, severe acute respiratory syndrome.